

ISOLATION, IDENTIFICATION AND CHARACTERIZATION OF NON-SACCHAROMYCES AND SACCHAROMYCES YEASTS IN TRADITIONAL WINES MADE FROM FIVE DIFFERENT NATIONAL GRAPES IN TURKEY

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ABSTRACT

ISOLATION, IDENTIFICATION AND CHARACTERIZATION OF NON-SACCHAROMYCES AND SACCHAROMYCES YEASTS IN TRADITIONAL WINES MADE FROM FIVE DIFFERENT NATIONAL GRAPES IN TURKEY

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Kalecik Karası, Öküzgözü, Dimrit and Boğazkere are local red grapes in Ankara, Elazığ, Cappadocia and Elazığ, respectively. Moreover, Emir is a local white grape grown in Cappadocia. Yeast populations are different in various types of grapes due to the unique properties of these grapes. In this study, 104 non-*Saccharomyces* and 293 *Saccharomyces* yeasts were isolated at different maceration and fermentation times from the grape's musts and wines made of these grapes. All 104 non-*Saccharomyces* isolates were sequenced. 29 (among 77 sequenced isolates) out of 293 total *Saccharomyces* yeasts were selected as those with high technological properties. Among all 181 (104 non-*Saccharomyces* and 77 *Saccharomyces*) yeast isolates, 16 different species of yeasts were identified by DNA sequencing of internal transcribed spacer (ITS) region and/or D1/D2 domain of large subunit ribosomal DNA as: *Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae*, *Hanseniaspora uvarum*, *Rhodotorula mucilaginosa*, *Wickerhamomyces anomalus*, *Metschnikowia aff. fructicola*, *Metschnikowia fructicola*, *Lachancea thermotolerans*, *Solicoccozyma aeria*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima*, *Metschnikowia*

sinensis, *Metschnikowia chrysoperlae*, *Starmerella bacillaris*, *Saccharomyces cerevisiae* and *Saccharomyces cf. cerevisiae/paradoxus*. Selected non-*Saccharomyces* isolates were tested for some phenotypic properties such as alcohol tolerance, SO₂ tolerance and H₂S production. In addition, selected *Lachancea thermotolerans* and *Saccharomyces cerevisiae* were used as starter cultures for sequential inoculation to produce Emir wine. Moreover, *Hanseniaspora uvarum* and *Saccharomyces cerevisiae* were added as a single starter culture separately for wine making from Kalecik Karası grapes. Volatile compounds of these wines were analyzed by gas chromatography-mass spectrometry (GS-MS). According to flavour profiles and amounts of volatile compounds, Emir wines with sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains were found successful.

Keywords: Non-*Saccharomyces*, *Saccharomyces*, wines starter cultures, identification, aroma compounds

ÖZ

TÜRKİYE'DEKİ BEŞ FARKLI YEREL ÜZÜMDEN YAPILAN GELENEKSEL ŞARAPLARDAN NON-SACCHAROMYCES VE *SACCHAROMYCES* MAYALARIN İZOLE EDİLMESİ, TANIMLANMASI VE KARAKTERİZASYONU

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Kalecik Karası, Öküzgözü, Dimrit ve Boğazkere sırasıyla Ankara, Elazığ, Kapadokya ve Elazığ'ın yerel kırmızı üzümeleridir. Buna ek olarak, Emir, Kapadokya'da yetişen yerel beyaz üzümdür. Bu üzümler kendilerine özgü özelliklere sahip oldukları için bu üzümlerdeki maya popülasyonu birbirlerinden farklıdır. Bu çalışmada, bu üzümlerin şıralarından ve şaraplarından değişik maserasyon ve fermantasyon zamanlarında 104 non-*Saccharomyces* ve 293 *Saccharomyces* maya izole edilmiştir. 104 non-*Saccharomyces* mayanın hepsi sekanslanmıştır. Toplam 293 *Saccharomyces* mayadan (77 sekanslanmış izolat arasından) 29 tanesi yüksek teknolojik özellik taşıyanlar olarak seçilmiştir. Tamamı 181 maya izolatı arasından (104 non-*Saccharomyces* ve 77 *Saccharomyces*) ITS bölgesine ve ribosomal DNA'nın büyük alt ünitesindeki D1/D2 domainindeki DNA dizilimine göre 16 farklı maya türü tanımlanmıştır: *Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae*, *Hanseniaspora uvarum*, *Rhodotorula mucilaginosa*, *Wickerhamomyces anomalus*, *Metschnikowia aff. fructicola*, *Metschnikowia fructicola*, *Lachancea thermotolerans*, *Solicoccozyma aeria*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima*, *Metschnikowia*

sinensis, *Metschnikowia chrysoperlae*, *Starmerella bacillaris*, *Saccharomyces cerevisiae* ve *Saccharomyces cf. cerevisiae/paradoxus*. Seçilen non-*Saccharomyces* izolatların alkol direnci, SO₂ direnci ve H₂S üretimi gibi bazı fenotipik özelliklerini test edilmiştir. Buna ek olarak seçilen *Lachancea thermotolerans* ve *Saccharomyces cerevisiae*, Emir şarabına starter kültür olarak sıralı inoküle edilmiştir. Bununla beraber, *Hanseniaspora uvarum* ve *Saccharomyces cerevisiae*, Kalecik Karası üzümünden şarap yapımı için tek starter kültür olarak kullanılmıştır. Bu şarapların aroma bileşikleri gaz kromatografisi-kütle spektrometresi ile analiz edilmiştir. Aroma profillerine ve miktarlarına göre *Lachancea thermotolerans* ve *Saccharomyces cerevisiae* suşlarımızla sıralı aşılama yöntemiyle yapılan Emir şarapları başarılı bulunmuştur.

Anahtar Kelimeler: Non-*Saccharomyces*, *Saccharomyces*, şarap starter kültürü, tanılama, aroma bileşikleri

To My Beloved Family...

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LIST OF ABBREVIATIONS

NS: Non-*Saccharomyces*

S: *Saccharomyces*

YPG: Yeast extract-peptone-glucose

ESA: Ethanol sulfate agar

v/v: Volume per volume

CM: Cold maceration

NM: Normal maceration

E-LT-SC: Wine made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Emir grapes

E-SC: Wine made by inoculation of our *Saccharomyces cerevisiae* strain in Emir grapes

E-WA-SC: Wine made by sequential inoculation of our *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* strains in Emir grapes

E-A: Traditional wine made from emir grapes (A-Parallel)

E-B: Traditional wine made from emir grapes (B-Parallel)

E-Laff: Wine made by inoculation of commercial *Saccharomyces cerevisiae* strain (Laffort) in Kalecik Karası grapes

K-13: Wine made by inoculation of our *Saccharomyces cerevisiae* strain in Kalecik Karası grapes

K-HU: Wine made by inoculation of our *Hanseniaspora uvarum* strain in Kalecik Karası grapes

K-SC: Wine made by inoculation of commercial *Saccharomyces cerevisiae* strain (Chr. Hansen) in Kalecik Karası grapes

K-A: Traditional wine made from Kalecik Karası grapes

O-LT-SC: Wine made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes

O-HO-SC: Wine made by sequential inoculation of our *Hanseniaspora opuntiae* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes

O-HG-SC: Wine made by sequential inoculation of our *Hanseniaspora guilliermondii* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes

O-SC: Wine made by inoculation of our *Saccharomyces cerevisiae* strain in Öküzgözü grapes

O-A: Traditional wine made from Öküzgözü grapes

O-CH: Wine made by inoculation of commercial *Saccharomyces cerevisiae* strain (Chr. Hansen) in Öküzgözü grapes

CHAPTER 1

INTRODUCTION

1.1. Grape Varieties

1.1.1. Grape Varieties in The World

Vineyards include 75% of the world area. According to the research for 44 countries at 2000, these areas are much more than 65,000 hectares for 10 main diversity.

There are three type of grapes with respect to usage area: table grapes (fresh grapes), dried grapes and wine grapes (Figure 1.1). According to this usage area, vineyards are taken forms. Production of grape wine becomes prominent in France and Spain while production of table and wine grapes comes to the forefront in Italy. In addition, production of three types grapes (fresh, dried and wine) come to the forefront in China and USA whereas production of table and dried grape becomes prominent in Turkey (Karabat, 2014).

In the world, there is 10000 known grape diversity and 13 of them encompass more than 33% of all vineyard field (Table 1.1).

According to the international catalogue of *Vitis* genus, there are 21045 names, and this includes 12250 for *Vitis vinifera*. However, this covers excessively synonyms and homonyms (Maul & Topfer, 2015). Indeed, the number of *Vitis vinifera* species is predicted as 6000 in the world (Lacombe, 2012).

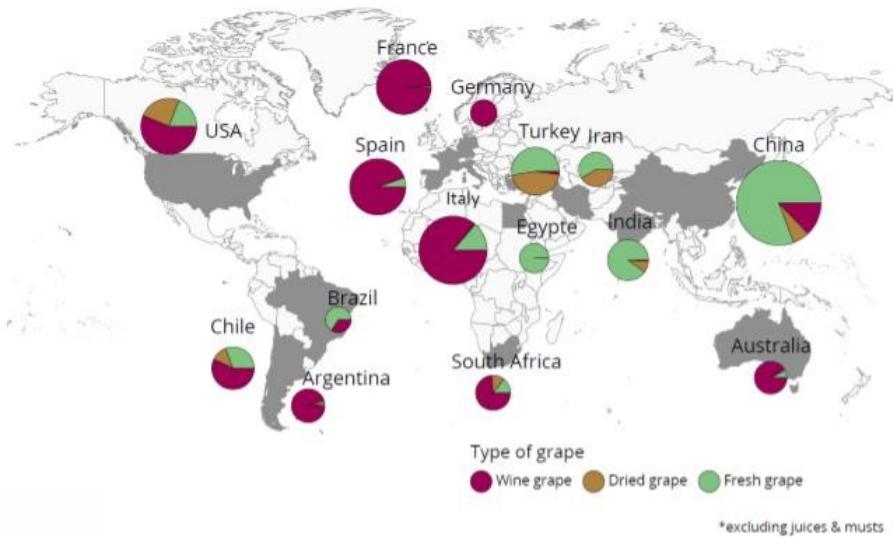


Figure 1.1. Grape Varieties in The World ("International Organisation of Vine and Wine", 2017)

Table 1.1. Distribution of Grape Variety in The World ("International Organisation of Vine and Wine", 2017)

Grape Variety	Grape Colour	Usage Area	Field (Hectares)
Kyoho	Black	Table	365000
Cabernet Sauvignon	Black	Wine	341000
Sultana	White	Table, Drying and Wine	273000
Merlot	Black	Wine	266000
Tempranillo	Black	Wine	231000
Airen	White	Wine, Brandy	218000
Chardonnay	White	Wine	210000
Syrah	Black	Wine	190000
Red Globe	Black	Table	159000
Garnacha Tinta/ Grenache Noir	Black	Wine	163000
Sauvignon Blanc	White	Wine	123000
Pinot Noir / Blauer Burgunder	Black	Wine	112000
Trebbiano Toscano / Ugni Blanc	White	Wine, Brandy	111000

1.1.1.1. Kyoho

Kyoho is main grape in China. China has more than 90% vines all over the world. Moreover, it has been produced in Japan since 1994 (Morinaga, 2001).

Kyoho has big purple grape including high sugar concentration. This brix value is between 18 and 20. It has nice foxy aroma. The shelf life of this grape is very short and the property of resistance of disease is medium level. It gives 12-15 tons per hectare product (Morinaga, 2001).

1.1.1.2. Cabernet Sauvignon

Cabernet Sauvignon comprises of Cabernet Franc and Sauvignon Blanc. The origin is Bordeaux. It has small grape with long ripeness period. It gives 2-14 tons per hectare product (Galet, 2015).

Wines with these grapes typically have violet and bell pepper aroma. They have high tannin content. Because of the high tannin content and color, these grapes are blended with other grapes such as Merlot, Cabernet Franc, Malbec and/or Petit Verdot. In addition, their wines are suitable for ageing. During ageing process, vegetal aroma has lost gradationally and develop more complex aroma.

Cabernet Sauvignon is the most cultivated grapes all over the world. It covers 341,000 hectares in the world (4% of the vineyards in the world). It is also mentioned an international variety. It produces in France, Argentina, United States, Italy, Spain, China, Australia, Chile and South Africa ("International Organisation of Vine and Wine", 2017).

1.1.1.3. Sultanina

The origin of Sultanina is Afghanistan. It is white seedless grape. Because of small grape, it is suitable for raisin. Moreover, it is used for table grape and wine grape especially in Turkey and The United States of America. Moreover, it can also be used for production of raki.

The production is 273,000 hectares approximately. It produces mainly in the Middle East such as Turkey, Pakistan, Iraq, Afghanistan and Iran, and Central Asia such as Tajikistan, Turkmenistan and Uzbekistan ("International Organisation of Vine and Wine", 2017).

1.1.1.4. Merlot

The origin of Merlot is Bordeaux, France. It has average ripeness time with early flowering. Moreover, it has small berries. Its wines have deep colour, round and structural properties. Because high tannin properties, they can be blended with other grapes.

It can be produced in 37 counties, and the production area is 266,000 hectares. This means 3% of vineyard in the world.

1.1.1.5. Tempranillo

The origin of Tempranillo is Spain. The production area is 231,000 hectares. It gives 2-10 tons per hectare product (Hidalgo, 2002).

Its wine has deep colour, high alcohol concentration and low acidity properties (Galet, 1990).

1.1.1.6. Airen

The origin of Airen is Spain. It is a white grape. Its wine has light color, distinctively fragrance and low acidity properties. It can be blended with other grapes frequently. Moreover, it can be used for making a red wine and spirit. It gives 5-20 tons per hectare product.

1.1.1.7. Chardonnay

The origin of Chardonnay is Burgundy. It is a white wine. It comprises of Gouais Blanc and Pinot (Bowers et al., 1999).

It has small berries and its wine has outstanding flavour. For example, its dry and sparkling wine with adding spirit, it gives dried fruit, walnuts or butter flavours. In addition, it is suitable for ageing process. It gives between 3 and 10 tons per hectare product (Galet, 2015).

It is most international white grapes all over the world. The production area is 210,000 hectares. It produces in Italy, France, Spain, Chile, The United States, and Australia.

1.1.1.8. Syrah

The origin of Syrah is France. It comprises of Mondeuse Blanche and Dureza (Meredith & Boursiquot, 2008).

It has a short ripening time and a late harvesting period. Size of berries changes between small and medium. Its wine has deep colour and high tannin content. It gives between 3 and 8 tons per hectare product.

The production area is 190,000 hectares. It is produced in The United States, Australia, Chile, South Africa and Argentina.

1.1.1.9. Red Globe

Red globe is the second produced table grape all over the world. 91% of total production area is China. The rest of producers are in Spain, Italy, Turkey, Portugal, Chile, The United States, Argentina, and South Africa. It gives between 8 and 30 tons per hectare product.

It can be used for addition of salads (Galet, 2015).

1.1.1.10. Garnacha Tinta (Grenache Noir)

The origin of this grape is Spain and its producers are mainly in Spain and France. It gives 2-8 tons per hectare product.

Its wine has deep colour and high alcohol concentration. Therefore, it can be blended with other grapes.

1.1.1.11. Sauvignon Blanc

The origin of Sauvignon Blanc is France. It is an international white variety. Total production area is 123,000 hectares and 20,500 of them is in New Zealand. It gives 5-10 tons per hectare product.

Its wine has thiol aroma such as exotic fruit, blackcurrant and boxwood. It can be used for producing young white wine. In addition, its dessert wine has high quality.

1.1.1.12. Pinot Noir (Blauer Burgunder)

The origin of Pinot Noir is Burgundy. It has small black berries. It has high sugar content and partially acidic properties. In addition, its skin has high polyphenol concentration. Its wine has a light colour and is suitable for ageing in barrels. Moreover, it can be blended with Chardonnay for making sparkling wines.

It produces in Romania, Spain, Italy, Germany, Hungary, Switzerland, Argentina, New Zealand, Chile, Australia, South Africa and the United States. Therefore, it is the fourth most produced grape type.

1.1.1.13. Trebbiano Toscano (Ugni Blanc)

The origin of this grape is Italy. Its producers are in Italy, France, and Portugal.

It is a white grape. Its wine has low alcohol content and high acidity properties. It can be blended with other grape varieties because of balancing neutral aroma. Therefore, it is used for making dry wines, liqueurs and sparkling wines. In addition, this grape is one of the main sources of ice wine (D'Agata, 2014).

According to distribution of grape varieties, the following figures are generated with most 5 grape producer countries (Figure 1.2-1.6):

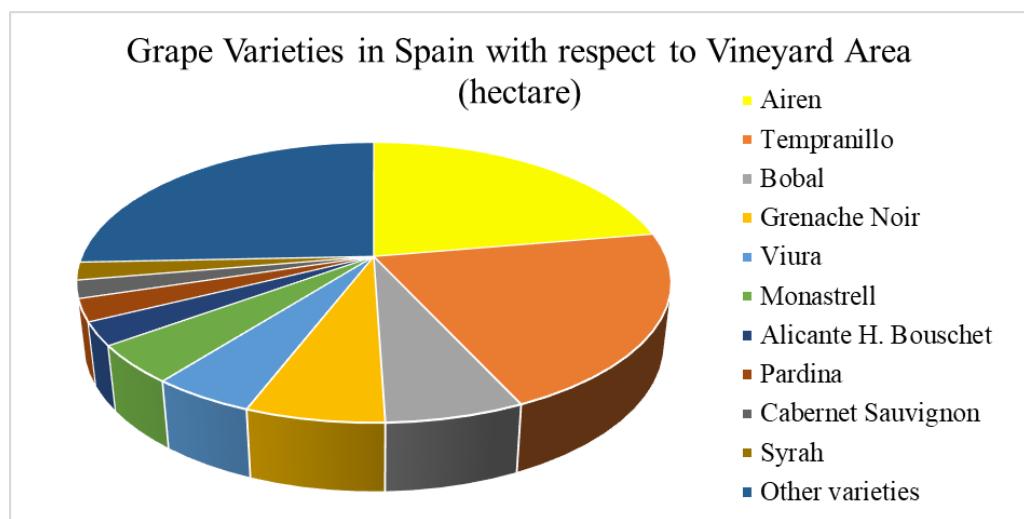


Figure 1.2. Grape Varieties in Spain ("International Organisation of Vine and Wine", 2017)

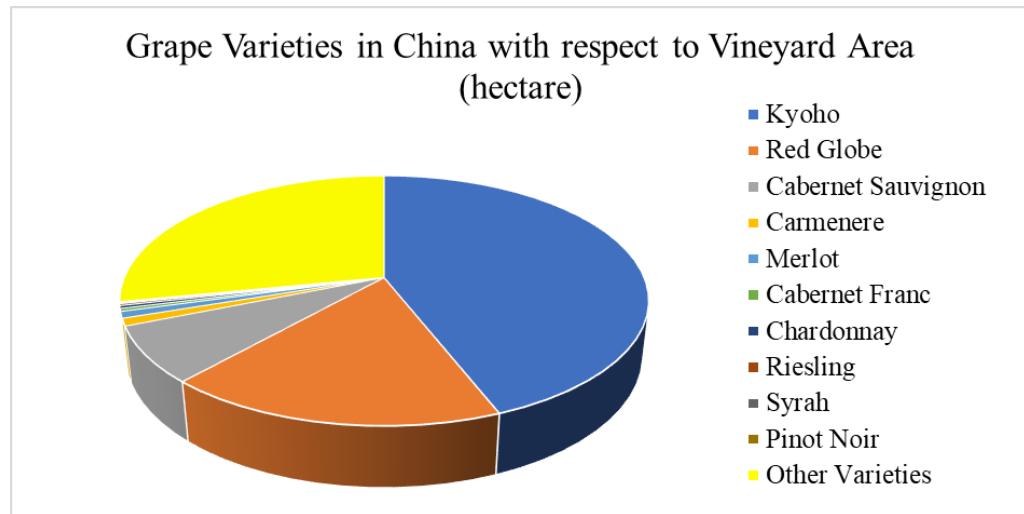


Figure 1.3. Grape Varieties in China ("International Organisation of Vine and Wine", 2017)

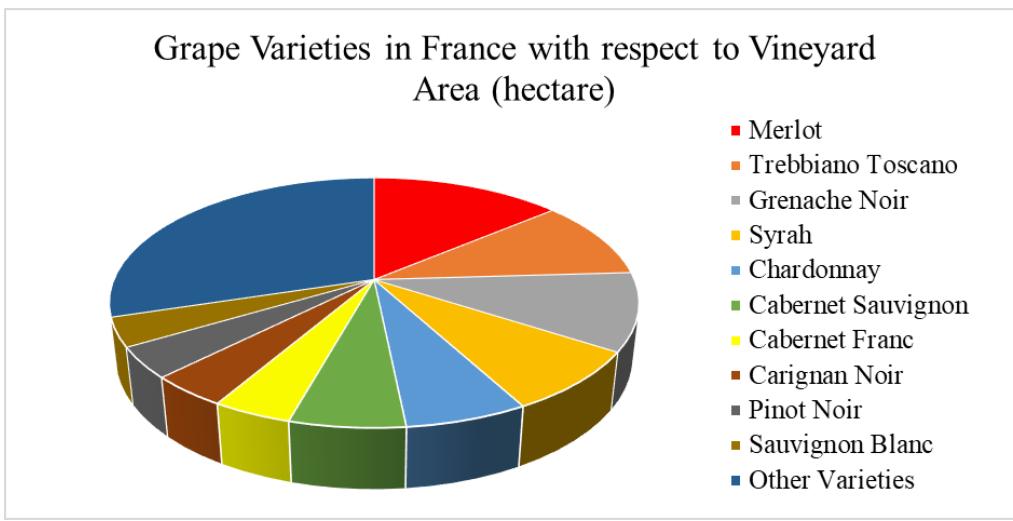


Figure 1.4. Grape Varieties in France ("International Organisation of Vine and Wine", 2017)

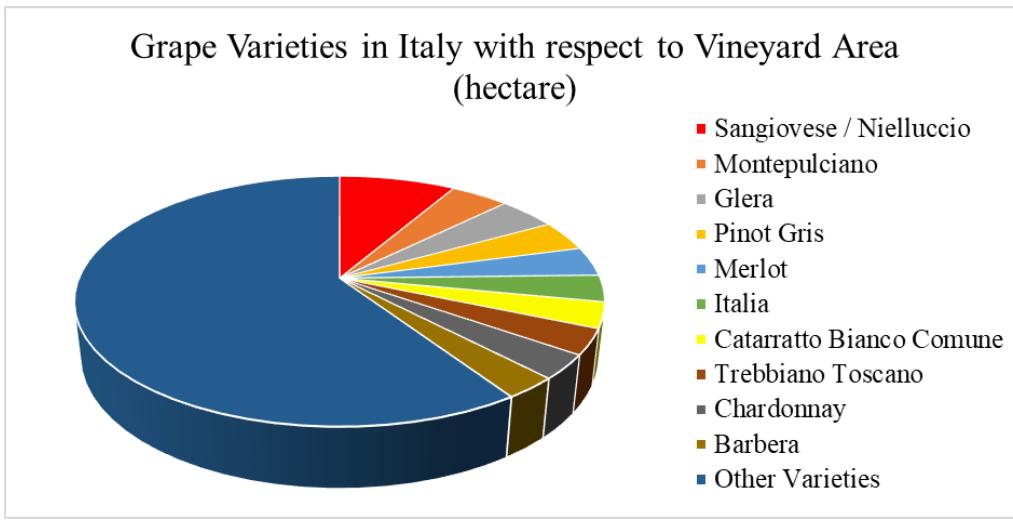
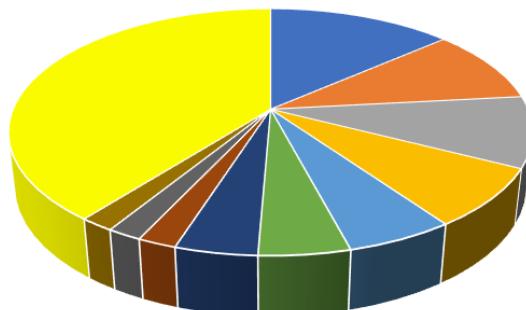


Figure 1.5. Grape Varieties in Italy ("International Organisation of Vine and Wine", 2017)

Grape Varieties in United States with respect to
Vineyard Area (hectare)



- Sultanina
- Chardonnay
- Cabernet Sauvignon
- Concord
- Pinot Noir
- Merlot
- Zinfandel / Primitivo
- Syrah
- Pinot Gris
- Colombard
- Other varieties

Figure 1.6. Grape Varieties in United States ("International Organisation of Vine and Wine", 2017)

1.1.2. Grape Varieties in Turkey

According to statistic information in 2011, vineyard area is 7,098,755 hectares in the world and 472,545 hectares in Turkey. The capacity of fresh grape is 4,296,351 tons in Turkey while this number is 69,093,293 tons all over the world (FAO, 2012).

According to production area, Turkey is the fifth major producing country (Spain, France, Italy, China, Turkey and USA). In addition, Turkey is the sixth country with respect to the amount of grape production (China, Italy, USA, France, Spain, and Turkey).

Grapes in Turkey separate three parts: fresh (table), dried and wine grapes. The percentages of total grape production are 52.8% for fresh grape, 36.4% for dried grape, and 10.8% of must-wine grape (Karabat, 2014).

There are more than 1200-1500 grape varieties. 600-800 of them are genetically different from each other in Turkey. The number of produced wine grape varieties is around 30: Adakarasi, Alicante Bouchet, Boğazkere, Bornova Misketi, Cabernet Franc, Cabernet Sauvignon, Carignan, Chardonnay, Cinsault, Çalkarasi, Çavuş,

Dimrit, Emir, Gamay, Grenache, Kalecik Karası, Karalahana, Kuntra (Karasakız), Malbec, Merlot, Narince, Öküzgözü, Papazkarası, Petit Verdot, Pinot Noir, Riesling, Sangiovese, Sauvignon Blanc, Semillon, Shiraz, Sultaniye, Tempranillo, Vasilaki, Viognier. 15 of them are Turkey's native grapes: Adakarasi, Boğazkere, Bornova Misketi, Çalkarasi, Çavuş, Dimrit, Emir, Kalecik Karası, Karalahana, Kuntra (Karasakiz), Narince, Öküzgözü, Papazkarası, Sultaniye, Vasilaki. ("Wines of Turkey", 2019). These wine grape varieties and their production areas are shown in Figure 1.7.



Figure 1.7. Grape Varieties in Turkey ("Balkan Pazar", n.d.)

1.1.2.1. Sultaniye

It grows in Denizli and Manisa. It is used for table grape and wine-making. Its wine has light colour and fresh fruity aroma. In addition, it can be made dry or semi-dry wine.

The aroma profile is asparagus, pineapple, mango, pear, floral, lemon, hay, golden and green apples.

1.1.2.2. Öküzgözü

The origin is Elazığ. It also grows in Cappadocia, Ankara and Uşak. It is medium bodied, producing round and fruity wines with tannin and high acidity property. The alcohol range changes between 12.5% to 13.5%. Its wine has floral and fruity aroma.

Therefore, its wine can be drunk easily and also suitable for ageing due to high acidity property. The color of Its wine is light red. It can be blended with Boğazkere grapes.

The aroma profile is raspberry, mint, sour cherry, dark cherry, pomegranate, chocolate, ripe plum, eucalyptus, cardamom.

1.1.2.3. Boğazkere

The origin is Diyarbakır. It also grows in Elazığ, Ankara, Uşak, Manisa, Denizli, and Elmalı. Its wine is full bodied, dense tannin with complex aroma profiles and medium acidity. Its wine reaches balance with ageing.

The aroma profile is raspberry, black cherry, blackberry, pepper, eucalyptus, leather, tobacco, dark chocolate, pine forest, clove, liquor ice.

1.1.2.4. Kalecik Karası

The origin is Ankara. It also grows in Denizli, Manisa, Uşak, Elmalı, and Cappadocia. Its wine has dried red rose colour and a sugar candy flavour. It also has moderate bodies on the palate with low tannins and crunch acidity properties. Its alcohol level is 14% (v/v).

The aroma profile is cherry, red berries, cotton candy, raspberry and strawberry.

1.1.2.5. Narince

The origin is Tokat. It also grows in Cappadocia. It is consumed both table and wine grape. It is suitable for dry or semi-dry wines. Its wine has yellow colour and citrus and fruity flavour. In addition, it has round, moderate to high body properties with balanced acidity. It is suitable for ageing with oak. This ageing process gives a complex flavour in this wine.

The aroma profile is grapefruit, white pineapple, orange, floral, acacia, ripe green apple, quince, walnut, plumeria, lime, basil, fruit blossom.

1.1.2.6. Emir

The origin is Cappadocia. It is used for sparkling and dry wines. Its wine has yellow colour with lively crunch flavour. It has light to moderate bodies with high fragile acidity. It is not suitable for ageing with oak. In addition, it is not designed to malolactic fermentation. It has been drunk within 1 or 2 years.

The aroma profile is pineapple, apple, lemon, kiwi, white rose, blood orange.

1.1.2.7. Çalkarası

The origin is Denizli. It is used for rosé wine. Its wine has lightly fruit red colour with high acidity. It has lively and balanced aroma. The alcohol level changes between 12% and 13.5% (v/v).

The aroma profile is strawberry, peaches, ripe white fruits, fresh red fruits.

1.1.2.8. Bornova Misketi

The origin is İzmir. It also grows in Manisa. Its wine has muscat flavour. It has light golden colour with lively aroma. It can be drunk easily. In addition, it can be suitable for dry and sweet wine.

The aroma profile is flowers, tropical fruits, citrus, bay leaves, thyme (“Wine of Turkey”, 2019).

The following tables and figure show distribution of the mentioned grape varieties with production capacity and national grape types with their origin and producers (Table 1.2 & 1.3 and Figure 1.8):

Table 1.2. Distribution of Grape Variety in Turkey

Grape Variety	Grape Colour	Usage Area	Output (Tons)
Sultaniye	White	Table and Wine	14000
Shiraz (Syrah)	Black	Wine	11932
Öküzgözü	Black	Wine	11830
Boğazkere	Black	Wine	8850
Kalecik Karası	Black	Wine	6885
Narince	White	Table and Wine	6150
Emir	White	Wine	5500
Çalkarası	Black	Wine	5000
Cabernet Sauvignon	Black	Wine	3125
Merlot	Black	Wine	2840
Sauvignon Blanc	White	Wine	1165
Chardonnay	White	Wine	1135
Bornova Misketi	White	Wine	910

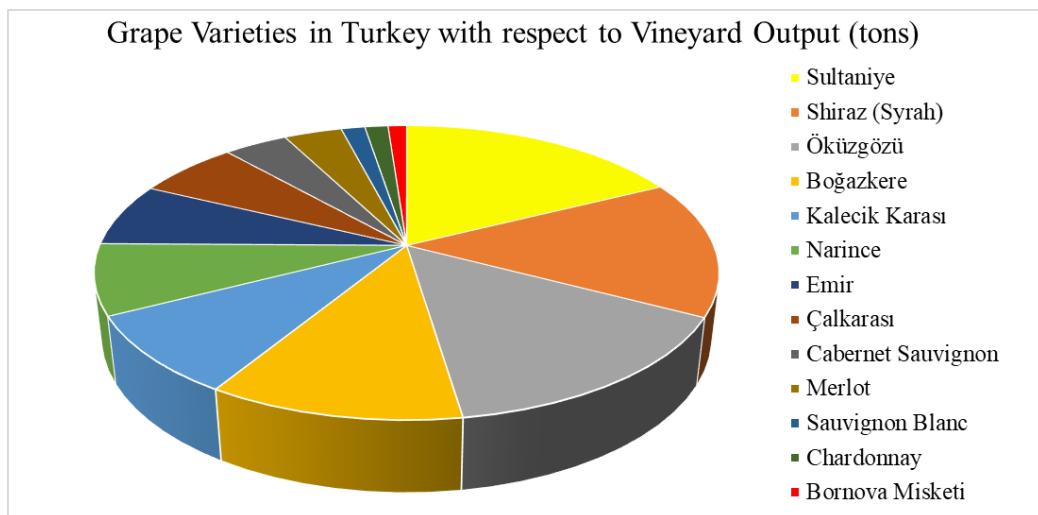


Figure 1.8. Grape Varieties in Turkey

Table 1.3. Origin of Grapes and Winemakers in Turkey

<i>Grape Type</i>	<i>Origin</i>	<i>Winemaker*</i>
Adakarası	Avşa	Büyülbağ
Boğazkere	Diyarbakır	Kavaklıdere, Suvla, Kayra, Doluca, Sevilen, Likya, Turasan, Buzbağ
Bornova Misketi	İzmir	Sevilen, Corvus Vineyards, Kavaklıdere, Nif
Çalkarası	Denizli	Sava, Kavaklıdere
Çavuş	Bozcaada	Corvus Vineyards, Talay Vinification
Dimrit	Kapadokya	Turasan, İllegüp
Emir	Kapadokya	Turasan, Kavaklıdere, Vinolus, Doluca, Kocabağ, Gilamada, Corvus, Yazgan
Kalecik Karası	Ankara	Kavaklıdere, Diren, Vinkara, Kayra, Doluca, Sevilen
Karalahana	Çanakkale	Corvus, Çamlıbağ
Karasakız (Kuntra)	Çanakkale	Suvla, Kocadere, Paşaeli
Narince	Tokat	Kavaklıdere, Doluca, Buzbağ, Kayra, Diren, Vinkara, Sava, Vinolus, Sevilen
Öküzgözü	Elazığ	Kavaklıdere, Diren, Suvla, Doluca, Vinkara, Kayra, Vinolus, Kuzeybağ, Corvus, Kocabağ, Buzbağ
Papazkarası	Tekirdağ	Melen, Chamlıja
Sultaniye	Denizli	Kavaklıdere, Doluca, Sevilen, Yazgan, Nif
Vasilaki	Bozcaada	Talay, Çamlıbağ, Corvus

*Winery A.Ş.

1.2. Wine Yeasts

1.2.1. Definition of Yeasts

Yeast is a eukaryotic microorganism categorized as fungi in the kingdom and described as single cell fungi which predominantly reproduced by budding or fission. According to type of reproduction, it is classified as ascomycetes (budding), basidiomycetes (fission), and imperfect fungi (Deuteromycetes).

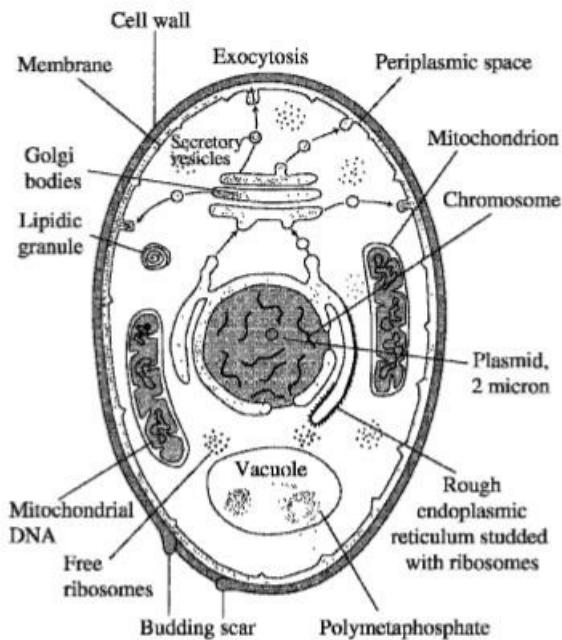


Figure 1.9. Yeast Cell (Gaillardin & Heslot, 1987)

Yeast cell includes cell envelopes, a cytoplasm with different organelles, and a nucleus encompassed by a membrane. Cellular envelope of yeast has a critical role. This is to provide alcohol fermentation successfully and release some components which contribute composition of final wine (Don, 2006).

1.2.2. Taxonomy of Wine Yeasts

Taxonomy of yeast contains classification and identification. Classification refers to indicate similarity or difference with ancestors. Identification demonstrates unknown microorganisms and put it to a group which has similar properties.

Taxonomy categorizes the microorganisms with respect to morphological, physiological, genetic and biochemical properties. The phenotypic properties of yeasts base on spore formation, size and shape of cell, ethnical characters, assimilation and fermentation of sugars and nitrogen sources, cycloheximide resistance (Don, 2006).

	F1 D-Glucose fermentation	F2 D-Galactose fermentation	F3 Maltose fermentation	F4 M ₆ α-D-glucoside fermentation	F5 Sucrose fermentation	F6 α-D-Trehalose fermentation	F7 Maltotriose fermentation	F8 Lactose Fermentation	F9 Cellobiose fermentation	F10 Melibiose fermentation	F11 Raffinose fermentation	F12 Inulin fermentation	F13 Starch fermentation	
<i>Candida stellata</i>	+	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Candida vini</i>	v	-	-	-	-	-	-	-	-	-	-	-	-	-
<i>Candida famata</i>	v v v - v v v	-	v v v -	-	+ + v v v +	-	-	v v v v v v v v	-	v v v v v v v v	-	v v - v	-	-
<i>Dekkera anomala</i>	+ v v v + v -	v v v v -	v v v -	-	v v v v v v v v	-	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	-
<i>Dekkera bruxellensis</i>	v v v + + + -	-	v v v -	-	v v v v v v v v	-	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	v v v v v v v v	-
<i>Hanseniaspora uvarum</i>	+ - - - -	-	v - - -	-	-	-	-	-	-	-	-	-	-	v -
<i>Metschnikowia pulcherrima</i>	+ v - - -	-	-	-	+ + v v v v	-	-	+ + + + + + +	-	-	-	-	-	+ - + - v + + +
<i>Pichia anomala</i>	+ v v v + v -	-	v - v -	-	+ v - v v	-	-	+ + v + v + +	-	v v - + v v v v	-	v v - + v v v v	-	-
<i>Pichia fermentans</i>	+ - - - -	-	-	-	+ - v - +	-	-	-	-	-	-	-	-	-
<i>Pichia membranefaciens</i>	v - - - -	-	-	-	+ - v v - v	-	-	-	-	-	-	-	-	v - v - v
* <i>Saccharomyces cerevisiae</i>	+ v v v v v v v	-	v v v - v	-	+ v - - -	-	-	v v v v v v v v	-	v v v v v v v v	-	v v - v v - - - v v	-	-
<i>Saccharomyces ludwigii</i>	+ - - + -	-	-	-	+ -	-	-	+ + + + + +	-	v -	-	v -	-	-
<i>Kluyveromyces thermolerens</i>	+ v v v + v +	-	-	-	v + v + -	-	-	+ + + + - - -	-	+ v + - + v + v v	-	+ v v -	-	-
<i>Schizosaccharomyces pombe</i>	+ v v v + - v	-	-	-	v v v v	-	-	+ + v v - - v	-	+ - v v -	-	+ - v v -	-	-
<i>Zygosaccharomyces bailii</i>	+ - - v v -	-	-	-	+ v v - -	-	-	v v - - -	-	v v - - v v v v v	-	v v - - v v v v v	-	-
	C30 Galactose growth	C31 myo-Inositol growth	C32 D-Glucano-1,5-lactone growth	C33 2-Keto-D-glucosate growth	C34 5-Keto-D-glucosate growth	C35 D-Glucosamine growth		C1 D-Glucose growth	C2 D-Galactose growth	C3 D-α-D-Glucoside growth	C4 D-Ribose growth	C5 D-Glucosamine growth	C6 D-Xylose growth	
	C38 D-Lactate growth	C39 Succinate growth	C40 Citrate growth	C41 Ethanol growth		N1 Nitrate growth		C10 Sucrose growth	C11 Malic acid growth	C12 D-Trehalose growth	C13 Me α-D-Glucoside growth	C14 Cellobiose growth	C15 Salicin growth	C16 Arabinose growth
	N2 Nitrite growth	N3 Dihydronicotinic acid growth	N4 L-Lysine growth	N5 D-Serine growth	N6 Creatine growth	N7 Creatinine growth	N8 Glucosamine growth	C17 Melibiose growth	C18 Lactose growth	C19 Raffinose growth	C20 Melibiose growth	C21 Inulin growth	C22 Starch growth	C23 Glycerol growth
	N9 Growth W/O vitamins	N10 Growth W/O myo-Inositol	N11 Growth W/O Patchatene	N12 Growth W/O Biotin	N13 Growth W/O Thiamin	N14 Growth W/O Biotin & Thiamin	N15 Growth W/O Pyridoxine	N16 Growth W/O Pyridoxine & Thiamin	N17 Growth at 30°C	N18 Growth W/O at 35°C	N19 Growth W/O at 37°C	N20 Growth W/O at 40°C	N21 Growth W/O at 40°C	N22 Growth W/O at 40°C
<i>Candida stellata</i>	- - - - -	- - - - -	- - - + - - -	- - - + + + - -	- - - + v v v +	- - v - - - - - v	- + v - - - - v	+ v - -	- - v	- -	- -	- -	- -	- -
<i>Candida vini</i>	- - - v -	v + +	- - + + + - -	- - + + v + v	- + v v v + +	- v + + + v + v	- + v + + + v +	v -	-	-	-	-	-	-
<i>Candida famata</i>	v - v + v	v + +	v + + + v -	v + + + + - -	v + + + v + v	v + + + + + + +	v + + + + + + +	+ v v -	v - +	-	-	-	-	-
<i>Dekkera anomala</i>	- - v - - v	v v - v	v v - v	v v + + + - -	v v + + + + -	v v + + + + + +	v v + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Dekkera bruxellensis</i>	- - v v -	v - v	v v + + -	v v + + + - -	v v + + + + -	v v + + + + + +	v v + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Hanseniaspora uvarum</i>	- - + + v	- - -	- - + + + - -	- - + + + + -	- - + + + + + +	- - v v v v v v v	- - v v v v v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Metschnikowia pulcherrima</i>	- - + + +	v + +	- - + + + - -	- - + + + + -	- - + + + + + +	- - + + + + + +	- - + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Pichia anomala</i>	- - + + v	+ + +	- - + + + - -	- - + + + + -	- - + + + + + +	- - + + + + + +	- - + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Pichia fermentans</i>	- - v - -	+ + +	- - + + + - -	- - + + + + -	- - + + + + + +	- - + + + + + +	- - + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Pichia membranefaciens</i>	- - v - -	v v - +	- - + + + - v	- - + + + + v	v + + v v v v v +	v + + v v v v v +	v + + v v v v v +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
* <i>Saccharomyces cerevisiae</i>	- - v - -	v v - v	- - + + + - -	- - + + + + -	- - + + + + + +	- - + + + + + +	- - + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Saccharomyces ludwigii</i>	- - v - -	v v - v	- - + + + - +	- - + + + + -	- - + + + + + +	- - v v v v v v v	- - v v v v v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Kluyveromyces thermolerens</i>	- - v v - v	- v - v	- - + + + - -	- - + + + + -	- - + + + + + +	- - v v v v v v v	- - v v v v v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v
<i>Schizosaccharomyces pombe</i>	- - v v - v	- - -	- - v v v - -	- - v v v - -	- - v v v v - -	- - v v v v v v v	- - v v v v v v v	+ + v v	v + +	v + +	v + +	v + +	v + +	v + +
<i>Zygosaccharomyces bailii</i>	- - v v -	- - +	- - + + + - -	- - + + + + -	- - + + + + + +	- - + + + + + +	- - + + + + + +	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v	+ v v v

+: test positive; -: test negative; v: variable result.

*With these tests they cannot be differentiated from *S. bayanus*, *S. paradoxus* and *S. pastorianus*.

Figure 1.10. Physiological Properties of Wine Yeasts (Barnett et al., 2000)

1.2.2.1. *Candida*

Candida, represents a very large group in wine, is an anamorphic genus. Its cell appears spherical, ellipsoidal or elongate under the microscope (Meyer et al., 1998).

Reproduction property of this genus is multilateral budding (Fugelsang & Edwards, 2007).

According to the species, sugars and nitrogen sources can be fermented or assimilated. For example, *Candida stellata* can use glucose, raffinose, and sucrose for fermentation and assimilation (Figure 1.10). However, *Candida pulcherrima* can ferment glucose, but assimilate galactose, sucrose, glucose, melezitose, mannitol, maltose (Meyer et al., 1998).

1.2.2.2. *Dekkera*

The other name is *Brettanomyces*. It is a spoilage microorganism for wine. There are a lot of species such as *D. anomala* (*B. anomala*), *D. bruxellensis* (*B. bruxellensis*), *B. custersianus*, *B. naardenensis*, and *B. nanus* (Smith, 1998).

Cell shape of *Brettanomyces* is boat-shaped or ogival. In addition, colour of colony in the agar is white to yellowish. This agar can be seen moist, straight, dull or glistening.

According to strains, they can ferment most sugars such as glucose, maltose, sucrose, galactose. *D. anomala* can ferment lactose with respect to strains while *D. bruxellensis* cannot use. This property can be used for identification of species.

1.2.2.3. *Hanseniaspora*

Cell shape of *Hanseniaspora* is spherical or ovoid for a young microorganism and lemon-shaped or apiculate for an older microorganism (Fugelsang & Edwards, 2007).

There are a lot of species in wine such as *H. uvarum*, *H. guillermondii*, *H. opuntiae*. For example, *H. uvarum* ferments only glucose, but it assimilates glucose, salicin and cellobiose. In addition, *Hanseniaspora* prefers to use fructose instead of glucose (Ciani and Fatichenti, 1999).

1.2.2.4. *Metschnikowia*

Reproduction property of *Metschnikowia* is multilateral budding. Generally, *Metschnikowia pulcherrima* found in the wine. It ferments glucose while galactose fermentation changes with strain to strain. In addition, it assimilates other sugars such as maltose, galactose, sucrose and lactose. Moreover, this species produces a brown or red pigment in the agar (Pallmann et al., 2001).

1.2.2.5. *Pichia*

Two species of *Pichia* found in wine: *P.anomala* and *P.membranifaciens*. *P.guilliermondii* found only in must or wine equipment (Dias et al., 2003).

The cell shape appears cylindrical, ovoid or ellipsoidal under the microscopy. In addition, reproduction property of this genus is multilateral budding. Colonies are seen cream or white, wrinkled, and dull on the agar (Fugelsang & Edwards, 2007).

P.anomala and *P. fermentans* ferment glucose while *P.membranifaciens* cannot ferment with respect to strains.

1.2.2.6. *Saccharomyces*

The cell shape appears ovoid under the microscopy. In addition, reproduction property of this genus is multilateral budding. Colonies are seen smooth on the agar.

Two species of *Saccharomyces* found in wine: *S. cerevisiae* and *S. bayanus*. *S. cerevisiae* ferments sucrose, glucose and raffinose. In addition, it assimilates glucose, maltose, sucrose, raffinose, but it doesn't utilize lactose. It can also not use five carbon sugars such as pentoses (Fugelsang & Edwards, 2007).

1.2.2.7. *Saccharomycodes*

The cell shape appears lemon-shaped under the microscopy. In addition, reproduction property of this genus is bipolar budding.

Saccharomycodes ludwigii ferments and assimilates sucrose and glucose.

1.2.2.8. *Schizosaccharomyces*

The cell shape appears ovoid and cylindrical under the microscopy. In addition, reproduction property of this genus is fission.

S. pombe ferments sucrose, maltose and glucose. In addition, it assimilates maltose, glucose, raffinose, and sucrose. However, it cannot use ethanol and nitrate.

1.2.2.9. *Zygosaccharomyces*

The cell shape of *Z. bailii* appears cylindrical and ovoid under the microscopy. In addition, reproduction property of this species is bipolar budding.

Z. bailii has alcohol tolerant property. It grows 18% (v/v) ethanol concentration (Thomas and Davenport, 1985).

1.2.3. Isolation of Yeasts

Bacteria and molds grow easily with yeast on standard medium. Therefore, some special mediums are used for isolation of yeasts. In these mediums, bacteria and molds don't grow. Generally, pH level, water activity, antibiotic or fungistatic addition are used for this purpose.

Generally, yeasts grow about 3.7 pH, but they don't resist high pH (4.5-5.0). However, *Schizosaccharomyces* only resist this pH ranges.

Usually, yeasts grow in high sugar concentration (30-50%), but bacteria don't grow in this medium. However, osmotolerant molds grow in this medium. There can be used with a rotary shaker in incubation time to inhibit molds.

A few media containing anti-microbials have been depicted which can be utilized to stifle co-occurring microorganisms. Tetracycline (50 mg/L), chloramphenicol (100-300 mg/L) or a combination of streptomycin sulfate and penicillin G (150-500 mg/L) are most used antibiotics to inhibition. Moreover, biphenyl (150 mg/L), cycloheximide (100-500 mg/L), pimaricin (5-100 mg/L) or cyclosporin A (4-10 mg/L)

can be used for inhibition of fungal (Kurtzman, 2011; Zott, Miot-Sertier, Claisse, Lonvaud-Funel, & Masneuf-Pomareda, 2008).

Selective media can also be used for specific yeast isolation. For example, the media including cycloheximide and sorbic acid (pH 4.8) can be used for *Dekkera* (Van der Walt and Van Kerken, 1961). Moreover, the medium including methanol, cycloserine and penicillin G can be utilized for inhibition of bacteria and isolation of yeast which use methanol (Van Dijken and Harder, 1974). In addition, lysine medium can be used for isolation of non-*Saccharomyces* (van der Aa Kühle & Jespersen, 1998), and ethanol sulfate agar (ESA) which containing 10 % (v/v) ethanol and SO₂ can be utilized for isolation of *Saccharomyces* (Kish, Sharf, & Margalith, 1983).

In our study, yeast isolation methods were used according to according to Zott et al., Kish et al., and van der Aa Kühle et al. as described in detail in part 2.2.2.

1.2.4. Identification of Yeasts

There are two type methods for identification of yeast: phenotypic identification and genotypic identification.

1.2.4.1. Phenotypic Identification

Phenotypic identification bases on morphological, physiological and biochemical tests. In morphological test, isolated yeasts are looked at the microscope. A yeast can be identified with respect to cell shape. For example, *Saccharomyces cerevisiae* has ovoid cell shape while *Hanseniaspora uvarum* has lemon-shape.

Physiological test bases on carbohydrate fermentation or assimilation properties (Figure 1.10), utilization of nitrogen sources, growth temperature, growth in high sugar concentration, and antibiotic resistances.

In addition, there are some automated systems such as API System, Biolog or Vitek. For example, API system kits provide 20-32 test for growing in wells. Time of reading is approximately between 18 and 24 hours. It can be identified between 42 and 63 species with respect to coding system (Kurtzman, Fell, Boekhout, & Robert, 2011).

1.2.4.2. Genotypic Identification

There are some genotypic identification methods:

- 5.8S- ITS Region Sequencing of Ribosomal DNA
- 5.8S- ITS Region Restriction Fragment Length Polymorphism (RFLP)
- D1/D2 Domain Sequencing of Ribosomal DNA
- Restriction Analysis of Mitochondrial DNA (mtDNA)
- Random Amplification of Polymorphic DNA (RAPD)
- Polymerase Chain Reaction (PCR) Amplification of Variable Regions of the Genome (Microsatellites and Minisatellites)
- Real-Time Polymerase Chain Reaction (q-PCR)
- Polymerase Chain Reaction Denaturing Gradient Gel Electrophoresis (PCR-DGGE)
- Pulsed-Field Gel Electrophoresis (PFGE)
- Amplified Fragment Length Polymorphism (AFLP)

1.2.4.2.1. 5.8S- ITS Region Sequencing of Ribosomal DNA

The ITS region contains the 5.8 rRNA gene coding and the two flanking non-coding and variable transcribed inner spacers (Figure 1.11). ITS1 and ITS2 demonstrate highly variability of interspecific size. However, their intraspecific polymorphism is low. In addition, the extremely conserved sequences of rRNA genes with ITS region enable universal primers to be used for fungi. Therefore, RFLP analysis of 5.8S ITS region is used for identification of *Saccharomyces* and non-*Saccharomyces* yeasts (Guillamón et al., 1998, Esteve-Zarzoso et al., 2009). In addition, the differences between sequencing of ITS regions in rDNA were utilized to identify species of yeast (Christoph & Thomas, 2001; Villa-Carvajal, Querol, & Belloch, 2006). The research study by Chavan et al. (2009), has demonstrated that *Saccharomyces* and non-*Saccharomyces* yeasts were identified by 5.8S ITS sequencing of rDNA.

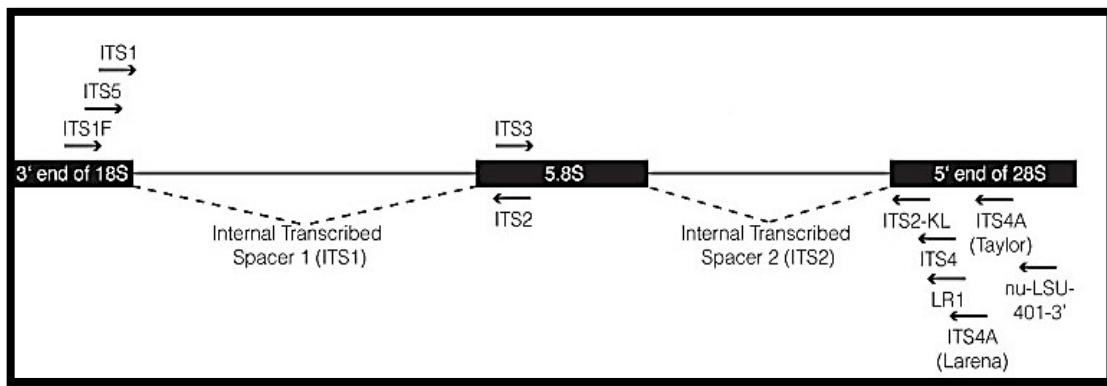


Figure 1.11. Primers of Internal Transcribed Spacer (ITS) Regions. SSU, small subunit (18 S); LSU, large subunit (25-28 S) (Nelsen, M. P., 2018)

The following sequences show the used ITS1 and ITS4 primers:

ITS1 5' TCCGTAGGTGAAACCTGC 3'

ITS4 5' TCCTCCGCTTATTGATATGC 3' (Chavan et al., 2009; White et al., 1990)

PCR amplicons with respect to ITS1-5.8S rDNA-ITS2 are different for each species.

The following table shows amplicons of some wine yeasts (Table 1.4):

Table 1.4. The ITS1-5.8 rDNA-ITS2 PCR Amplicons of Yeasts (Ghosh, 2017; Pham et al., 2011).

<i>Yeasts</i>	<i>ITS1-5.8 rDNA-ITS2 PCR Amplicons (bp)</i>
<i>Metschnikowia pulcherrima</i>	390
<i>Metschnikowia aff. pulcherrima</i>	390
<i>Pichia fermentans</i>	445
<i>Starmerella bacillaris</i>	475
<i>Dekkera bruxellensis</i>	468
<i>Dekkera anomala</i>	540
<i>Wickerhamomyces anomalus</i>	600
<i>Rhodotorula mucilaginosa</i>	610
<i>Lanchancea thermotolerans</i>	675
<i>Kluyveromyces marxianus</i>	720
<i>Kluyveromyces lactis</i>	740
<i>Hanseniaspora uvarum</i>	747
<i>Hanseniaspora guilliermondii</i>	749
<i>Torulaspora delbreuckii</i>	798
<i>Saccharomyces cerevisiae</i>	842

1.2.4.2.2. D1/D2 Domain Sequencing of Ribosomal DNA

D1/D2 domain covers 600 nucleotides in the sequence (Guadet et al., 1989). According to this domain, *Saccharomyces* and non-*Saccharomyces* can be identified (Kurtzman & Robnett, 1998). The research studies by Baleiras-Couto et al. (2006, 2010), demonstrated that wine yeast species were examined by restriction profiles of D1/D2 domain in the large unit of ribosomal DNA. In addition, sequencing of this domain has facilitated identification of species accurately and rapidly (Hesham et al., 2014). This identification method was used for *Saccharomyces* (Hesham et al., 2011), *Kluyveromyces* (Belloch, Barrio, Garcia, & Querol, 1998), and other wine yeast species (Guillamon, Sabate, Barrio, Cano, & Querol, 1998) (Table 1.5).

The following sequences show the used NL1 and NL4 primers (Figure 1.12):

NL-1 5' GCATATCAATAAGCGGAGGAAAAG 3'

NL-4 5' GGTCCGTGTTCAAGACGG 3' (O'Donnell, 1993).

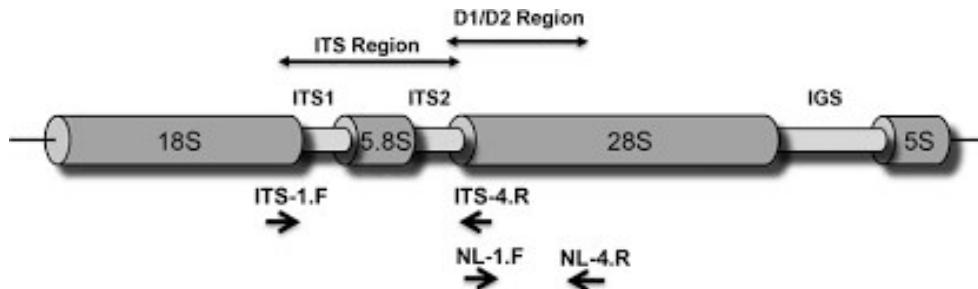


Figure 1.12. ITS Region and D1/D2 Domains of rRNA (Romanelli, Fu, Herrera, & Wickes, 2014)

Table 1.5. Used Identification Methods in the Literature (Fernandez-Espinar, Llopis, Querol, & Barrio, 2011)

<i>Studies</i>	<i>Used Methods</i>	<i>References</i>
NS and S	ITS Region and D1/D2 Domain	Tofalo et al. (2009)
NS	ITS Region	Zott et al. (2008)
NS	D1/D2 Domain	Romancino et al. (2008)
NS and S	ITS Region	Rodriguez et al. (2004)
NS and S	ITS Region	Esteve-Zarzoso et al. (2001)
S	ITS Region	Fernández-Espinar et al. (2000)
NS and S	ITS Region	Guillamon et al. (1998)

NS: Non-*Saccharomyces*; S: *Saccharomyces*.

1.2.5. Diversity of Indigenous Non-*Saccharomyces* Yeasts Isolated from Musts of Grapes

Indigenous non-*Saccharomyces* yeasts play role in wine quality due to metabolites they produced (Jolly, Augustyn, & Pretorius, 2006). These yeast strains indicate differences in their desirable characteristics according to regions from which they are isolated. The following table shows diversity of indigenous non-*Saccharomyces* yeasts according to regions from which they are isolated (Table 1.6):

Table 1.6. Diversity of Indigenous Non-Saccharomyces Yeasts According to Regions (Jolly et al., 2006)

Name of Isolated Yeast	Region or Country	Grape Variety
<i>Brettanomyces bruxellensis</i> (<i>D. bruxellensis</i>)	Tenerife	White Listan
<i>Candida colliculosa</i> (<i>T. delbrueckii</i>)	Catalonia	Macabeo and Grenache
<i>Candida guilliermondii</i> (<i>P. guilliermondii</i>)	Israel	Cabernet Sauvignon
<i>Candida krusei</i> (<i>I. orientalis</i>)	Argentina	Malbec
<i>Candida pulcherrima</i> (<i>M. pulcherrima</i>)	Israel	Cabernet Sauvignon and Muscat d'Alexandrie
	Argentina	Malbec
<i>Candida stellata</i>	Argentina	Malbec
<i>Candida valida</i> (<i>P. membranifaciens</i>)	Argentina	Malbec
<i>Kloeckera apiculata</i> (<i>H. uvarum</i>)	California	Vineyard
	Australia	Riesling, Semillon, Malbec and Hermitage
	Bordeaux	Semillon and Merlot
	Argentina	Malbec
<i>Kloeckera apis</i> (<i>H. guilliermondii</i>)	Spain	Abarino, Godello and Mencia
<i>Kluyveromyces thermotolerans</i> (<i>L. thermotolerans</i>)	Catalonia	Macabeo and Grenache
	Tenerife	White Listan
<i>Rhodotorula mucilaginosa</i>	Western Cape	Grapes
	Spain	Abarino, Godello and Mencia
<i>Pichia anomala</i> (<i>W. anomalus</i>)	Majorca	Chenin blanc
<i>Saccharomycodes ludwigii</i>	Spain	Ribeiro
<i>Zygosaccharomyces bailii</i>	Spain	White and red varieties

In addition to this table, non-*Saccharomyces* yeasts were isolated from different regions recently. In India, 10 different non-*Saccharomyces* yeasts (*Candida quercitrusa*, *Candida azyma*, *Hanseniaspora guilliermondii*, *Hanseniaspora uvarum*, *Hanseniaspora viniae*, *Issatchenkia terricola*, *Issatchenkia orientalis*, *Pichia membranifaciens*, *Zygoascus steatolyticus*) were isolated from Bangalore Blue, Cabernet, Zinfandel, Sauvignon Blanc, Chenin Blanc and Shiraz grape varieties (Chavan et al., 2009) while 17 yeast species (*Candida bombi*, *Candida inconspicua*, *Candida zemplinina*, *Candida quercitrusa*, *Cryptococcus carnescens*, *Cryptococcus flavesiens*, *Cryptococcus magnus*, *Hanseniaspora guilliermondii*, *Hanseniaspora uvarum*, *Issatchenkia terricola*, *Issatchenkia orientalis*, *Metschnikowia pulcherrima*, *Pichia fermentans*, *Pichia guilliermondii*, *Sporidiobolus pararoseus*, *Zygosaccharomyces bailii*, *Zygosaccharomyces fermentati*) were identified from Chardonnay, Cabernet Sauvignon, and Merlot grape varieties in China (Li et al., 2010).

Lately, few studies have been carried out to study the non-*Saccharomyces* yeasts isolated from grape and grape must of vineyards in Turkey. Çelik et al. (2017) found 8 different non-*Saccharomyces* yeasts species (*Candida zemplinina*, *Hanseniaspora guilliermondii*, *Hanseniaspora uvarum*, *Metschnikowia* spp., *Lachancea thermotolerance*, *Pichia kluveri*, *Pichia occidentalis*, *Torulaspora delbrueckii*) in Narince grapes from the region Tokat. In addition, 18 non-*Saccharomyces* (*Debaryomyces hansenii*, *Hanseniaspora guilliermondii*, *Hanseniaspora uvarum*, *Hanseniaspora opuntiae*, *Kluyveromyces marxianus*, *Metschnikowia chrysoperlae*, *Metschnikowia fructicola*, *Metschnikowia pulcherrima*, *Pichia anomala*, *Pichia kudriavzevii*, *Pichia sporocuriosa*, *Rhodotorula mucilaginosa*, *Pichia guilliermondii*, *Pichia sporocuriosa*, *Issatchenkia terricola*, *Issatchenkia terricola*, *Zygoascus hellenicus*, *Zygoascus meyerae*) species were isolated from grapes which were harvested from Adana, Urla, and Tekirdağ (Ükelgi, 2011). Moreover, Nurgel et al. (2005) found dominant non-*Saccharomyces* yeasts (*Kloeckera apiculate*, *Lachancea thermotolerance*, *Candida pulcherrima*) during fermentation of Emir grapes and

isolated non-*Saccharomyces* yeasts (*Kloeckera apiculata*, *Metschnikowia pulcherrima*, *Candida holmii*, *Candida valida*, *Candida guilliermondii* and *Candida sp.*) during alcoholic fermentation of Kalecik Karası grapes. Although this study of yeast flora for Kalecik Karası and Emir grape varieties in the Anatolia region was carried out, a study of yeast populations for five different grape varieties (Kalecik Karası, Öküzgözü, Boğazkere, Dimrit, Emir) taken from three different regions (Ankara, Elazığ and Cappadocia) in Turkey hasn't been conducted yet.

1.2.6. Non-*Saccharomyces* Yeasts Used as a Starter Culture

1.2.6.1. *Hanseniaspora guilliermondii*

Hanseniaspora genera comes to the forefront of acetate ester producer. A study was carried out by Viana et al. (2008), that 11027 and 11102 strains of *Hanseniaspora guilliermondii* were selected as a starter culture due to ester profile for enological characterization. In addition, different strains of *Hanseniaspora guilliermondii* strongly produce ethyl acetate and 2-phenylethyl acetate which give flowery and fruity aroma in must (Nathalie Moreira, Mendes, Hogg, & Vasconcelos, 2005; Rojas, Gil, Piñaga, & Manzanares, 2003). In addition, some strains of this microorganism have ethanol tolerance like *Saccharomyces cerevisiae*. This microorganism resists up to 25% (v/v) ethanol level (Pina, Santos, Couto, & Hogg, 2004).

1.2.6.2. *Hanseniaspora opuntiae*

Hanseniaspora opuntiae affects positively wine aroma, especially sensory of wines. this microorganism with inoculation of *Saccharomyces cerevisiae* increases aroma of the wines and gains specific profile in to the wine. A study showed that usage of *Hanseniaspora opuntiae* and *Saccharomyces cerevisiae* var. *bayanus* in Cabernet sauvignon, Chenin Blanc, Rose grapes gave a typical wine flavour. These wines were preferred by consumers due to positive sensory qualities (Assis et al., 2014).

1.2.6.3. *Hanseniaspora uvarum*

Hanseniaspora uvarum is used microorganism as a starter culture for winemaking process. Although this microorganism causes stuck fermentation, some researchers used this microorganism with sequential or co-inoculation with *Saccharomyces cerevisiae* to use industrial production of wine. This usage decreases volatile acidity component of a wine and gives organoleptic quality to the wine (Tristezza et al., 2016).

1.2.6.4. *Rhodotorula mucilaginosa*

A study in China showed that *Rhodotorula mucilaginosa* which has a high level of β-glucosidase gave fruity and floral aroma in the wine due to glycosides of C₁₃-norisoprenoids and benzenic compounds (Hu et al., 2016). In addition, *Rhodotorula mucilaginosa* and *Saccharomyces cerevisiae* were used with sequential inoculation in dry white wine to enhance aroma profile. Nerol oxide, certain ethyl and acetates compounds and (Z)-3-hexene-1-ol were found in this mixed fermentation. They gave berry, citrus, floral, sweet and acid fruit aroma in this wine (Wang et al., 2017).

1.2.6.5. *Wickerhamomyces anomalus* (*Pichia anomala*)

A study of the mixed culture of *Saccharomyces cerevisiae* and *Wickerhamomyces anomalus* showed that this co-inoculation affects quality of cider positively. In addition, it reached same ethanol level as monoculture of *Saccharomyces cerevisiae*. This mixed culture enhanced remarkably higher alcohols, ethyl esters, acetate esters, ketones and aldehydes than pure culture fermentation. It found more desirable than the product of monoculture fermentation (Ye, Yue, & Yuan, 2014).

1.2.6.6. *Metschnikowia* spp.

Metschnikowia pulcherrima is very use species in this genus. It significantly reduces volatile and total acidity of the wines. This usage of mixed inoculation with *Saccharomyces cerevisiae* affect isoamyl acetate, 2-phenyl ethanol, and medium-chain fatty acids production positively. It also increases polysaccharides content in the wine (Comitini et al., 2011).

1.2.6.7. *Lachancea thermotolerans* (*Kluyveromyces thermotolerans*)

Lachancea thermotolerans and *Saccharomyces cerevisiae* are used for enhancement of acidity and wine quality improvement by simultaneous and sequential fermentation. A study has shown that glycerol and 2-phenylethanol enhanced with co-inoculation of these microorganisms. In addition, spicy aroma and total acidity increased with these two microorganisms using starter cultures (Gobbi et al., 2013).

Lachancea thermotolerans is used a commercial starter culture by Chr. Hansen (Viniflora® SYMPHONY). It is used for improving aroma in white (Riesling, Pinot Blanc, Chardonnay and Pinot Gris) and red (Pinot Noir, Shiraz, Cabernet Sauvignon and Merlot) grapes. It gives tropical fruit and floral aroma (Jolly et al., 2006).

1.2.6.8. *Starmerella bacillaris* (*Candida zemplinina*)

A study was carried out to the molecular and physiological characterization of *Starmerella bacillaris* yeasts isolated from grapes in Italy. This study has shown as this microorganism produced a low amount of acetic acid and ethanol as a high level of glycerol. Therefore, this microorganism is suitable for application of mixed fermentation with *Saccharomyces cerevisiae* for winemaking. In addition, this mixed culture can be used for reduction of ethanol in wine (Englezos et al., 2015).

1.2.6.9. *Williopsis saturnus*

Williopsis saturnus (*Hansenula saturnus*) produces a significant level of volatile esters, particularly isoamyl acetate and ethyl acetate. Isoamyl acetate gives banana-like aroma in the wine (Wang et al., 2017). Although this non-*Saccharomyces* yeast does not find grapes surfaces and equipment of winery, it produces desirable flavour. *Williopsis saturnus* potentially enhances fruity aroma in the wines which were obtained from characteristics of neutral cultivar. Therefore, *Williopsis saturnus* was used as a starter culture with *Saccharomyces cerevisiae* to produce the wine. Erten and Tanguler (2010) carried out the study of mixed inoculation of *Williopsis saturnus* and *Saccharomyces cerevisiae* in Emir wine.

1.2.7. *Saccharomyces* Yeasts Used as a Starter Culture

Saccharomyces cerevisiae is known as the wine yeast due to an important species in alcoholic fermentation (Tofalo et al., 2013). It converts all sugars in the grapes to ethanol and CO₂ during fermentation process due to ethanol resistance. Therefore, it is an indispensable species for wine-making.

1.2.8. Characterization of Yeasts and Selected Yeasts for Starter Culture

Isolated yeasts should be characterized to be selected as a starter culture for wine-making. The followings show the features of a starter culture:

- Having high alcohol tolerance
- Having SO₂ tolerance
- Having temperature tolerance
- Having pH tolerance
- Producing low level H₂S
- Producing low volatile acid
- Converting sugars to ethanol at suitable rate in the must
- Fermenting all sugars
- Having high fermentation speed
- Producing low level of sulphur dioxide
- Producing low foam
- Having a killer property
- Producing low level of acetaldehyde
- Producing enough level of glycerol
- Settling easily after fermentation (Bağder, 2008; Nikolaou, Soufleros, Bouloumpasi, & Tzanetakis, 2006)

1.3. Fermentation Process

Wine is the result of a complicated biochemical and biological interaction between grapes and various microorganisms such as non-*Saccharomyces* and *Saccharomyces*

yeasts, acetic acid and lactic acid bacteria, other fungi (Fleet, 2003). This process begins to a vineyard and moving through fermentation and ageing.

Yeast have a critical role in this process because they convert sugar grape to ethanol and carbon dioxide (CO_2). In addition, even though flavour and quality of the wine depend on grape variety directly and other factors such as weather, soil structure, wine making practices, it affects yeasts with products of metabolism through growth and autolysis (Fleet, 2003; Jolly et al., 2006; Swiegers & Pretorius, 2005). Moreover, maceration process is important for yeasts especially cold maceration. *Saccharomyces* yeasts stand at stationary phase, non-*Saccharomyces* yeasts grow at 4°C . Therefore, cold maceration provides to flavour improvement and colour extraction (Hierro, González, Mas, & Guillamón, 2006).

Fermentation process occurs by two ways: spontaneous (traditional) or inoculation way.

1.3.1. Spontaneous Fermentation

Spontaneous fermentation occurs without addition of yeasts. It is a traditional method. Yeasts which locate on the grapes or wine equipment's start fermentation process. Therefore, this wine has a unique property. Aroma profile of this wine doesn't appear similar the other parallel wines. Sometimes, it has high quality. However, it can be bad because of dominant spoilage yeasts such as *Dekkera bruxellensis*. In addition, stuck fermentation can occur in this fermentation (Jolly et al., 2006).

This method is not preferable in Turkey excluding homemade wine while this is used by commercially producers in Europe (Pretorius, 2000).

1.3.2. Inoculated Fermentation

Inoculated fermentation is a more reliable method than spontaneous fermentation because the wine can have the wanted properties such as alcohol level or H_2S production level. This quality of wine doesn't change year by year with the help of the used commercial strain.

1.3.2.1. Inoculation of Non-*Saccharomyces* Yeasts

Many studies have shown that non-*Saccharomyces* yeasts contribute to a more complicated aroma and enhanced quality of wine (Maurizio Ciani, Beco, & Comitini, 2006; Henick-Kling, Edinger, Daniel, & Monk, 1998; Romano, Suzzi, Comi, Zironi, & Maifreni, 1997).

A lot of studies have indicated that using non-*Saccharomyces* starter cultures improve the quality of wine especially in a controlled manner (Maurizio Ciani et al., 2006; Erten, 2002; Toro & Vazquez, 2002).

1.3.2.2. Inoculation of *Saccharomyces* Yeasts

Although non-*Saccharomyces* yeasts improve wine quality, generally, they don't complete fermentation because of low level of alcohol tolerance. In other words, non-*Saccharomyces* yeast are not able to convert all sugar in the grapes to ethanol. Therefore, usage of *Saccharomyces* yeast is needed. In addition, *Saccharomyces* plays an important role for final quality and bouquet of the wine (Cocolin, Pepe, Comitini, Comi, & Ciani, 2004).

There are three type of inoculation: pure, sequential or co-inoculation. In pure fermentation, a yeast is added as a starter culture purely. In sequential inoculation, firstly, a non-*Saccharomyces* yeast is added, then, *Saccharomyces cerevisiae* can be added after 4-7 days later with respect to fermentation temperature (Maurizio Ciani et al., 2006). In co-inoculation, non-*Saccharomyces* and *Saccharomyces cerevisiae* are added in the must at the same time (Padilla, Gil, & Manzanares, 2016).

1.3.2.3. Mixed Inoculation

Selected non-*Saccharomyces* and *Saccharomyces cerevisiae* are combined with respect to abilities. These combinations have unique aromatic properties. According to their ability to generate enzymes that improve flavor or alter secondary metabolites concentration, various combinations of mixed starter cultures have been created and analyzed to improve wine quality (Padilla et al., 2016).

There are two mixed inoculation methods: sequential or co-inoculation. These two methods are used by researchers and winemakers. The following table was shown inoculation types and effects of the wine quality for using non-*Saccharomyces* and *Saccharomyces* mixed culture (Table 1.7).

Table 1.7. Effects of Mixed Inoculations (Padilla et al., 2016)

<i>Mixed Inoculation</i>	<i>Effects</i>	<i>Type of Inoculation</i>	<i>References</i>
<i>C. zemplinina</i> & <i>S. cerevisiae</i>	Decreasing acetic acid	Sequential, co-inoculation	(M. Ciani & Ferraro, 1998; Rantsiou et al., 2012)
<i>H. guilliermondii</i> & <i>S. cerevisiae</i>	Increasing acetate ester and sulphur compound	Co-inoculation	(Moreira et al, 2008)
<i>H. uvarum</i> & <i>S. cerevisiae</i>	Increasing acetate ester	Co-inoculation	(Andorrà et al., 2010)
<i>H. vineae</i> & <i>S. cerevisiae</i>	Increasing ethyl ester and acetate	Sequential, co-inoculation	(Medina et al., 2013)
<i>I. orientalis</i> & <i>S. cerevisiae</i>	Deacidification of wine	Co-inoculation	(Kim, Hong, & Park, 2008)
<i>L. thermotolerans</i> & <i>S. cerevisiae</i>	Acidification of wine	Sequential, co-inoculation	(Gobbi et al., 2013b)
<i>M. pulcherrima</i> & <i>S. cerevisiae</i>	Decreasing acetic acid and increasing α -terpineol, ethyl ester, alcohol level	Sequential, co-inoculation	(Comitini et al., 2011; Zohre & Erten, 2002)
<i>P. fermentans</i> & <i>S. cerevisiae</i>	Decreasing acetic acid and increasing alcohol level	Sequential, co-inoculation	(Clemente-Jimenez, et al, 2005; Comitini et al., 2011)
<i>T. delbrueckii</i> & <i>S. cerevisiae</i>	Decreasing acetic acid and increasing alcohol level, ethyl ester, acetate, linalool and α -terpineol	Sequential, co-inoculation	(Azzolini et al, 2015; Comitini et al., 2011)
<i>W. anomalus</i> & <i>S. cerevisiae</i>	Increasing ethyl ester and acetate	Sequential	(Cañas, García, & Romero, 2011)
<i>W. saturnus</i> & <i>S. cerevisiae</i>	Increasing acetate ester	Co-inoculation	(Erten & Tanguler, 2010)
<i>Z. bailii</i> & <i>S. cerevisiae</i>	Increasing ethyl ester	Co-inoculation	(Garavaglia et al., 2015)

1.4. Aroma Profiles of Wine

Aroma profile plays an important role in the wine quality. Wine aroma consists of 100s of various compounds with levels varying from 10^{-1} to 10^{-10} g/kg (Rapp & Mandery, 1986). Interaction and balance of these compounds affect quality of wine aroma.

There are three type of wine aroma: primary (varietal) aroma, secondary (fermentation) aroma, and tertiary (bouquet) aroma (Padilla et al., 2016).

1.4.1. Primary Aroma

Primary aroma relates to variety of grape. It affects ripeness of grapes. Primary aroma includes C₁₃-norisoprenoids, terpenes, methoxypyrazines, and volatile sulfur compounds (Ebeler & Thorngate, 2009). Carotenoids is derived by C₁₃-norisoprenoids. Non-floral aroma is given by C₁₃-norisoprenoids, especially β -damascenone and β -ionone (Fang & Qian, 2016). Terpenes found all grape varieties, but especially, it is high level in Muscat and Rhine Riesling (King & Dickinson, 2000). 70 terpenes found in the grape and their wines such as monoterpenoid alcohols (Mateo & Jimenez, 2000). They are α -terpineol, linalool, citronellol, geraniol, and nerol which are widest and stronger participants to aroma of wines (Carrau et al., 2005). In addition, they have low value of thresholds and give floral aroma (Zalacain, Marin, Alonso, & Salinas, 2007). Methoxypyrazines are produced by metabolism of amino acids. They give green, vegetal, and herbaceous aroma in wine (Sidhu et al., 2015). Volatile sulfur compounds also give aromatic thiols in the wine. Especially, it is important for Sauvignon Blanc wine (Bouchilloux, Darriet, & Henry, 1998).

1.4.2. Secondary Aroma

Secondary aroma relates to yeasts and conditions of fermentation. Secondary aroma includes high levels of alcohols, volatile fatty acids, esters, and low level of aldehydes (Rapp & Versini, 1995). In addition, compounds containing sulfur or nitrogen contribute wine aroma.

1.4.2.1. Higher Alcohols

Most aromatic compounds are higher alcohols. They give complex aroma under 300 mg/L concentration level. On the contrary, above 400 mg/L concentration level, they give negative effect on wine (Rapp & Mandery, 1986).

Pure *Hanseniaspora* species produce higher alcohol than its mixed inoculation with *S. cerevisiae* during must fermentation. Moreover, *Zygosaccharomyces* produces low level of high alcohols while *C. zemplinina* produces high level of these alcohols and it exceeds 400 mg/L (Andorrà et al., 2010; Romano et al., 1993).

2-phenylethyl alcohol, especially, gives pleasant aroma in wines and it is a characteristic's property of some non-*Saccharomyces* such as *M. pulcherrima*, *L. thermotolerans*, *C. zemplinina* (Andorrà et al., 2010; Beckner et al., 2015; Josefa María Clemente-Jimenez, Mingorance-Cazorla, Martínez-Rodríguez, Las Heras-Vázquez, & Rodríguez-Vico, 2004).

1.4.2.2. Volatile Fatty Acids

The majority (90%) of volatile fatty acids in wines is acetic acid. The rest of them are butanoic and propanoic acids. Acetic acid gives unpleasant flavour between 0.7 and 1.1 g/L range while 0.2-0.7 g/L is optimal range (Lambrechts & Pretorius, 2000).

Some non-*Saccharomyces* yeasts produce high amount of acetic acids such as *Hanseniaspora*, *Zygosaccharomyces*, and *Schizosaccharomyces pombe* (Mendoza, De Nadra, & Farias, 2007; Snow and Gallander, 1979). However, this property can change strain to strain. For example, strains of *H. uvarum* produce between 0.6 g/L and 3.4 g/L acetic acid (Romano, Fiore, Paraggio, Caruso, & Capece, 2003a). On the other hand, the other non-*Saccharomyces* have a property of low acetic acid production such as *T. delbrueckii*, *L. thermotolerans* and *C. zemplinina* (Comitini et al., 2011). *L. thermotolerans* produces low acetic acid whereas high L-lactic acid. *C. zemplinina* also produces low acetic acid and ethanol whereas high glycerol (Englezos et al., 2015; Rantsiou et al., 2012).

1.4.2.3. Esters

Esters consist of isoamyl acetate, 2-phenylethyl acetate, ethyl acetate, isobutyl acetate, deriving from acetic acid, and ethyl caprylate, ethyl butanoate, ethyl caprate, ethyl caproate, originating from saturated fatty acids of esters. Ethyl acetate is primary ester and has negative effect at 150-200 mg/L concentration (Lambrechts & Pretorius, 2000).

Some non-*Saccharomyces* have capacity of high level of ethyl acetate production such as *Candida*, *Pichia*, and *Hanseniaspora*. *Pichia* and *Hanseniaspora* have similar ethyl acetate production level. In addition, *Hanseniaspora* produces isoamyl acetate and 2-phenylethyl acetate which are esters of fruity acetate. In the species level, *H. uvarum* is generally a successful producer of esters while *H. osmophila* and *H. guilliermondii* give 2-phenylethyl acetate. Moreover, *Rhodotorula* and *Pichia* give isoamyl acetate significantly (Romano et al., 1997; Viana et al., 2008).

1.4.2.4. Aldehydes

Aldehydes give apple-like flavour and their thresholds values are low. The concentration level in the wines changes between 10 mg/L and 300 mg/L (Lambrechts & Pretorius, 2000).

Usually, *Saccharomyces cerevisiae* generates greater concentrations of acetaldehyde (5-120 mg/L) than non-*Saccharomyces* (max 40 mg/L) such as *M. pulcherrima*, *C. krusei*, *H. anomala*, *C. stellata*. *H. uvarum* can produce 25 mg/L acetaldehyde with respect to the strains (Romano, Fiore, Paraggio, Caruso, & Capece, 2003).

1.4.2.5. Phenols

Vinylphenols and ethylphenols are very important compounds for white and red wines, respectively. *p*-coumaric and ferulic acid produce these phenols. They give an undesirable flavour in wines. *Brettanomyces* produces ethylphenol (Lambrechts & Pretorius, 2000). On the other hand, *H. osmophila*, *H. guilliermondii*, and *P.*

membranifaciens produce neither ferulic nor *p*-coumaric acids (Padilla et al., 2016; Viana et al., 2008).

1.5. The Objectives of the Study

Starter cultures are the microorganisms used for the production of bakery and dairy products such as cheese and yogurt or fermented alcoholic beverages such as beer and wine. The following table shows international starter culture producers and their products (Table 1.8):

Table 1.8. International Starter Culture Producers

<i>Company Name</i>	<i>Products</i>
AB Mauri Food Inc. (Fleischmann)	Baker's yeast
Chr. Hansen A/S	Cheese and dairy products and alcoholic beverages' cultures
LAFFORT® Company	Wine yeast
Lesaffre Yeast Corporation (Red Star)	Wine and baker's yeast
Lallemand Inc.(Lalvin, Anchor)	Beer and wine yeasts
Mangrove Jack's, BSGi NZ Ltd	Beer, cider and wine yeasts
Bio Sunkeen Co., Ltd.	Beer, wine and baker's yeasts
Martin Vialatte	Wine yeast
Angel Yeast Co., Ltd. Gloripan	Baker's yeast
Koninklijke DSM N.V.	Cheese and dairy products and alcoholic beverages' cultures
Dow Inc. (DuPont (Danisco, Holdback))	Cheese and dairy products' cultures
Biolacter Inc	Cheese and dairy products' cultures
Proxis Développement Sas (Bioprox)	Lactic starter cultures for cheese, yogurt, fermented milk, butter, bakery and oenology

On the other hand, there is only one starter culture producer in Turkey. Pak Food Production and Marketing Inc. (Pakmaya) produces bakery yeasts. However, there are no starter culture producers for winemaking in Turkey.

Traditional wine is the product of spontaneous fermentation by indigenous yeasts in the grapes. Although this wine process doesn't achieve standard properties of wine due to changes in climate, regions and years, starter culture addition in the wine is preferred. However, technology with starter cultures provides standard and ordinary products. Thus, researchers and winemakers have begun to prefer using native strains as starter cultures to adapt in the native grape must easily and give specific odour properties in this wine (Capece et al., 2010; Cocolin et al., 2004). Native non-*Saccharomyces* species produce authentic wines because they impart significant regional and desirable properties (Jolly et al., 2006). In addition, native *Saccharomyces cerevisiae* strains yield easily in native grape types due to high adaptation properties (Cappello, Bleve, Grieco, Dellaglio, & Zacheo, 2004).

The objectives of this study are:

- To learn about non-*Saccharomyces* and *Saccharomyces* yeast populations in local grapes at different maceration and fermentation times in Turkey
- To isolate non-*Saccharomyces* and *Saccharomyces* yeasts in local microflora of the grapes
- To identify and characterize the isolates
- To select high properties yeasts as starter culture
- To make wine with using selected yeasts and analyze volatile compounds of these wines.

Therefore, this study can provide to encourage using these selected native strains as a starter culture for boutique or industrial wine-making production of native grapes in Turkey.

CHAPTER 2

MATERIALS AND METHODS

2.1. Materials

Commercial *Saccharomyces cerevisiae* starter cultures were Chr. Hansen: Viniflora® Merit and Laffort: Zymoflore VL1 Vin Blanc, used for making commercial red and white wines, respectively.

Yeast extract, peptone from casein (tryptone), lysine agar, lactose, mannitol, maltose, sucrose were purchased from Lab M Limited. Moreover, D-(+)-glucose monohydrate, Biggy agar, agar-agar, cycloheximide were bought from Sigma-Aldrich Chemical Co. In addition, the brand of chloramphenicol and buffered peptone water was VWR International Company.

2.2. Methods

2.2.1. Winemaking

2.2.1.1. Source of Grapes and Production

Wines were produced with the traditional method and the commercial method by the using commercial strain. Using different *Saccharomyces cerevisiae* strains cause different taste and aroma profile (Bisson, 2012).

5 different wines were produced from 5 different kinds of grapes: Kalecik Karası, Öküzgözü, Boğazkere, Dimrit and Emir (Figure 2.1). These grapes were gathered different time and provinces as shown in Table 2.1.

Table 2.1. Grape Variety Collected from Three Different Region from Turkey

Grape Variety	Region	Time
Kalecik Karası	Ankara (Kalecik)	11.09.2017
Dimrit	Kapadokya, Ürgüp	19.09.2017
Emir	Kapadokya, Ürgüp	29.09.2017
Öküzgözü	Elazığ	01.10.2017
Boğazkere	Elazığ	01.10.2017
Emir	Kapadokya, Ürgüp	19.09.2018
Öküzgözü	Elazığ	21.09.2019

The grapes mentioned in the Table 2.1 could reach Ankara under 24 hours.



Figure 2.1. Grape Varieties Collected from Three Different Region of Turkey

2.2.1.2. Separation of Grapes from Stems and Crushing

Incoming grapes were separated from the stalks and detonated. At this stage, attention was given to blowing the seeds without damaging them. In this way, the bitterness of grape seed was prevented from getting into the must.

Each grape was broken down to 3 immediately after the explosion and the wine production process begun. The first two parallel were used for traditional wine production, the last parallel was used for commercial wine production. In this commercial wine production, concentration of SO₂ was calculated 30 mg/L for adding potassium metabisulphite (Fugelsang & Edwards, 2007). In addition to traditional wines, commercial wines have been obtained with the use of commercial strains. The use of two parallel runs in this phase was intended to prevent any possible

contamination. The purpose of doing commercial wine was to provide positive control. At this stage, samples were taken from each grape variety for measuring specific gravity, pH, temperature, brix, dry matter and isolating yeast.

After crushing of grapes, specific gravity of grapes varies between 1100 and 1090 showed the amount of sugar content of grapes. This refers the potential of alcohol content at the final wine. In addition, pH can be changed between 3.3 and 3.7 (Baker, J. & Clarke, 2012).

2.2.1.3. Maceration

There were two types of maceration in wine making: one was cold maceration and the other was normal maceration. The application of cold maceration is not obligatory, but it applies to enrich the aroma of wine (Hierro et al., 2006).

Therefore, the musts were first kept at 4 °C for 4 days and the specific weights were measured every day by hydrometer. At the end of the 4th day, the specific weights were measured, and after the samples were taken, they were left at the normal maceration stage at 20°C. During this process, the temperature was stabilized at 20°C by help of air conditioner.

In both stages of the maceration, the musts were mixed twice a day in order to get oxygen and prevent growing molds (Figure 2.2).



Figure 2.2. Mixing Process of Must Grapes in The Maceration Stage

2.2.1.4. Inoculation of Commercial Yeast, Fermentation Process, Removal of Pulp

In the traditional wine making process, any inoculation method was not applied. Only in the commercial wine making process, the commercial strain was used for positive control. The brand name of this commercial strain was Viniflora® Merit of Chr. Hansen which is a pure *Saccharomyces cerevisiae* starter culture (Figure 2.3).

Firstly, this starter culture, which had been waiting at 4°C, was weighed according to 1 g yeast/4 L must and secondly, activated with pure water for 10 minutes at 35°C. After that, a little bit must be taken and added this activated culture. Then, it was kept at 35°C for 20 minutes in order to adapt this yeast in must. Finally, this mix was added in the must.

When the red grapes reach the specific gravity between 1010 and 1020, the stage of maceration was terminated, and the must was squeezed with the help of a cheesecloth (Hyma et al., 2011). The must which was removed from the pulp was taken into glass jars and the mouth of the vessel was closed with an airlock to prevent oxygen ingress. Thereafter, alcoholic fermentation started in an anaerobic environment and continued at room temperature of 25 °C until specific gravity reaches 990 which refers to 15% (v/v) and 12% (w/w) alcohol content (Baker, J. & Clarke, 2012).

The first measurements of white grapes were made and after the sample was taken, it was squeezed with the help of a cheesecloth and the mouths were closed with air locks in glass jars to start alcoholic fermentation for an anaerobic environment. Alcoholic fermentation continued at 18°C due to the improvement of specific aroma components of white wines (Çelik et al., 2017). For commercial white wine, Laffort: Zymoflore VL1 Vin Blanc *Saccharomyces cerevisiae* culture was used (Figure 2.4).

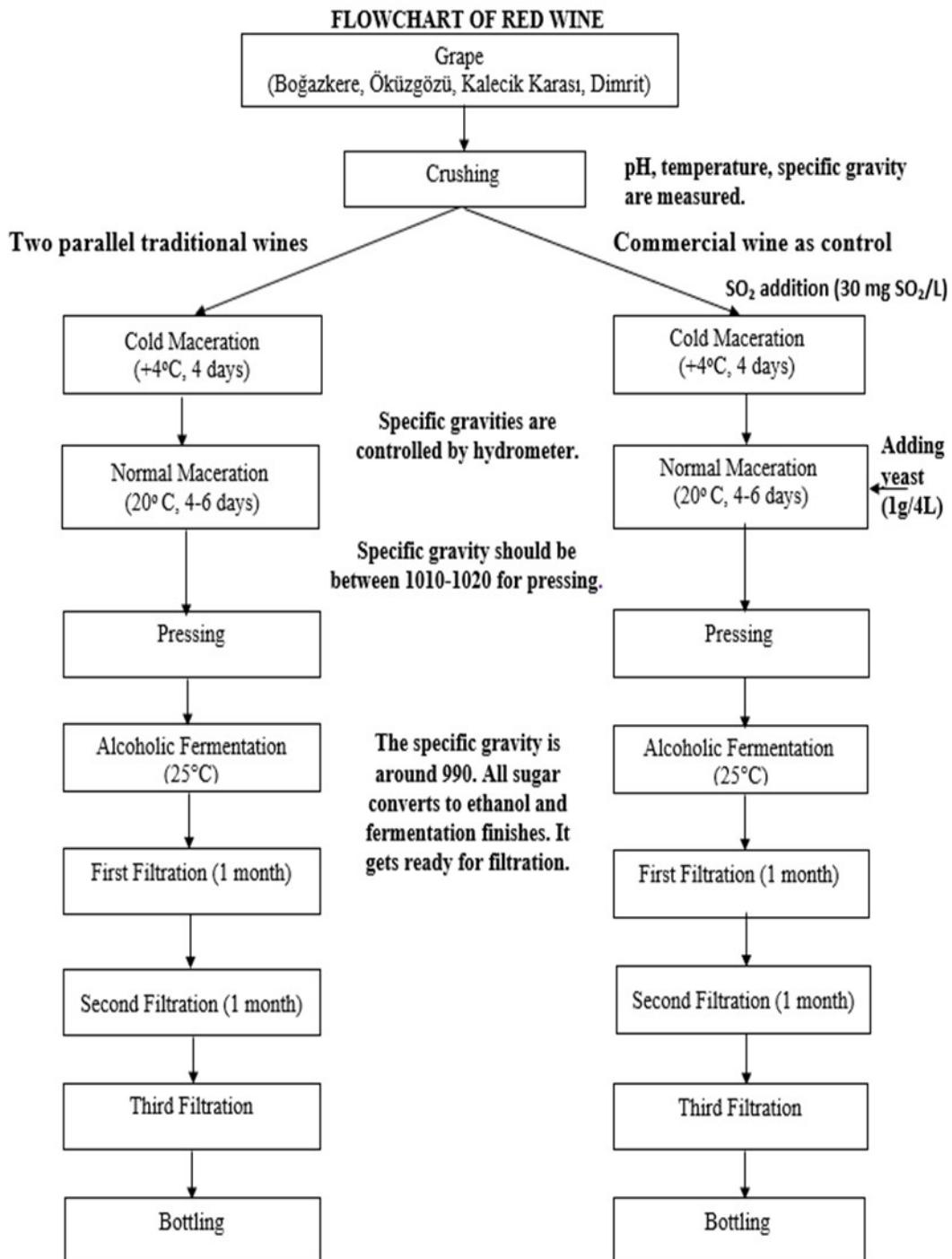


Figure 2.3. Flowchart of Red Wine Making

FLOWCHART OF WHITE WINE

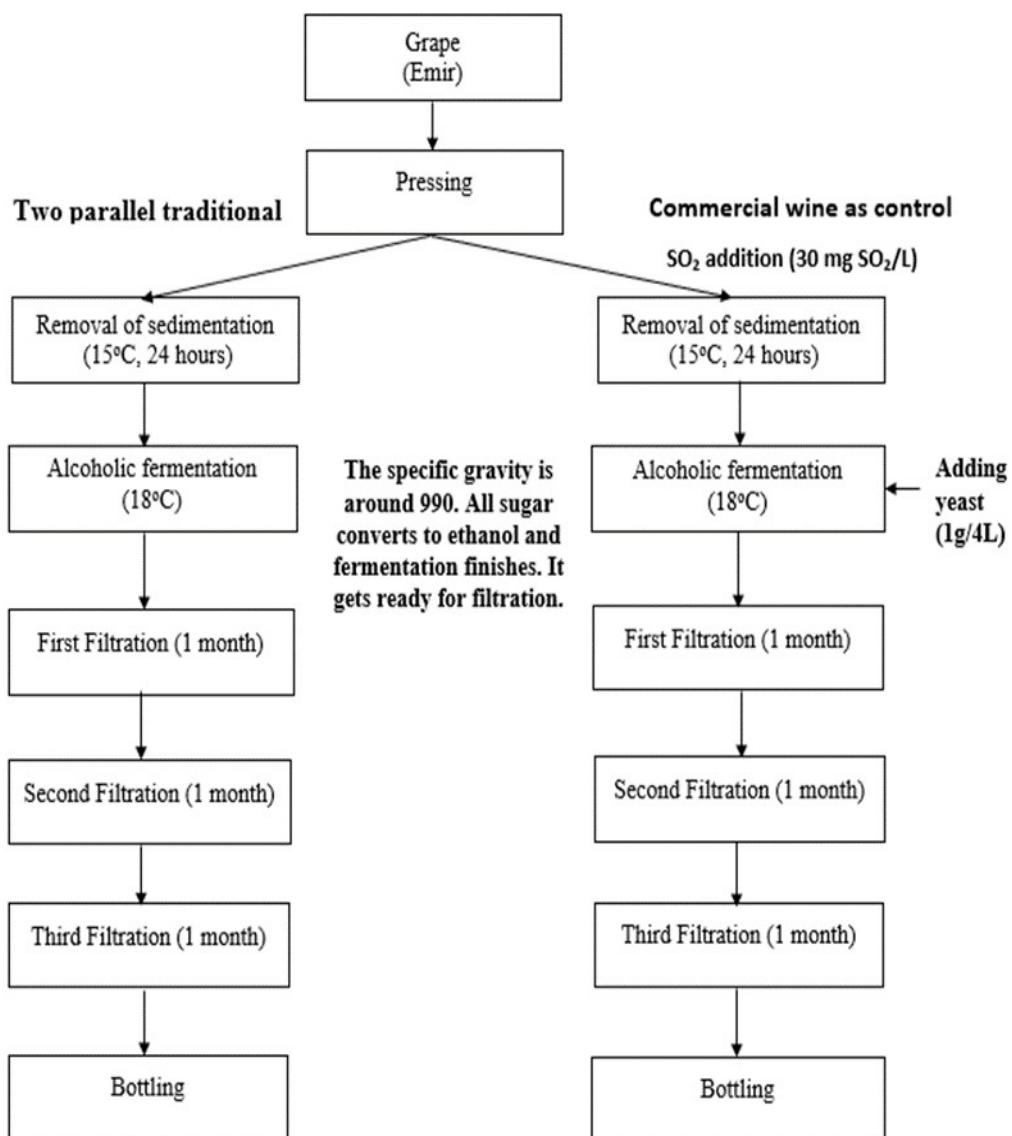


Figure 2.4. Flowchart of White Wine Making

2.2.1.5. Filtration and Bottling

Filtration was applied by siphonage method with 3 times totally in order to clarify the wine. After the filtration process, the wines were taken in green bottles with the help of siphonage method and the cork caps were attached (Figure 2.5).



Figure 2.5. Bottling

2.2.2. Isolation of Non-*Saccharomyces* and *Saccharomyces* Yeasts

The samples were taken at the specified times and diluted with peptone water. In this isolation process, two different media were used for non-*Saccharomyces* and *Saccharomyces* yeasts. The first one was selective mediums for non-*Saccharomyces* yeast. One of the selective mediums non-*Saccharomyces* yeast was yeast extract-peptone-glucose (YPG) agar (10 g/L yeast extract, 10 g/L peptone from casein (tryptone), 20 g/L glucose, 20 g/L agar) supplemented with cycloheximide (1 µg/mL), biphenyl (0.15 g/L of a 7.5 g/L stock solution), and chloramphenicol (0.1 g/L of a 5 g/L stock solution) (Zott et al., 2010). The other medium was lysine agar containing 66 g/L lysine agar, 1 ml potassium lactate, and 0.1 ml of 10% lactic acid (Van Der Aa Kühle & Jespersen, 1998).

The selective medium which was used for *Saccharomyces* was ethanol sulfate agar (ESA) containing 20 g/L glucose, 5 g/L yeast extract, 5 g/L yeast extract, 20 g/L agar, 0.15 g/L sodium metabisulfite, and 10% (v/v) ethanol. (Kish et al., 1983; Heard et al., 1985). Growing microorganisms on these mediums were subjected to purification twice in YPG agar. At the last stage, a single colony was obtained and inoculated on

the liquid medium. Thereafter, stock cultures were made to be 15% glycerol solution and put at -80 ° C (Figure 2.6).

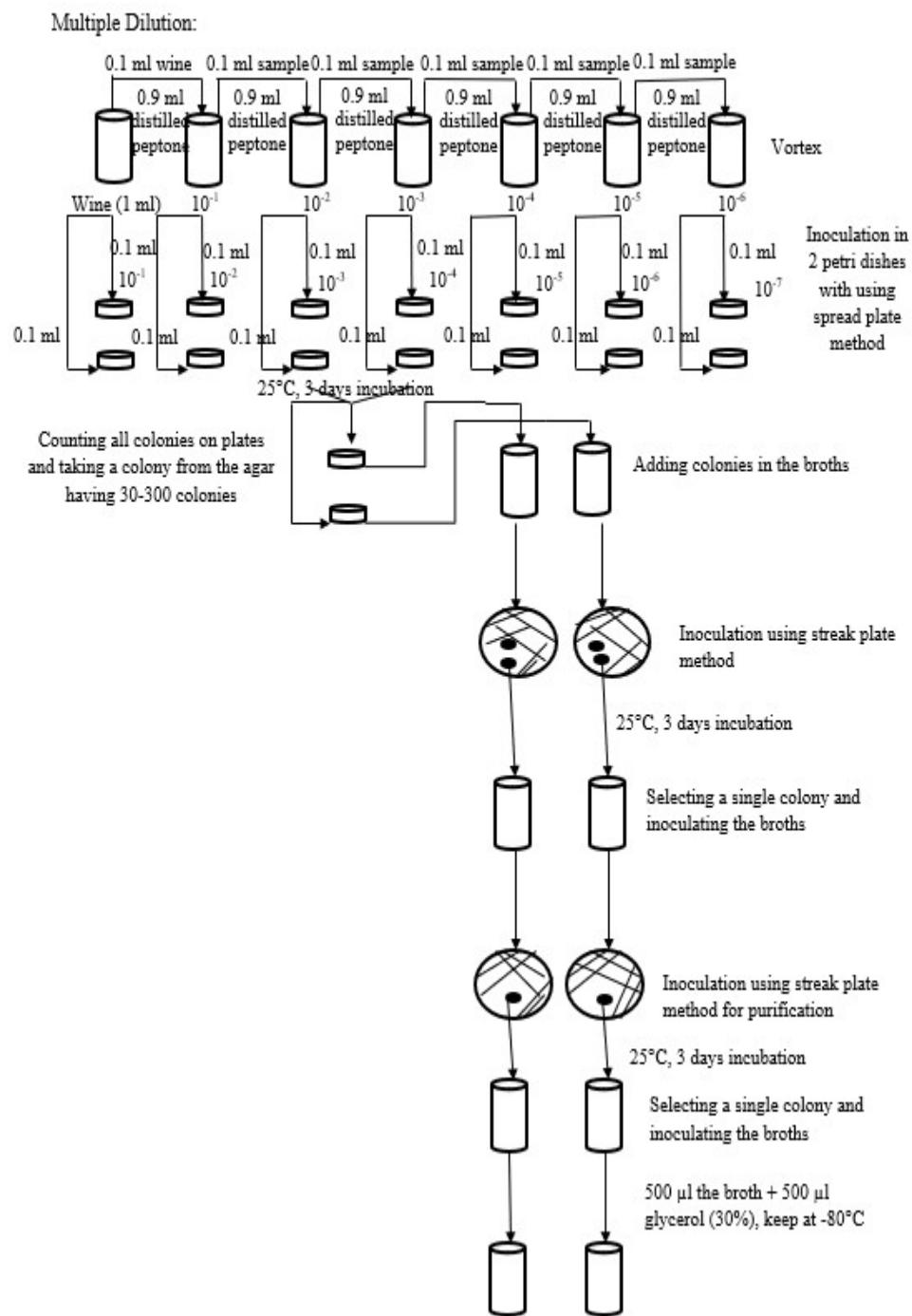


Figure 2.6. Isolation Method of *Saccharomyces* and Non-*Saccharomyces* Yeasts

2.2.3. Identification of Non-*Saccharomyces* and *Saccharomyces* with Molecular and Biochemical Techniques

2.2.3.1. Identification with Molecular Technique

EurX Gene Matrix Bacterial & Yeast DNA isolation kit (E3580) was used for DNA isolation according to the instruction. Spectrophotometric measurements were made on a Thermo Scientific Nanodrop 2000 (USA) instrument in order to check the amount of the DNA and purity. The ratio of absorbance at 260 nm and 280 nm was used to assess the purity of DNA. Approximately, a ratio of 1.8 was accepted as pure for DNA. PCR amplifications were performed using universal primers ITS1 and ITS4 primers. NL-1 and NL-4 primers were used for identification of repeated yeasts.

The following sequences were the used primers for ITS region and D1/D2 domain, respectively:

ITS1 5' TCCGTAGGTGAACCTGCGG 3'

ITS4 5' TCCTCCGCTTATTGATATGC 3' (Chavan et al., 2009; White et al., 1990)

NL-1 5' GCATATCAATAAGCGGAGGAAAAG 3'

NL-4 5' GGTCCGTGTTCAAGACGG 3' (O'Donnell, 1993).

The PCR conditions were as follows:

- Initial denaturation at 95 ° C for 5 minutes
- During 35 cycles
 - Denaturation at 95 ° C for 45 seconds
 - Annealing at 57 ° C for 45 seconds
 - Extension at 72 ° C for 60 seconds
- Final extension for 5 minutes at 72 ° C
- The temperature was reduced to 4 ° C and the PCR was completed.

PCR reaction was performed with Solis Biodyne (Estonia) FIREPol® DNA Polymerase, Taq polymerase enzyme. After PCR, Kyratec SuperCycler Trinity (Australia) was used. In agarose gel electrophoresis, a single band was obtained for our samples and PCR was successful. The marker was a 100 bp Solis Biodyne (Estonia) DNA ladder Ready to Load.

In order to purify PCR products, ExoSAP-IT™ PCR Product Cleanup Reagent (ThermoFisher Scientific, USA) was used. Purified PCR products were sequenced with the aid of forward and reverse primers. After gel electrophoresis, sequencing of PCR products was commercially provided by BM Laboratory Systems.

2.2.3.2. Identification with Biochemical Technique

2.2.3.2.1. Carbohydrate Fermentation Test

Isolates were subjected to the test for the property of carbohydrate fermentation. In this test, firstly, isolates were grown in 15 ml YPG (10 g/L yeast extract, 10 g/L peptone from casein (tryptone), 20 g/L glucose) broths at 28°C. The amounts of cells in the broths were measured in the spectrometer at 600 NM. When OD reached 1 in spectrometer, it refers to 10^6 CFU/ml in the broths. After the number of microorganisms reached 10^6 CFU/ml, they were centrifuged. They were washed twice with autoclaved distilled water in order to get rid of the medium especially glucose.

Moreover, 4.5 g/L yeast extract and 7.5 g/L peptone were dissolved in distilled water and bromothymol blue was added in order to give adequately green color. Bromothymol blue stock solution was prepared 50 mg/ 75 ml with distilled water, and 4 ml of this stock solution was added for 100 ml the medium. After autoclaving, 7.8 g/L glucose, sucrose, maltose, mannitol and lactose were added in each medium separately with filter sterilization.

10 µl microorganism was added in 100 µl medium containing a sugar which was tested in a well. This microtiter plate was incubated at 25 °C for 3 days.

Each sugar was tested in duplicate. Positive control contained glucose as a carbon source, while negative control did not contain any carbon sources.

If the microorganism uses the sugar, the color of medium will change from green to yellow at the end of the incubation time as shown in Figure 2.7 and 2.8. (Kurtzman, Fell, Boekhout, & Robert, 2011; Ru, Bernal-grande, Cordero-bueso, & Hughes-herrera, 2017).



Figure 2.7. Carbohydrate Fermentation Test on Microplate Before Adding the Microorganisms (right to left: positive control, sucrose, maltose, mannitol, lactose, negative control)



Figure 2.8. Carbohydrate Fermentation Test on Microplate After 3 Days (right to left: positive control, sucrose, maltose, mannitol, lactose, negative control)

2.2.4. Characterization of Non-*Saccharomyces* and *Saccharomyces*

2.2.4.1. Ethanol Tolerance Test

In order to observe alcohol tolerance, firstly, the isolates were grown in the YPG broth for 24 hours at 25°C. After 24 hours, the amounts of cells in the broths were measured in the spectrometer at 600 NM. When OD reached 1 in spectrometer, it refers to 10^6 CFU/ml in the broths. After measuring, 0.1 ml of each having microorganism broth

was added to the YPG broth which contains different ethanol concentrations. These new YPG broths were prepared at 0, 10, 13 and 15% (v/v) ethanol concentrations respectively. Moreover, this test was occurred duplicate. After inoculation, these broths incubated for 3 days at 30°C (Guimarães et al, 2006).

2.2.4.2. Sulfur Dioxide Resistant Test

In order to observe sulfur dioxide resistance, firstly, the isolates were grown in the YPG broth for 24 hours at 25°C. After 24 hours, the amounts of cells in the broths were measured in the spectrometer at 600 NM. After reaching 10^6 CFU/ml in the broths, 0.1 ml of each having microorganism broth was added to the YPG broth which contains different concentration of SO₂. These concentrations are 0, 50, 100, 150, 200 mg/L, respectively. Moreover, this test was occurred duplicate. After inoculation, these broths incubated for 3 days at 30°C (Bağder, 2008).

2.2.4.3. Hydrogen Sulfide Production Test

The isolates were grown in the YPG broth for 24 hours at 25°C. After 24 hours, the samples in broths were inoculated on BIGGY (Bismuth Sulfite Glucose Glycine Yeast) agar (1 g/L yeast extract, 3 g/L sodium sulfite, 10 g/L glycine, 10 g/L dextrose, 5 g/L bismuth ammonium citrate, 16 g/L agar). After inoculation, these broths incubated for 3 days at 30°C (Vicente et al, 2006). The characteristics of H₂S production of the strains were determined according to the colors of the colonies formed on the BIGGY agar; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark-brown, 6: black (Sipiczki et al, 2001). In addition, *Candida* spp. was used for a positive control on the BIGGY agar.

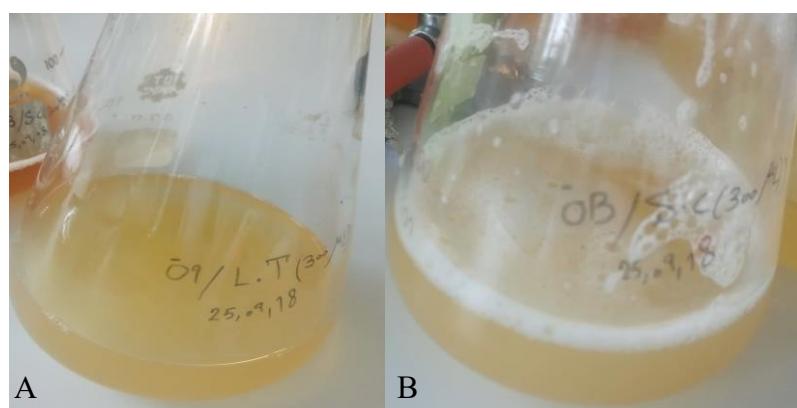
2.2.5. Sequential Inoculation of Non-*Saccharomyces* and *Saccharomyces cerevisiae* in the Wines

2.2.5.1. Sequential Inoculation of *Lachancea thermotolerans*, *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* in Emir Grapes

Emir grapes were taken from Cappadocia at 2018. These grapes were separated three parts for sequential inoculation our non-*Saccharomyces* and *Saccharomyces cerevisiae* strains, and pure inoculation of our *Saccharomyces cerevisiae* strain. They were added 30 mg/L SO₂ in order to kill all indigenous yeasts and molds. After 24 hours, selected yeasts were inoculated in these grapes.

For sequential inoculation, the microorganisms were grown in the YPG broth at 25°C. After reaching 10⁶ cells/ml, our *Lachancea thermotolerans* strain (OB 4.CM NS4) was inoculated for sequential inoculation with respect to the flowchart of white wine (Figure 2.4). After 7 days, our *Saccharomyces cerevisiae* strain (OB 4.CM S1) was added in this Emir must. (Maurizio Ciani et al., 2006). In addition, our *Wickerhamomyces anomalus* strain (KA 0.CM S1) was added with *Saccharomyces cerevisiae* strain in the same way.

For single *Saccharomyces cerevisiae* inoculation, our *Saccharomyces cerevisiae* strain (OB 4CM S1) was inoculated as a single culture after 24 hours of SO₂ treatment.



*Figure 2.9. YPG Broths with the Selected Yeasts (A: *Lachancea thermotolerans* strain (OB 4.CM NS4, B: *Saccharomyces cerevisiae* strain (OB 4CM S1))*

2.2.5.2. Inoculation of *Hanseniaspora uvarum* and *Saccharomyces cerevisiae* in Kalecik Karası Grapes

Kalecik Karası grapes were taken from Ankara at 2017. These grapes were separated into two parts for pure non-*Saccharomyces* (*Hanseniaspora uvarum*) and pure *Saccharomyces cerevisiae* inoculation. These two grapes were added 30 mg/L SO₂ in order to destroy all indigenous yeasts and molds. After 24 hours, selected yeasts were inoculated in these grapes.

The microorganisms were grown in the YPG broth at 25°C. After reaching 10⁶ cells/ml, our *Hanseniaspora uvarum* (Strain 1) and *Saccharomyces cerevisiae* (Strain 13) strains were added in Kalecik Karası musts, separately, with respect to the flowchart of red wine (Figure 2.3).

2.2.5.3. Inoculation of *Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae* *Lachancea thermotolerans* and *Saccharomyces cerevisiae* in Öküzgözü Grapes

Öküzgözü grapes were taken from Elazığ at 2018. These grapes were separated into four parts for sequential inoculation our non-*Saccharomyces* and *Saccharomyces cerevisiae* strains, and pure inoculation of our *Saccharomyces cerevisiae* strain (Table 2.2). They were added 30 mg/L SO₂ in order to kill all indigenous yeasts and molds. After 24 hours, selected yeasts were inoculated in these grapes.

For sequential inoculation, the microorganisms were grown in the YPG broth at 25°C. After reaching 10⁶ cells/ml, our *Lachancea thermotolerans* strain (OB 4.CM NS4) was inoculated for sequential inoculation with respect to the flowchart of red wine (Figure 2.3). After 4 days, *Saccharomyces cerevisiae* strain was added in this Öküzgözü must. (Maurizio Ciani et al., 2006). In addition, our *Hanseniaspora guilliermondii* (KA 0.CM NS1) and *Hanseniaspora opuntiae* (KA 0.CM NS2) strains were separately added with *Saccharomyces cerevisiae* strain in the same way.

For single *Saccharomyces cerevisiae* inoculation, our *Saccharomyces cerevisiae* strain (OB 4.CM S1) was inoculated after 24 hours.

Table 2.2. Inoculation of Microorganisms

<i>Microorganisms</i>	<i>Grapes</i>	<i>Inoculation Type</i>
<i>L. thermotolerans & S. cerevisiae</i>	Emir	Sequential Inoculation
<i>W. anomalus & S. cerevisiae</i>	Emir	Sequential Inoculation
<i>S. cerevisiae</i>	Emir	Pure Inoculation
<i>H. uvarum</i>	Kalecik Karası	Pure Inoculation
<i>S. cerevisiae</i>	Kalecik Karası	Pure Inoculation
<i>L. thermotolerans & S. cerevisiae</i>	Öküzgözü	Sequential Inoculation
<i>H. guilliermondii & S. cerevisiae</i>	Öküzgözü	Sequential Inoculation
<i>H. opuntiae & S. cerevisiae</i>	Öküzgözü	Sequential Inoculation
<i>S. cerevisiae</i>	Öküzgözü	Pure Inoculation

2.2.6. Wine Analysis

2.2.6.1. Determination of Alcohol, Acid, pH, Volatile Acid, Reducing Sugar and Malic Acid Content

Ethanol, total acidity, pH, volatile acid, reducing sugar and malic acid contents of wine samples were measured with Foss WineScan™ Auto Equipment (Type 79067, Denmark) at Kavaklıdere Winery A.Ş. WineScan™ composes of the analyzer and Foss Integrator software (Figure 2.10 and 2.11).

10 mL of wine samples was put in standard tubes of instrument and analysis of samples started by immersing the specific probe in each sample. Ethanol, total acidity, pH, volatile acid, reducing sugar and malic acid measurement of wine samples were obtained by Foss Integrator software.



Figure 2.10. Foss WineScan™ Auto Equipment

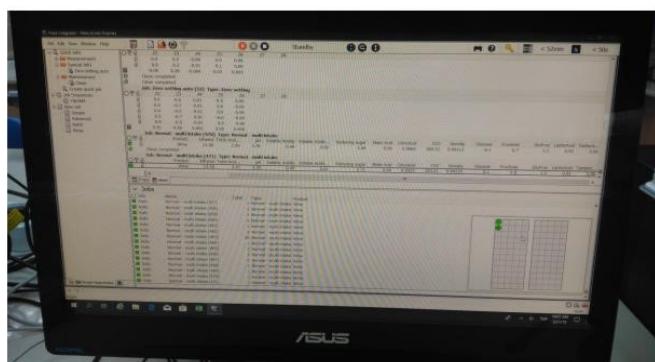


Figure 2.11. Foss Integrator Software

2.2.6.2. Determination of Free and Total Sulphur Dioxide

The amounts of free and total SO₂ were measured by Flastar 5000 Analyzer (Figure 2.12). There were 5 solution boxes in this analyzer. These boxes were dinitro benzoic acid, two buffer solution, diluted HCl, distilled water.

0.74 g sodium disulfide and 50 ml ethanol were mixed, and it covered 500 ml with distilled water to prepare mixture 1.

1, 2, 4, 8, 14 and 20 ml mixture 1 were covered 100 ml with distilled water in volumetric flasks to prepare 10, 20, 40, 80, 140, and 200% SO₂. These volumetric flasks were put in the machine with wine samples.

The program (“Sofia”) was opened and total SO₂ graph was drawn ($r=0.99$).

The references volumetric flashes of 140% and 200% SO₂ were taken off the machine due to measurement of free SO₂. Then, free SO₂ graph was drawn with this software ($r=0.99$). The results denominated as mg/L.

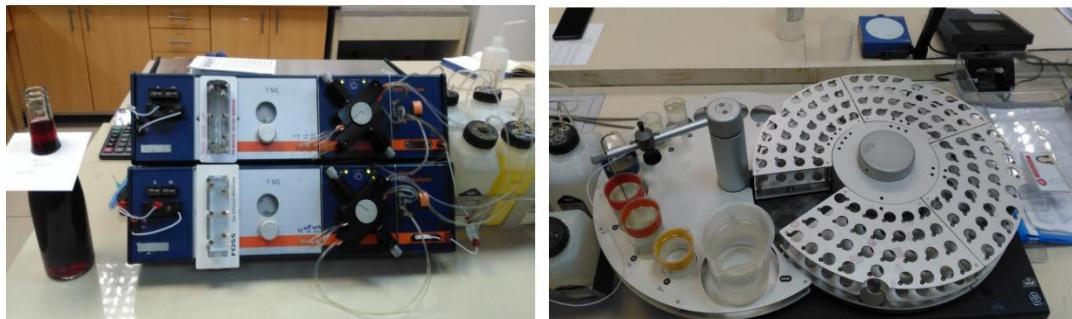


Figure 2.12. Flastar 5000 Analyzer for Determination of Free and Total SO₂

2.2.7. Volatile Compounds and Sensory Analysis of Wines

2.2.7.1. Volatile Compounds

Analysis of volatile compounds was carried out at Prof. Dr. Serkan Selli's Laboratory in Cukurova University.

2.2.7.1.1. Extraction of Volatile Compounds in Wine Samples

Volatile compounds of wine samples were obtained with liquid-liquid extraction method and defined with gas chromatography and mass spectrometry whose brand names are Agilent 6890 GC and 5973 MSD. Extraction was carried out by mixing 45 ml of the sample in a 500 ml Erlene with 40 µg of 4-nonal (internal standard) and 50 ml of a high purity dichloromethane solvent in a magnetic stirrer at 4-5 °C for 45 minutes under nitrogen gas. After extraction, the samples were centrifuged at 0°C 5500 rpm for 15 minutes. After centrifugation, water which may be present in dichloromethane and aromatic extract was separated by separatory funnel. It was removed with sodium sulfate and this mixture was concentrated to 1 mL at 40°C in a distillation column of Vigreux. The aromatic extracts were concentrated again to 0.5 ml under nitrogen gas and directly injected into GC-FID, GC-MS systems to

determine the flavoring agents. Extraction was done with three replications (Sellı et al., 2007) (Figure 2.13).

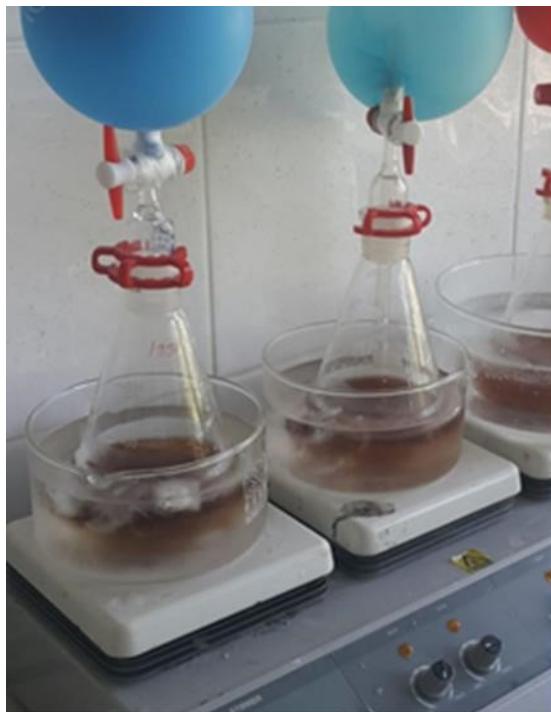


Figure 2.13. Extraction of Volatile Compounds in Wine Samples (Provided kindly by Prof. Dr. Serkan Sellı)

2.2.7.1.2. Identification and Quantification of Volatile Compounds

For identification of volatile compounds, Agilent 6890N gas chromatography and Agilent 5975B VL MSD mass spectrometry were used simultaneously. In addition, Agilent 6890N flame ionization detector (FID) gas chromatography was used for quantification of volatile compounds. The separation of the aromatics was carried out using DB-WAX capillary column (60 m x 0.25 mm x 0.4 μ m). The injector temperature was at 220°C, detector temperature was 250°C, column temperature was 60°C for 3 minutes. Then, column temperature was increased by 2°C per minute to 220°C and then 3°C per minute to 245°C. This temperature was remained constant for 20 minutes by a program. The amount injected into the device was 3 μ l. Helium was

used as the carrier gas and the flow rate was fixed at 1.5 ml/min. Temperatures of detector and injector were 250°C.

For identification of volatile compounds, Agilent 5975B VL MSD mass spectrometry was used. Injector type and temperature program had the same conditions as gas chromatography. The speed of helium used as carrier gas was 1.5 ml/min. The ionization energy of the mass spectrometer was screened at 70 eV, the ion source temperature was 250°C, the quadrupole temperature was 120°C, and it was scanned between 29-350 mass /load (m/e) at the intervals of one second. The peak diagnosis was made by comparing the mass spectrum for the non-standard compounds to the standard mass spectra in the libraries of the aroma substances in the computer memory (Wiley 9.0, NIST-11, and Flavor.2L) and for the standard solution to the standard of for the compounds. After the diagnosis of peaks, concentrations of flavoring substances were calculated according to the following formula according to the internal standard method (Sell et al., 2007). Each analysis was performed with 3 replications.

$$C_i = (A_i/A_{st}) \times C_{st} \times RF \times HF$$

C_i : Concentration of the compound

A_i : Peak area of the compound

A_{st} : Peak area of internal standard

C_{st} : Concentration of internal standard (40 µg/50 mL)

RF: Answer factor

HF: Calculation factor (factor for converting the sample amount to L).

2.2.7.2. Statistical Analysis

According to GS-MS results, statistical analyses of volatile compounds in wine were made with the help of MiniTab. The results were evaluated with ANOVA and Tukey's multiple range test. In these tests, the significant level was chosen $P < 0.05$.

2.2.7.3. Sensory Analysis of Wine

Sensory analyses of wines were realized with four degustators. In these analyses, there were 9 criteria: color, flavor, fullness, sweetness, acidness, bitterness, astringency, final astringency, and overall impression. Degustators were graded from 1 (weak) to 5 (strong) with respect to these criteria. Sensory profile was formed in radar charts by Excel Office 365.

CHAPTER 3

RESULTS AND DISCUSSION

3.1. Isolation of Microorganisms

469 microorganisms were obtained with the isolation method. It was thought that 204 of these yeast isolates belong to non-*Saccharomyces*, 265 belong to *Saccharomyces* yeasts. Figure 3.1 showed yeast colonies grown on selective media. Non-*Saccharomyces* yeast colonies were grown on lysine agar (A), YPG agar containing cycloheximide, biphenyl, and chloramphenicol (B) and *Saccharomyces* yeast colonies on the ethanol sulfate agar (C). Each colony was randomly picked from these specific agars (Figure 3.2) and streaked on YPG agar for purification (Figure 3.3.).

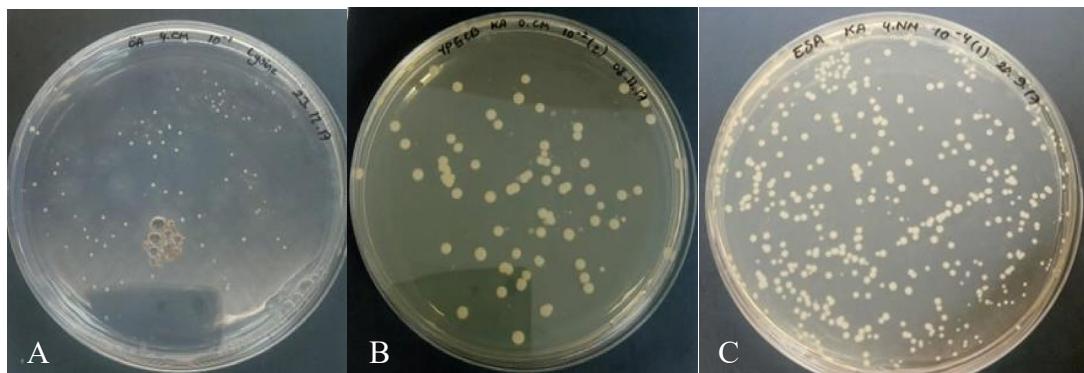


Figure 3.1. Yeast Colonies Grown on Selected Agars (A, lysine agar; B, YPG agar containing cycloheximide, biphenyl, and chloramphenicol; C, ESA)

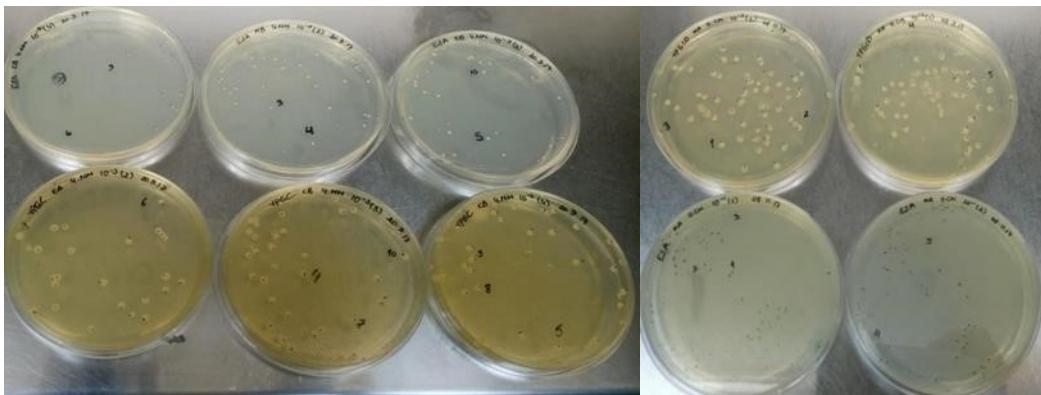


Figure 3.2. Selection and Isolation of Growing Microorganisms in Selective Agars



Figure 3.3. Purification Method of Selected Colonies on YPG Agar

469 microorganisms were obtained with the isolation method. It was thought that 204 of these yeast isolates belong to non-*Saccharomyces*, 265 belong to *Saccharomyces* yeasts (Figure 3.4). The number of isolated *Saccharomyces* and non-*Saccharomyces* yeasts from five different grape varieties of Turkey with using selective mediums (ESA, lysine agar and YPG agar containing cycloheximide, biphenyl, and chloramphenicol) were depicted in Figure 3.5.

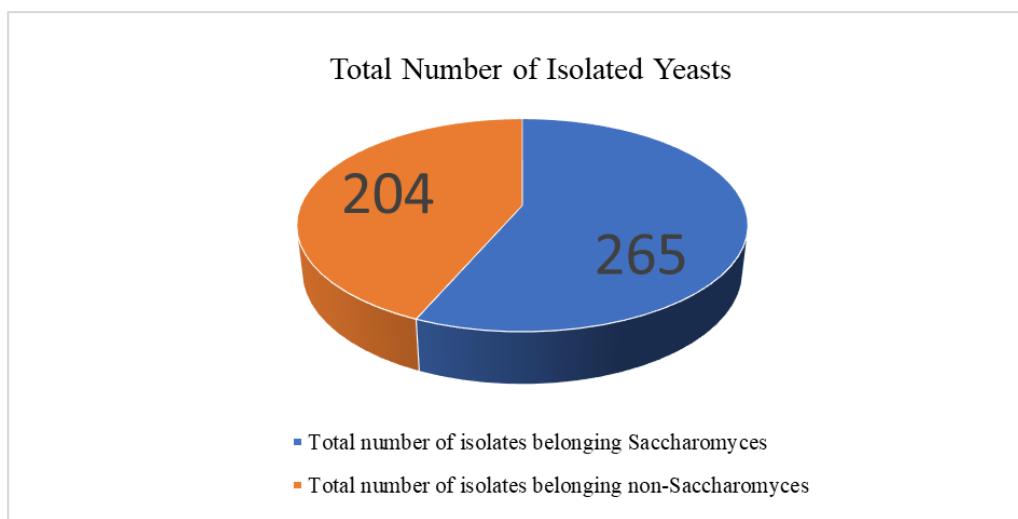


Figure 3.4. Total Number of Isolated Yeasts

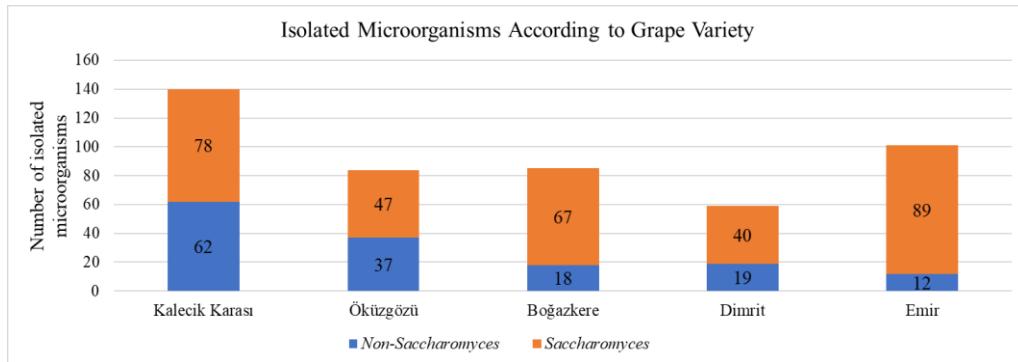


Figure 3.5. Numbers of Isolated Non-Saccharomyces and Saccharomyces Yeast According to Grape Variety

Total number of isolated microorganisms were 140 from Kalecik Karası grape. 60 were considered as non-*Saccharomyces*, while 80 as *Saccharomyces* according to isolates from selective agars (YPG agar containing cycloheximide, biphenyl, chloramphenicol, and ESA, respectively) (Figure 3.6.).

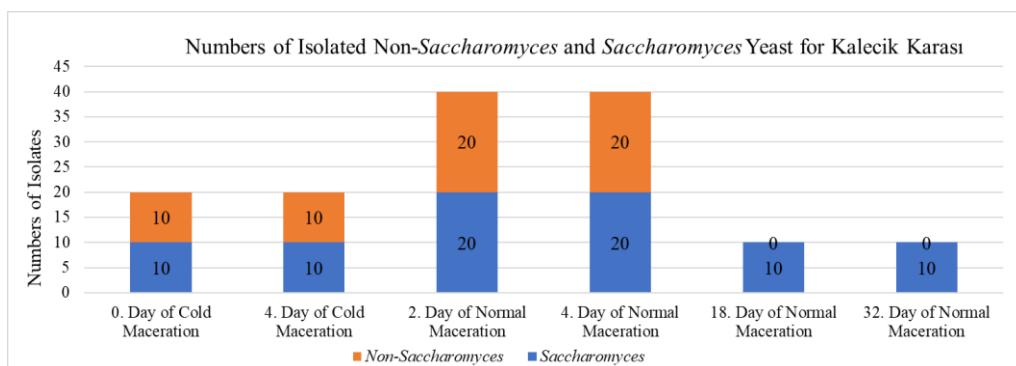


Figure 3.6. Numbers of Isolated Non-Saccharomyces and Saccharomyces Yeasts for Kalecik Karası Grape

Total number of isolated microorganisms were 84 from Öküzgözü grape. 40 were considered as non-Saccharomyces, while 44 as Saccharomyces yeasts according to isolates from lysine agar and ESA, respectively (Figure 3.7).

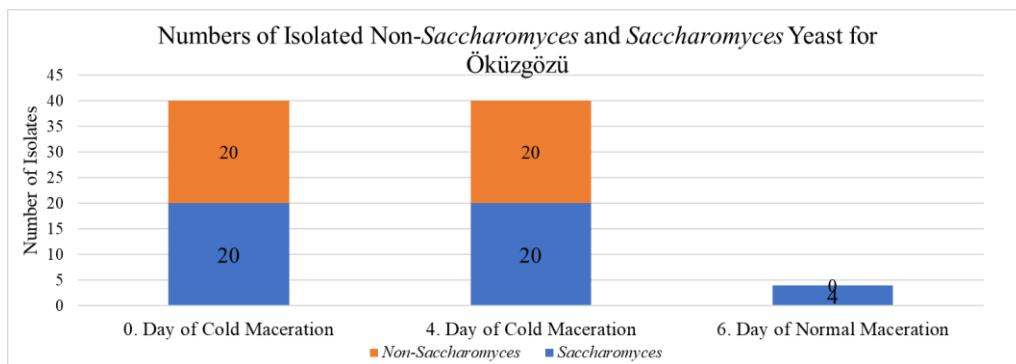


Figure 3.7. Numbers of Isolated Non-Saccharomyces and Saccharomyces Yeast for Öküzgözü Grape

Total number of isolated microorganisms were 85 from Boğazkere grape. 41 were considered as non-Saccharomyces, while 44 as Saccharomyces according to isolates from selective agars (lysine agar and ESA) as depicted in Figure 3.8.

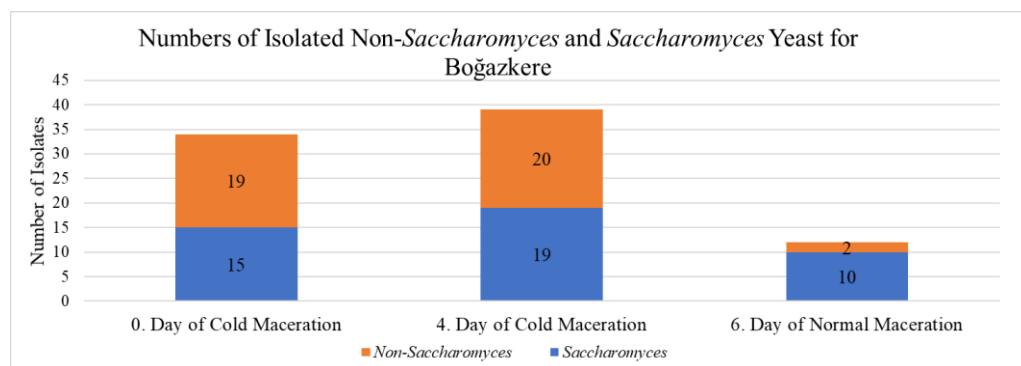


Figure 3.8. Numbers of Isolated Non-*Saccharomyces* and *Saccharomyces* Yeasts for Boğazkere Grape

Total number of isolated microorganisms were 59 from Dimrit grape. 19 were considered as non-*Saccharomyces*, while 40 as *Saccharomyces* according to isolates from selective agars (lysine agar and ESA) (Figure 3.9).

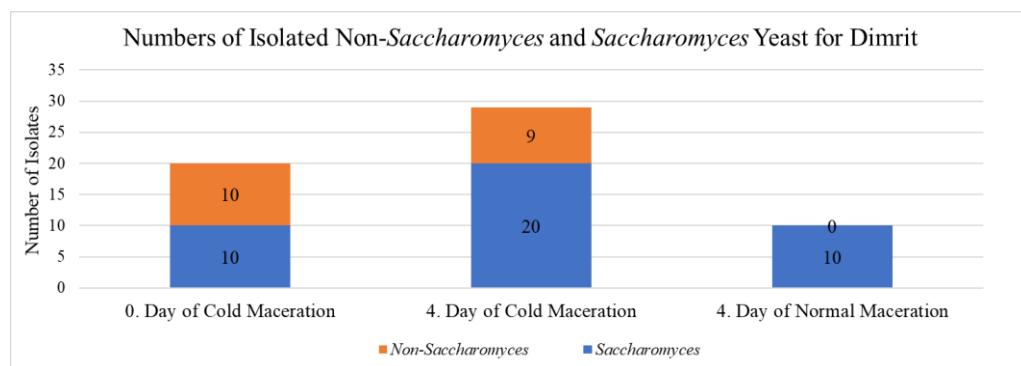


Figure 3.9. Numbers of Isolated Non-*Saccharomyces* and *Saccharomyces* Yeasts for Dimrit Grape

Total number of isolated microorganisms were 101 from Emir grape. 44 were considered as non-*Saccharomyces*, while 57 as *Saccharomyces* according to isolates from lysine agar and ESA, respectively (Figure 3.10).

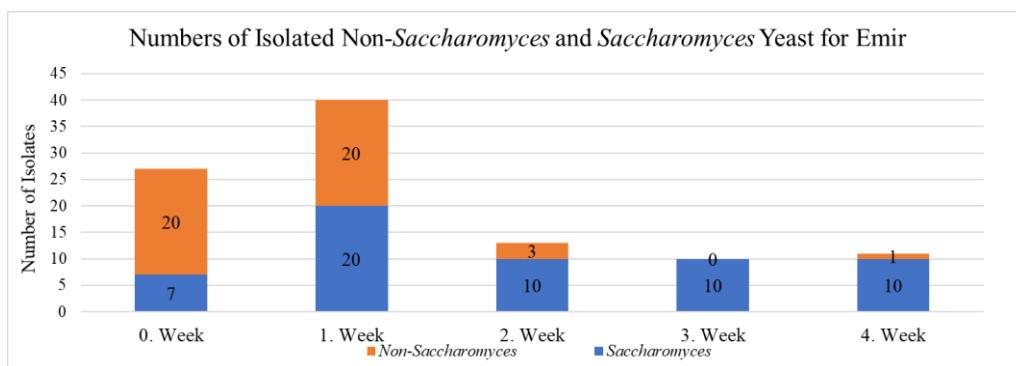


Figure 3.10. Numbers of Isolated Non-Saccharomyces and Saccharomyces Yeasts for Emir Grape

3.2. Phenotypic Characterization of *Saccharomyces* Yeasts

Firstly, H₂S production test was applied. Biggy agar was used to test of hydrogen sulfide production. Production of H₂S was indicated by the intensity of colours. Low H₂S production was observed for the isolate DA 4.NM S1 (Figure 3.11.A). Another isolate (OB 4.CM S2) from Öküzgözü grape produced intermediate H₂S amount (Figure 3.11.B). Moreover, the commercial *Saccharomyces cerevisiae* was used in order to compare strains. The level of H₂S production of this strain was intermediate (3) (Figure 3.11.C). On the other hand, *Candida* spp. was used as positive control, and the level of H₂S production is high (4) on Figure 3.11.C.

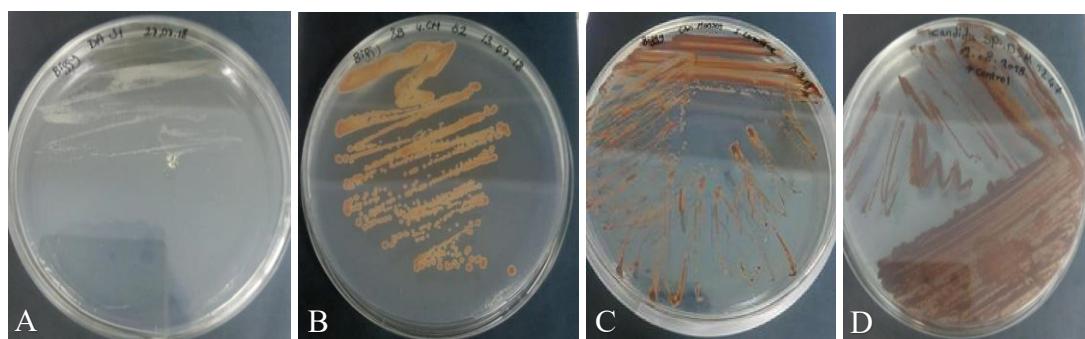


Figure 3.11. Biggy Agars (From left to right these isolates are given 1, 3, 3 and 4 of H₂S production.)

Secondly, test of alcohol tolerance was applied. The results of isolates from Dimrit were given as an example (Table 3.1). The other results of isolates from Kalecik Karası, Öküzgözü, Boğazkere and Emir were given in Appendix C.

Table 3.1. The Alcohol Tolerance and H₂S Production Results of *Saccharomyces* Isolates from Dimrit

Name	Alcohol Tolerance (%)				H ₂ S Production
	0	10	13	15	
DA 0CM S2	+	+	+	+	3
DA 0CM S3	+	+	+	+	3
DA 0CM S4	+	+	-	-	2
DA 0CM S5	+	+	w	w	2
DA 4CM S1	+	+	+	-	2
DA 4CM S2	+	+	+	w	2
DA 4CM S3	+	+	w	w	2
DA 4CM S4	+	+	+	+	2
DA 4CM S5	+	+	w	w	3
DA 4CM S6	+	w	-	-	2
DA 4CM S7	+	+	w	-	2
DA 4CM S8	+	+	+	w	2
DA 4CM S9	+	w	-	-	2
DA 4CMS10	+	w	-	-	ng
DA 4NM S1	+	+	+	w	1
DA 4NM S2	+	+	+	w	3
DA 4NM S3	+	+	-	-	3
DA 4NM S4	+	+	w	-	3
DB 0CM S1	+	w	w	-	3
DB 0CM S2	+	+	w	-	3
DB 0CM S3	+	+	w	w	4
DB 0CM S4	+	+	+	+	2
DB 0CM S5	+	+	+	w	3
DB 4CM S1	+	+	w	-	3
DB 4CM S2	+	+	-	-	3
DB 4CM S3	+	+	w	-	2
DB 4CM S4	+	+	w	w	3
DB 4CM S5	+	+	w	w	4
DB 4CM S6	+	+	w	w	3
DB 4CM S7	+	+	-	-	2
DB 4CM S8	+	+	w	w	4
DB 4CM S9	+	+	+	w	3
DB 4CMS10	+	+	+	-	3
DB 4NM S1	+	+	w	w	3
DB 4NM S2	+	+	w	w	3
DB 4NM S3	+	+	w	w	4
DB 4NM S4	+	-	-	-	ng
DB 4NM S5	+	+	+	w	3
DB 4NMS6	+	+	w	w	4

DA: A Parallel of Wine Made from Dimrit Grape, DB: B Parallel of Wine Made from Dimrit Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 4NM: 4. Day of Normal Maceration; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; ng: not growth.

According to H₂S production, some strains were eliminated. Strains with low level H₂S production were selected for the following tables (Table 3.2-3.6). However, isolates of Öküzgözü were chosen among intermediate level (3) H₂S producing strains due to the lack of low level (1 and/or 2) H₂S producing strains. Then, SO₂, temperature, pH tolerances, and carbohydrate fermentation tests were performed for these low level H₂S producing strains.

In addition, phenotypic properties of all *Saccharomyces* yeasts were given Appendix C.

The number of low H₂S producing *Saccharomyces* strains isolated from Kalecik Karası must and wine was 10 (Table 3.2).

*Table 3.2. Phenotypic Characterization and Carbohydrate Fermentation Test Results of *Saccharomyces* Strains Isolated from Kalecik Karası Must and Wine*

Name	Alcohol Tolerance (%)						SO ₂ Tolerance (mg/L)						Temperature Tolerance (°C)						pH Tolerance				H ₂ S Product
	0	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose					
KB 2NM S4	+	+	-	-	+	+	+	+	+	+	w	+	+	+	+	+	-	-	-	-	1		
KB 4NM S2	+	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	-	-	1		
KA 32NM S2	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	w	-	-	1		
KA 4NM S8	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	+	-	-	-	2		
KB 4NM S10	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	-	-	-	2		
KA 4NM S2	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	+	-	-	-	2		
KA 18NM S2	+	-	-	-	+	+	+	+	-	+	-	-	-	-	-	+	+	-	-	-	2		
KB 18NM S1	+	-	-	-	w	w	w	+	-	+	-	-	-	-	-	-	+	-	-	-	2		
KB 18NM S3	+	w	+	+	+	+	+	+	+	+	-	w	w	-	-	+	-	-	-	-	2		
KB 18NM S4	+	-	-	-	+	w	+	+	+	+	-	w	-	-	-	-	+	-	-	-	2		

KA: A Parallel of Wine Made from Kalecik Karası Grape, KB: B Parallel of Wine Made from Kalecik Karası Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 2NM: 2. Day of Normal Maceration, 4NM: 4. Day of Normal Maceration, 18NM: 18. Day of Normal Maceration (Middle of the fermentation), 32NM: 32. Day of Normal Maceration (End of the fermentation); S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black.

The only one strain isolated from Öküzgözü must and wine had low H₂S production (2). Therefore, isolates with intermediate level of H₂S production were added to the following table (Table 3.3). Moreover, the level of H₂S production of commercial *Saccharomyces* strain was also 3.

Table 3.3. Phenotypic Characterization and Carbohydrate Fermentation Test Results of *Saccharomyces* Strains Isolated from Öküzgözü Must and Wine

Name	Alcohol						Temperature				pH				Carbohydrate Fermentation Test				<i>H₂S</i> Product	
	SO ₂ Tolerance (mg/L)						Tolerance (°C)				Tolerance									
	0	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose		
OA 0CM NS3	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
OB 0CM S6	+	+	+	+	+	+	+	+	+	+	-	+	+	+	-	+	w	w	3	
OA 0CM S10	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	
OB 0CM S5	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	
OB 4CM S2	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	
OB 4CM S1	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	

OA: A Parallel of Wine Made from Öküzgözü Grape, OB: B Parallel of Wine Made from Öküzgözü Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black.

The low level of producing H₂S *Saccharomyces* strains isolated from Boğazkere and Dimrit musts and wines was represented in Table 3.4 and Table 3.5, respectively.

Table 3.4. Phenotypic Characterization and Carbohydrate Fermentation Test Results of *Saccharomyces* strains Isolated from Boğazkere Must and Wine

Name	Alcohol						Temperature				pH				Carbohydrate Fermentation				<i>H₂S</i> Product	
	SO ₂ Tolerance (mg/L)						Tolerance (°C)				Tolerance				Test					
	0	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose		
BA 0CM NS3	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
BA 0CM NS5	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
BB 0CM NS6	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
BA 4CM NS6	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
BB 4CM S10	+	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	2	

BA: A Parallel of Wine Made from Boğazkere Grape, BB: B Parallel of Wine Made from Boğazkere Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 6NM: 6. Day of Normal Maceration; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black.

Table 3.5. Phenotypic Characterization and Carbohydrate Fermentation Test Results of *Saccharomyces* Strains Isolated from Dimitrit Must and Wine

Name	Alcohol Tolerance(%)					SO ₂ Tolerance (mg/L)				Temperature Tolerance(°C)			pH Tolerance			Carbohydrate Fermentation Test				<i>H₂S</i> Production
	0	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose		
DA 4NM S1	+	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
DA 4CM S3	+	+	w	w	+	+	+	+	+	+	-	w	w	-	+	+	w	w	2	
DA 4CM S1	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	
DA 4CM S4	+	+	+	+	+	+	+	+	+	+	-	+	+	+	-	+	w	w	2	
DA 4CM S9	+	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
DA 4CM S2	+	+	+	w	+	+	+	+	+	w	-	w	w	-	-	+	w	-	2	
DA 4CM S8	+	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
DA 0CM S5	+	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
DA 0CM S4	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	2	
DA 4CM S7	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	
DA 4CM S6	+	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	2	
DB 4CM S7	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	
DB 0CM S4	+	+	+	+	+	+	+	+	+	w	+	+	+	-	+	-	-	-	2	
DB 4CM S3	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	

DA: A Parallel of Wine Made from Dimitrit Grape, DB: B Parallel of Wine Made from Dimitrit Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration 4NM: 4. Day of Normal Maceration; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black.

The number of low H₂S producing *Saccharomyces* strains isolated from Emir must and wine was 24 (Table 3.6). The highest number of low H₂S producing *Saccharomyces* isolates were obtained from Emir must and wine.

Table 3.6. Phenotypic Characterization and Carbohydrate Fermentation Test Results of *Saccharomyces* Strains Isolated from Emir Must and Wine

Name	Alcohol Tolerance(%)						SO ₂ Tolerance (mg/L)			Temperature Tolerance(°C)			pH Tolerance			Carbohydrate Fermentation Test				<i>H₂S</i> Product
	0	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose		
	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-		
EA 3W S4	+	+	-	-	+	+	+	+	+	+	-	+	+	+	-	-	w	w	1	
EB 3W S5	+	+	+	w	+	+	+	+	+	+	-	w	+	w	+	+	-	-	1	
EA 4W S5	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
EA 1W NS5	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
EA 1W NS6	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
EB 1W NS1	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
EB 1W NS7	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	
EA 0W NS5	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 0W NS1	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EA1WNS10	+	+	+	+	+	+	+	+	+	+	w	+	+	+	+	+	-	-	2	
EB 1W NS2	+	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 1W NS3	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 1W NS9	+	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EA 2W NS1	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EA 0W NS9	+	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EA 3W S2	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	
EA 3W S5	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 2W S5	+	+	+	w	+	+	+	+	+	+	-	+	+	+	-	-	w	w	2	
EB 3W S1	+	+	-	-	+	+	+	+	+	+	-	+	+	+	-	+	-	-	2	
EB 4W S1	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 0W S2	+	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 1W S3	+	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	
EB 1W S5	+	+	+	+	+	+	+	+	+	+	-	+	+	+	-	-	-	-	2	
EA 3W S3	+	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	-	2	

EA: A Parallel of Wine Made from Emir Grape, EB: B Parallel of Wine Made from Emir Grape; 0W: 0. Week, 1W: 1. Week, 2W: 2. Week; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black.

3.3. Identification of *Saccharomyces* Yeasts by DNA-Sequencing Methods

3.3.1. Identification of *Saccharomyces* Yeasts by Sequencing of ITS Region

According to phenotypic properties, selected *Saccharomyces* isolates were identified by sequencing of ITS1-5.8S-ITS2 regions.

The following figure (Figure 3.12) showed PCR products of isolated *Saccharomyces* strains amplified by ITS1 and ITS4 primers. The results of DNA-sequencing for all strains were given in Appendix A.



Figure 3.12. PCR Products of *Saccharomyces* Yeasts Amplified by ITS1 and ITS4 Primers. K40, *Hanseniaspora opuntiae*; K41-42, *Wickerhamomyces anomalus* (600 bp); K43, *Metschnikowia* spp.; K44-47, *Saccharomyces cerevisiae* (842 bp); K48-K50, *Saccharomyces cerevisiae* (K, isolates from Kalecik Karası grapes).

3.3.2. Identification of *Saccharomyces* Yeasts by Sequencing of D1/D2 Domain

Contradictory results were obtained for few isolates using the ITS and D1/D2 domain (Table 3.7).

Table 3.7. The Results of Sequencing with respect to ITS Region and D1/D2 Domain

Name	Region	Similarity	BLAST Result	Name	Region	Similarity	BLAST Result
K48	ITS	100%	<i>Saccharomyces</i> cf. <i>cerevisiae/paradoxus</i>	K48	D1/D2	100%	<i>Saccharomyces</i> Domain
B35	ITS	100%	<i>Saccharomyces</i> cf. <i>cerevisiae/paradoxus</i>	B35	D1/D2	100%	<i>Saccharomyces</i> Domain

Fernández-Espinar et al. (2000) has reported that ITS1-5.8S-ITS2 region showed greater interspecific differences than genes of 18S and 26S rRNA. Therefore, ITS region often allowed the differentiation of closely related species. Moreover, ITS region simplified the methodology due to the smaller size compared to D1/D2 domain. It was stated that this situation was also related to *Saccharomyces* genus (Fernandez-Espinar, Esteve-Zarzoso, Querol, & Barrio, 2000).

It was decided to determine the identity of these strains with further identification studies in the future.

3.4. High Technological Properties of *Saccharomyces* Strains

According to phenotypic properties, selected isolates of *Saccharomyces* were identified by DNA sequencing methods. Table 3.8 showed these selected isolates with high technological properties (high alcohol tolerance and low hydrogen sulfur production except OB 4 CM S1). OB 4 CM S1 was randomly chosen as *Saccharomyces cerevisiae* strain before phenotypic characterization. It was used for wine production. Although it produced relatively high H₂S (3) product, it has 13% alcohol resistance and high aroma quality was obtained in the wines produced with this microorganism.

Table 3.8. Selected Isolates of *Saccharomyces* with High Technological Properties

Name	Alcohol Tolerance (%)					SO ₂		Temperature			pH	Carbohydrate Fermentation Test					H ₂ S Production		Identification	
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2	
KA 32NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	1	Nd	+
KB 4NM S2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
KB 4NMS10	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
KA 4NM S2	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	Nd	+
EA 1W NS5	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
EA 1W NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
EB 1W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
EB 1W NS7	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
EB 3W S5	+	+	w	+	+	+	+	+	+	-	w	+	w	+	+	-	-	1	Nd	+
EA 0W NS5	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EB 0W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EA 1W NS10	+	+	+	+	+	+	+	+	+	w	+	+	+	+	+	-	-	2	+	Nd
EB 1W NS2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EB 1W NS3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EB 1W NS9	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EA 2W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
EA 0W NS9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
OA 0CM NS3	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
OB 4CM S1*	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	Nd	+
OB 4CM S2	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	Nd	Nd
BA 0CM NS3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
BA 0CM NS5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd
BB 4CM S10	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	2	Nd	+
BB 0CM NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
BA 4CM NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd
DA 4NM S1	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	Nd	+
DA 4CM S1	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	Nd	+
DA 4CM S3	+	w	w	+	+	+	+	+	+	-	w	w	-	+	+	w	w	2	Nd	+
DA 4CM S8	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	Nd	+
Commercial	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	Nd	+

KA: Kalecik Karası grape wines A parallel, KB: Kalecik Karası grape wines B parallel; EA: Emir grape wine A parallel, EB: Emir grape wine B parallel; OA: Öküzgözü grape wines A parallel, OB: Öküzgözü grape wines B Parallel; BA: Boğazkere grape wines A parallel, BB: Boğazkere grape wines B parallel; DA: Dimitri grape wines A Parallel, DB: Dimitri grape wines B parallel; 0CM: 0. day of cold maceration, 4CM: 4. day of cold maceration, 4NM: 4. day of normal maceration, 32NM: 32. day of normal maceration (end of the fermentation), 1W: 1. Week, 2W: 2. Week, 3W: 3. Week; S: *Saccharomyces*; +: positive growth , w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; Nd: not determined; *: Randomly chosen *Saccharomyces cerevisiae* was used for wine production, Commercial: Commercial *Saccharomyces cerevisiae* strain.

According to literature, *Saccharomyces cerevisiae* ferments glucose, sucrose and maltose as carbohydrate. On the other hand, it doesn't use mannitol and lactose (Kurtzman *et al.*, 2011.) Although Kurtzman et al. (2011) have reported that *Saccharomyces cerevisiae* ferments sucrose, some of our strains did not utilize sucrose. Moreover, they were identified as *Saccharomyces cerevisiae* by DNA sequencing method. On the other hand, although Guimarães isolated sucrose negative strains, they eliminated these strains (Guimaraes *et al.*, 2006). Similarly, Barnett et al. (2000) emphasized that sucrose, mannitol and maltose may vary from strain to strain. In the light of the contradictory results in the literature, sucrose negative isolates were not selected in Table 3.8 and, strains which have high alcohol tolerance (13% and above 13%) and low H₂S production (1 and 2) were chosen. Ethyl alcohol tolerance higher than 12-13% (v/v) is a desirable technological property for yeasts used as fermentation starters (Vaughan-Martini & Martini, 1998). 29 strains with high technological properties were shown in Table 3.8 and identified by ITS region and/or D1/D2 domain of rDNA sequencing. Therefore, the desired number of yeasts was reached to be used as starter cultures for winemaking.

In addition, isolates with the highest technological characteristics were selected from each type of grapes, and if possible, 2 isolates were selected from these grapes (Table 3.9). Only one isolate from Öküzgözü grape produced a low amount of H₂S and this isolate was selected from this grape variety. Grape varieties of the cultures of this grape, the region-specific characters and authentic features are added to the wine. In addition, it is thought that they will maintain the fermentation process more effectively by being superior to climate and environmental conditions (Jolly *et al.*, 2005). As a result, these isolates will be the strains which can be suggested for the wines to be prepared from the grapes.

The commercial *Saccharomyces cerevisiae* strain was added in Table 3.8 and 3.9 as a reference strain to compare the results.

Table 3.9. Selected High Technological Properties of *Saccharomyces* Strains

Name	Alcohol Tolerance (%)				SO ₂ Tolerance (mg/L)				Temperature Tolerance (°C)				pH Tolerance			Carbohydrate Fermentation Test			H ₂ S Production		Identification	
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	DI/D2			
KA 32NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	1	Nd	+		
KB 4NM S2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd		
EA 1W NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd		
EB 1W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd		
OA 0CM NS3	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	Nd		
BA 0CM NS3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	Nd		
BB 4CM S10	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	2	Nd	+		
DA 4NM S1	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	Nd	+		
DA 4CM S3	+	w	w	+	+	+	+	+	+	-	w	w	-	+	+	w	w	2	Nd	+		
Commercial	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	Nd	+		

KA: Kalecik Karası grape wines A parallel, KB: Kalecik Karası grape wines B parallel; EA: Emir grape wine A parallel, EB: Emir grape wine B parallel; OA: Öküzgözü grape wines A parallel, OB: Öküzgözü grape wines B Parallel; BA: Boğazkere grape wines A parallel, BB: Boğazkere grape wines B parallel; DA: Dimitrit grape wines A Parallel, DB: Dimitrit grape wines B parallel; 0CM: 0. day of cold maceration, 4CM: 4. day of cold maceration, 4NM: 4. day of normal maceration, 32NM: 32. day of normal maceration (end of the fermentation) 1W: 1. Week; S: *Saccharomyces*; +: positive growth, w: weak growth, -: negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; Nd: not determined; *: Randomly chosen *Saccharomyces cerevisiae* was used for wine production, Commercial: Commercial *Saccharomyces cerevisiae* strain.

3.5. Identification of Non-*Saccharomyces* Yeasts by DNA-Sequencing Methods

3.5.1. Identification of Non-*Saccharomyces* Yeasts by Sequencing of ITS Region

Isolated non-*Saccharomyces* yeasts can be identified by sequencing of ITS1-5.8S-ITS2 regions.

The following figures (Figure 3.13, 3.14, 3.15, 3.16) showed PCR products of isolated non-*Saccharomyces* strains amplified by ITS1 and ITS4 primers.

The results of DNA-sequencing for all non-*Saccharomyces* strains were given in Appendix A.

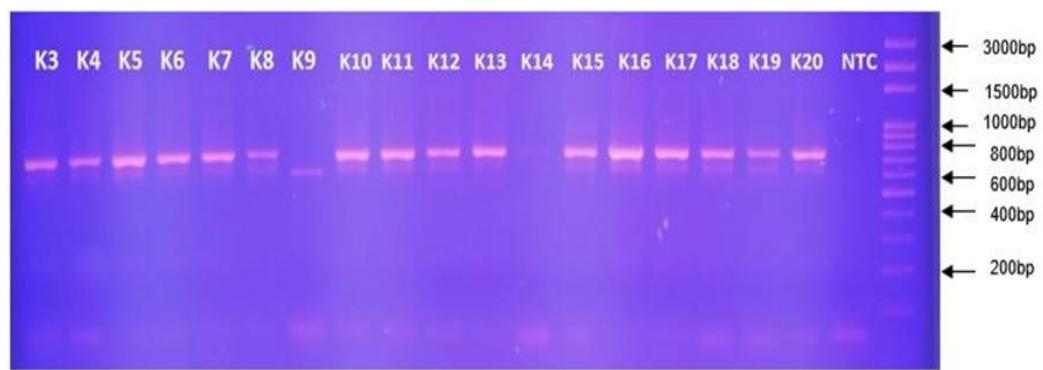


Figure 3.13. PCR Products of Non-*Saccharomyces* Yeasts Amplified by ITS1 and ITS4 Primers. K3, *Hanseniaspora uvarum* (747bp); K4, not identified; K5, *Hanseniaspora guilliermondii* (749 bp); K6, not identified; K7-8, *Hanseniaspora opuntiae*; K9, *Rhodotorula mucilaginosa* (610 bp); K10-12, *Hanseniaspora opuntiae*; K13, *Hanseniaspora uvarum* (747 bp); K14, not identified; K15-20, *Hanseniaspora opuntiae* (K, yeast isolates from Kalecik Karası grapes).

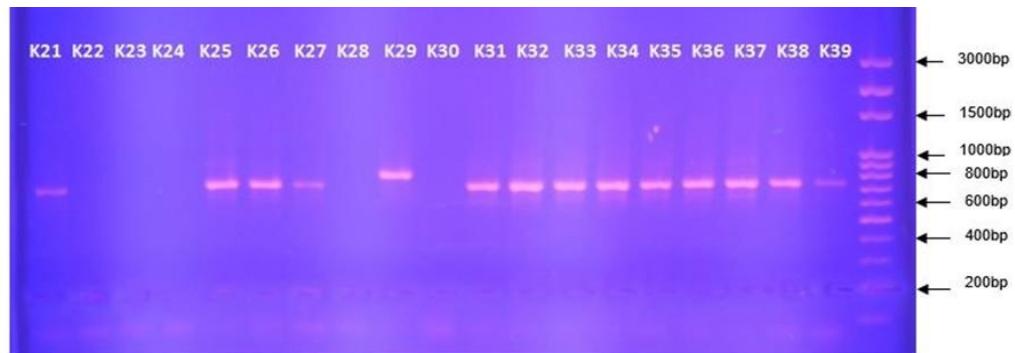


Figure 3.14. PCR Products of Non-*Saccharomyces* and *Saccharomyces* Yeasts Amplified by ITS1 and ITS4 Primers. K21, *Hanseniaspora opuntiae*; K22-24, not identified; K25-26, *Hanseniaspora uvarum* (747 bp); K27, *Hanseniaspora opuntiae*; K28, not identified; K29, *Saccharomyces cerevisiae* (842 bp); K30, not identified; K31, *Hanseniaspora guilliermondii* (749 bp); K32, *Hanseniaspora opuntiae*; K33, *Hanseniaspora guilliermondii* (749 bp); K34-35, *Hanseniaspora opuntiae*; K36, *Hanseniaspora guilliermondii* (749 bp); K37-38, *Hanseniaspora opuntiae*; K39, *Hanseniaspora guilliermondii* (749 bp). (K, yeast isolates from Kalecik Karası grapes)

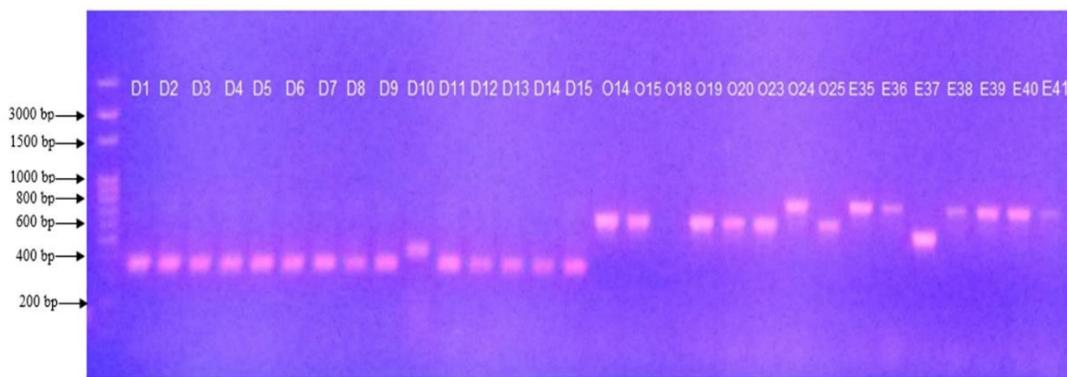


Figure 3.15. PCR Products of Non-*Saccharomyces* and *Saccharomyces* Yeasts Amplified by ITS1 and ITS4 Primers. D1, not identified; D2*, *Metschnikowia* spp.; D3*, *Metschnikowia sinensis*; D4, *Metschnikowia pulcherrima* (390 bp); D5, not identified; D6, *Metschnikowia chrysoperlae*; D7, not identified; D8*, *Metschnikowia* spp.; D9, not identified; D10, *Starmerella bacillaris* (475 bp); D11-14, *Metschnikowia pulcherrima*; D15*, *Metschnikowia* spp.; O14-23, *Lachancea thermotolerans*; O24, *Saccharomyces cerevisiae*; O25, *Lachancea thermotolerans* (675 bp); E35-36, *Saccharomyces cerevisiae* (842 bp); E37, *Wickerhamomyces anomalus* (600 bp); E38-41, *Saccharomyces cerevisiae* (D, yeast isolates from Dimrit grapes; O, yeast isolates from Öküzgözü grapes; E, yeast isolates from Emir grapes).

(*) These microorganisms are identified accurately according to D1/D2 Domain.

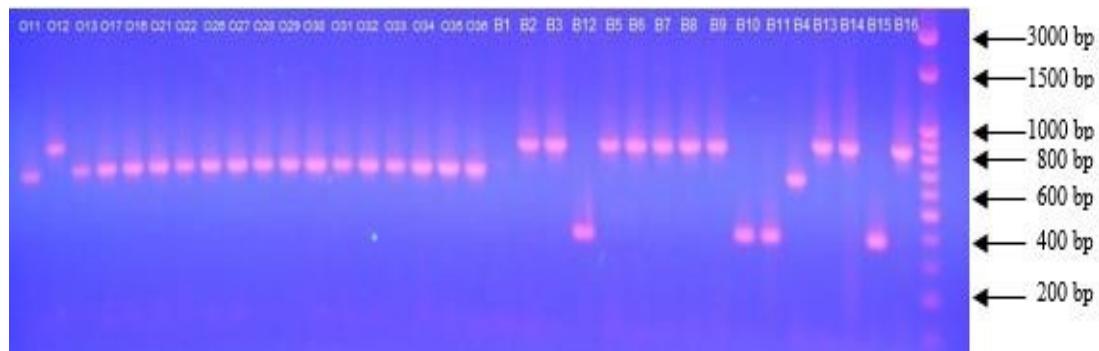


Figure 3.16. PCR Products of Non-*Saccharomyces* and *Saccharomyces* Yeasts Amplified by ITS1 and ITS4 Primers. O11, *Lachancea thermotolerans* (675 bp); O12, *Saccharomyces cerevisiae* (842 bp); O13-36, *Lachancea thermotolerans*; B1, not identified; B2-3, *Saccharomyces cerevisiae*; B12, *Metschnikowia pulcherrima* (390 bp); B5-9, *Saccharomyces cerevisiae*; B10, *Metschnikowia pulcherrima*; B11*, *Metschnikowia* spp.; B4, *Solicoccozyma aeria*; B13-14, *Saccharomyces cerevisiae*; B15, *Metschnikowia pulcherrima*; B16, *Saccharomyces cerevisiae* (O, yeast isolates from Öküzgözü grapes; B, yeast isolates from Boğazkere grapes).

(*) These microorganisms are identified accurately according to D1/D2 Domain.

Table 3.10 showed PCR amplicons of non-*Saccharomyces* and *Saccharomyces* yeasts by using ITS1 and ITS4 primers (Ghosh, 2017; Pham et al., 2011).

Table 3.10. The ITS1-5.8 rDNA-ITS2 PCR Amplicons of Yeasts (Ghosh, 2017; Pham et al., 2011)

<i>Yeasts</i>	<i>ITS1-5.8 rDNA-ITS2 PCR Amplicons (bp)</i>
<i>Metschnikowia pulcherrima</i>	390
<i>Metschnikowia aff. pulcherrima</i>	390
<i>Starmerella bacillaris</i>	475
<i>Wickerhamomyces anomalus</i>	600
<i>Rhodotorula mucilaginosa</i>	610
<i>Lachancea thermotolerans</i>	675
<i>Kluyveromyces marxianus</i>	720
<i>Kluyveromyces lactis</i>	740
<i>Hanseniaspora uvarum</i>	747
<i>Hanseniaspora guilliermondii</i>	749
<i>Saccharomyces cerevisiae</i>	842

According to the results of our PCR products, *Hanseniaspora uvarum* strains gave 747 bp while *Rhodotorula mucilaginosa* gave 610 bp in size (Figure 3.13). In addition, isolates of *Hanseniaspora* spp. utilized nearly the same band size in Figure 3.14. On the other hand, *Metschnikowia pulcherrima* gave 390 bp in size while *Starmerella bacillaris* utilized 475 bp in Figure 3.15. *Lachancea thermotolerans* isolates also gave 675 bp in Figure 3.16.

The result of our PCR products (bp) of non-*Saccharomyces* and *Saccharomyces* yeasts amplified by ITS1 and ITS4 primers were similar to amplicons (bp) of Ghosh (2017) as shown in Table 3.10.

3.5.2. Identification of Non-*Saccharomyces* Yeasts by Sequencing of D1/D2 Domain

Basic Local Alignment Search Tool (BLAST) results for identification of yeasts by ITS sequencing (Appendix A) were contradictory for few yeast isolates. Therefore, it was decided to carry out sequencing of 26/28S rDNA D1/D2 domain for these isolates. PCR amplicon of these isolates were displayed in Figure 3.17.

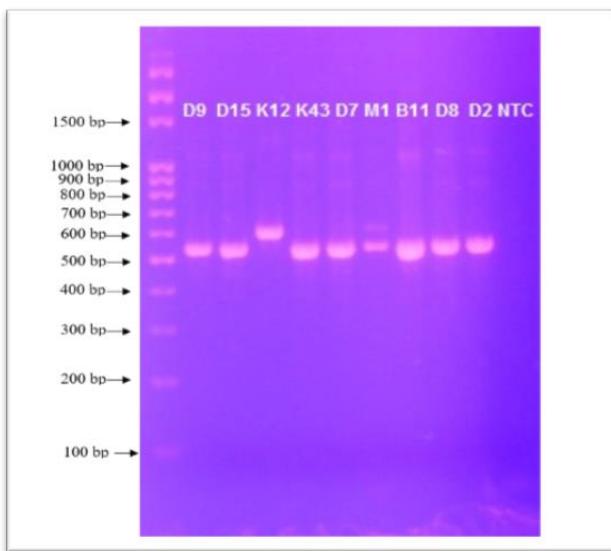


Figure 3.17. PCR Amplicon of Non-*Saccharomyces* Yeasts Amplified by NL1 and NL4 Primers. D9, not identified; D15, *Metschnikowia fructicola*; K12, *Hanseniaspora opuntiae*; K43, *Metschnikowia aff. fructicola*; D7, not identified; M1, *Metschnikowia aff. fructicola*; B11, *Metschnikowia aff. pulcherrima*; D8, *Metschnikowia pulcherrima*; D2, *Metschnikowia aff. pulcherrima*

These results of D1/D2 domain sequencing were compared with those obtained for ITS region sequencing. Contradictory results were obtained particularly for *Metschnikowia* spp. by two identification methods while the other yeast isolates gave the same identification results (Table 3.11). Table 3.12 showed also differences between yeasts identified by ITS region and D1/D2 domain sequencing.

Table 3.11. The Results of Sequencing with respect to ITS Region and D1/D2 Domain

Name	Region	Similarity	BLAST Result	Name	Region	Similarity	BLAST Result
K5	ITS	100%	<i>Hanseniaspora guilliermondii</i>	K5	D1/D2	100%	<i>Hanseniaspora guilliermondii</i>
K12	ITS	100%	<i>Hanseniaspora opuntiae</i>	K12	D1/D2	100%	<i>Hanseniaspora opuntiae</i>
K25	ITS	100%	<i>Hanseniaspora uvarum</i>	K25	D1/D2	100%	<i>Hanseniaspora uvarum</i>
K35	ITS	99%	<i>Hanseniaspora opuntiae</i>	K35	D1/D2	100%	<i>Hanseniaspora opuntiae</i>
K43	ITS		Not identified	K43	D1/D2	99%	<i>Metschnikowia aff. fructicola</i>
K62	ITS	100%	<i>Hanseniaspora opuntiae</i>	K62	D1/D2	100%	<i>Hanseniaspora opuntiae</i>
K65	ITS	99%	<i>Hanseniaspora opuntiae</i>	K65	D1/D2	99%	<i>Hanseniaspora opuntiae</i>
K70	ITS	98%	<i>Hanseniaspora uvarum</i>	K70	D1/D2	98%	<i>Hanseniaspora uvarum</i>
O1	ITS	99%	<i>Lachancea thermotolerans</i>	O1	D1/D2	99&	<i>Lachancea thermotolerans</i>
O7	ITS	100%	<i>Hanseniaspora opuntiae</i>	O7	D1/D2	100%	<i>Hanseniaspora opuntiae</i>
B4	ITS	100%	<i>Solicoccozyma aeria</i>	B4	D1/D2	99%	<i>Solicoccozyma aeria</i>
B11	ITS	98%	<i>Metschnikowia pulcherrima</i>	B11	D1/D2	99%	<i>Metschnikowia aff. pulcherrima</i>
B33	ITS	99%	<i>Metschnikowia pulcherrima</i>	B33	D1/D2	100%	<i>Metschnikowia aff. fructicola</i>
D2	ITS	100%	<i>Metschnikowia chrysoperlae</i>	D2	D1/D2	99%	<i>Metschnikowia aff. pulcherrima</i>
D3	ITS	94%	<i>Metschnikowia sinensis</i>	D3	D1/D2	99%	<i>Metschnikowia pulcherrima</i>
D8	ITS	95%	<i>Metschnikowia chrysoperlae</i>	D8	D1/D2	99%	<i>Metschnikowia pulcherrima</i>
D10	ITS	100%	<i>Starmerella bacillaris</i>	D10	D1/D2	100%	<i>Starmerella bacillaris</i>
D15	ITS	97%	<i>Metschnikowia aff. chrysoperlae</i>	D15	D1/D2	99%	<i>Metschnikowia fructicola</i>

K, Yeast isolates from Kalecik Karası; O, Yeast isolates from Öküzgözü; B, Yeast isolates from Boğazkere; D, Yeast isolates from Dimrit grapes.

Table 3.12. The Microorganisms Shown Different Results with respect to ITS Region and D1/D2 Domain Sequencing

Name	Region	Similarity	BLAST Result	Name	Region	Similarity	BLAST Result
K43	ITS		Not identified	K43	D1/D2 Domain	99%	<i>Metschnikowia aff. fructicola</i>
B11	ITS	98%	<i>Metschnikowia pulcherrima</i>	B11	D1/D2 Domain	99%	<i>Metschnikowia aff. pulcherrima</i>
B33	ITS	99%	<i>Metschnikowia pulcherrima</i>	B33	D1/D2 Domain	100%	<i>Metschnikowia aff. fructicola</i>
D2	ITS	100%	<i>Metschnikowia chrysoperlae</i>	D2	D1/D2 Domain	99%	<i>Metschnikowia aff. pulcherrima</i>
D3	ITS	94%	<i>Metschnikowia sinensis</i>	D3	D1/D2 Domain	99%	<i>Metschnikowia pulcherrima</i>
D8	ITS	95%	<i>Metschnikowia chrysoperlae</i>	D8	D1/D2 Domain	99%	<i>Metschnikowia pulcherrima</i>
D15	ITS	97%	<i>Metschnikowia aff. chrysoperlae</i>	D15	D1/D2 Domain	99%	<i>Metschnikowia fructicola</i>

K: Isolates from Kalecik Karası, B: Isolates from Boğazkere, D: Isolates from Dimrit.

Cordero-Bueso et al. and García et al. (2017) reported the difficulty of distinguishing *Metschnikowia fructicola* species from *Metschnikowia pulcherrima* species. Moreover, Hesham et al. at 2014 performed researches about the identification of biofuel yeast by sequencing of ITS region and D1/D2 domain. According to this study, the sequencing of D1/D2 domain of 26S ribosomal DNA was accepted as an accurate identification method for the unknown yeast species. Therefore, accurate results for our strains were also obtained by this method for non-Saccharomyces yeasts shown contradictory result respect to ITS region sequencing. Although K43 was not identified according to the ITS region sequencing, it was defined as *Metschnikowia aff. fructicola* by the sequencing of the D1/D2 domain.

Moreover, Boekhout et al. (2014) reported that PCR products of D1/D2 domain of yeast rDNA changing between 600 and 650 bp in size. In our study, *Hanseniaspora* spp. gave the same results within 600-650 bp in size, while *Metschnikowia* spp. gave different pattern with 520 bp in size (Figure 3.18) as Goldhawke et al (2016) mentioned in their studies (Figure 3.18).

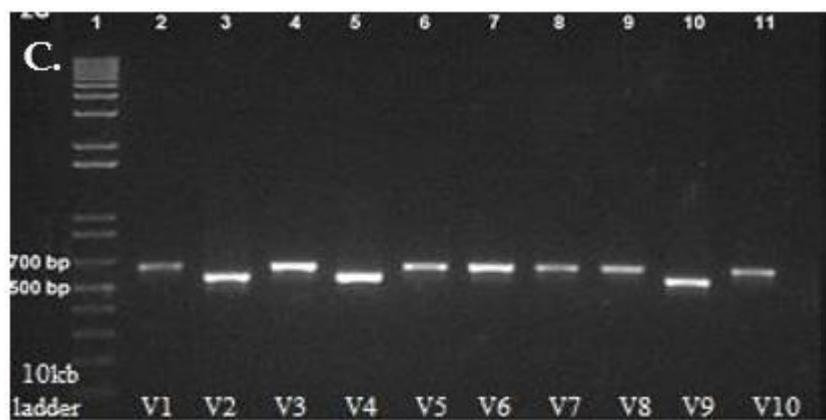


Figure 3.18. PCR Products of D1/D2 Domain. V1, V3, V5, V6, V7, V8, V10, *Hanseniaspora uvarum*; V2, V4, *Metschnikowia fructicola*; V9, *Metschnikowia* spp. (Goldhawke et al., 2016)

The D1/D2 nucleotide sequence of isolate (D3 strain) which was identified as *Metschnikowia sinensis* by ITS sequencing was compared with D1/D2 sequence of *Metschnikowia pulcherrima*. Only four nucleotide difference was observed according to alignment (Figure 3.19). However, ITS sequences of these two species were highly different (Figure 3.20). Therefore, further studies will be performed to clarify this confusing issue in the future.

Query 1	GGCAAAAGCTCAAATTGAAATCCCCGGAAATTGTAATTGAAGAGATTGGTCCGGC	60
Sbjct 13	GGCAAAAGCTCAAATTGAAATCCCCGGAAATTGTAATTGAAGAGATTGGTCCGGC	72
Query 61	CGGCGGGGTTAACACTGGAAAGTGGGCCACAGAGGGTGACAGCCCCGTGAACCCC	120
Sbjct 73	CGGCGGGGTTAACACTGGAAAGTGGGCCACAGAGGGTGACAGCCCCGTGAACCCC	132
Query 121	TTCAACGCCCTATCCAAATCTCAAGAGTCGAGTTGGGAATGCAGCTAAGTG	180
Sbjct 133	TTCAACGCCCTATCCAGATCTCAAGAGTCGAGTTGGGAATGCAGCTAAGTG	192
Query 181	GGTGGTAAATTCCATCTAAAGCTAAATACCGCGAGAGACCGATAGCGAACAGTACAGT	240
Sbjct 193	GGTGGTAAATTCCATCTAAAGCTAAATACCGCGAGAGACCGATAGCGAACAGTACAGT	252
Query 241	GATGGAAAGATGAAAAGCACTTGTAAAAGAGAGTGAAAAGTACGTGAAATTGTTGAAAG	300
Sbjct 253	GATGGAAAGATGAAAAGCACTTGTAAAAGAGAGTGAAAAGTACGTGAAATTGTTGAAAG	312
Query 301	GGAAGGGCTTGCAAGCAGACACTTAACACTGGCCAGCATGGGGCGGGGAAGCAAAACC	360
Sbjct 313	GGAAGGGCTTGCAAGCAGACACTTAACACTGGCCAGCATGGGGCGGGAGCAAAACC	372
Query 361	ACCGGGAAATGTACCTTCGAGGATTATAACCCCGTCCTTACTCCATGCCGCCCCGAG	420
Sbjct 373	ACCGGGAAATGTACCTTCGAGGATTATAACCCCGTCCTTACTCCATACCACCCGAG	432
Query 421	GCCTGCAATCTAAGGATGCTGGCTAATGGTGCAAGTCGCCGTCTGA	470
Sbjct 433	GCCTGCAATCTAAGGATGCTGGCTAATGGTGCAAGTCGCCGTCTGA	482

Figure 3.19. Alignment of D3 strain (Query) and *Metschnikowia pulcherrima* (Sbjct) according to D1/D2 Domain

Query 43	AAAAAACTTTAACACGGATCTTGGGCTCGCATCGATGAAAAACGCACCGAATTGC	102
Sbjct 243	AAAAAACTTTAACACGGATCTTGGGCTCGCATCGATGAAGAACGCAGCGAATTGC	184
Query 103	GATACGTAATATGACTTGAGACCGAATCATTGAATCTTGAACGCACATTGCCCG	162
Sbjct 183	GATACGTAATATGACTTGAGACGTGAATCATTGAATCTTGAACGCACATTGCCCG	124
Query 163	GGGTATTCAGGGGATGCGTGGGGAGCGATATTACTCTAAACCTCGGGTTGGG-	221
Sbjct 123	GGGTATTCAGGGCATGCGTGGGTGAGCGATATTACTCTAAACCTCTGGTTGGT	64
Query 222	CCTGCTTGGCTAATATCAACGGGCTNTAATAAGTTAGCCCCATTCTTTTC-TC	280
Sbjct 63	CCTGCTCAGGCTAATATCAACGGCGTAGAATAAGTTAGCCCCATTCTTTTCCTC	4
Query 281	ACC 283	
Sbjct 3	ACC 1	

Figure 3.20. Alignment of *Metschnikowia sinensis* (D3 strain)(Query) and *Metschnikowia pulcherrima* (Sbjct) according to ITS Region

Finally, 16 species were identified according to ITS region and/or D1/D2 domain sequencing results. 14 out of 16 yeast species were non-*Saccharomyces* as following: *Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae*, *Hanseniaspora uvarum*, *Rhodotorula mucilaginosa*, *Wickerhamomyces anomalus*, *Metschnikowia aff. fructicola*, *Metschnikowia fructicola*, *Lachancea thermotolerans*, *Solicoccozyma aeria*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima*, *Metschnikowia sinensis*, *Metschnikowia chrysoperlae*, *Starmerella bacillaris*. The others were *Saccharomyces cerevisiae* and *Saccharomyces cf. cerevisiae/paradoxus*. However, *Metschnikowia sinensis* was identified as *Metschnikowia pulcherrima* and *Saccharomyces cf. cerevisiae/paradoxus* as *Saccharomyces cerevisiae* by D1/D2 sequencing. The results of *Metschnikowia* spp. were given according to D1/D2 domain in Figure 3.21-3.25.

Hanseniaspora spp. (*Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae*, *Hanseniaspora uvarum*) was found predominant species in must of Kalecik Karası grapes. Moreover, *Rhodotorula mucilaginosa*, *Wickerhamomyces anomalus*, *Metschnikowia aff. fructicola* were also isolated and identified from Kalecik Karası grapes. Kalecik Karası had rich microflora according to isolated non-*Saccharomyces* yeast species. On the other hand, *Lachancea thermotolerans*, *Hanseniaspora opuntiae*, and *Wickerhamomyces anomalus* were isolated from Öküzgözü while *Solicoccozyma aeria*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima* were found in must of Boğazkere although Öküzgözü and Boğazkere grapes were taken from the same region (Elazığ). Emir grapes had less rich microflora than the others because *Hanseniaspora uvarum* and *Wickerhamomyces anomalus* were just isolated. On the other hand, *Metschnikowia* spp. and *Starmerella bacillaris* were isolated from Dimrit must although Emir and Dimrit grapes were taken from the same region (Cappadocia). In addition, *Metschnikowia* spp. (*Metschnikowia fructicola*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima*, *Metschnikowia chrysoperlae*) was found remarkably in Dimrit's must (Figure 3.21).

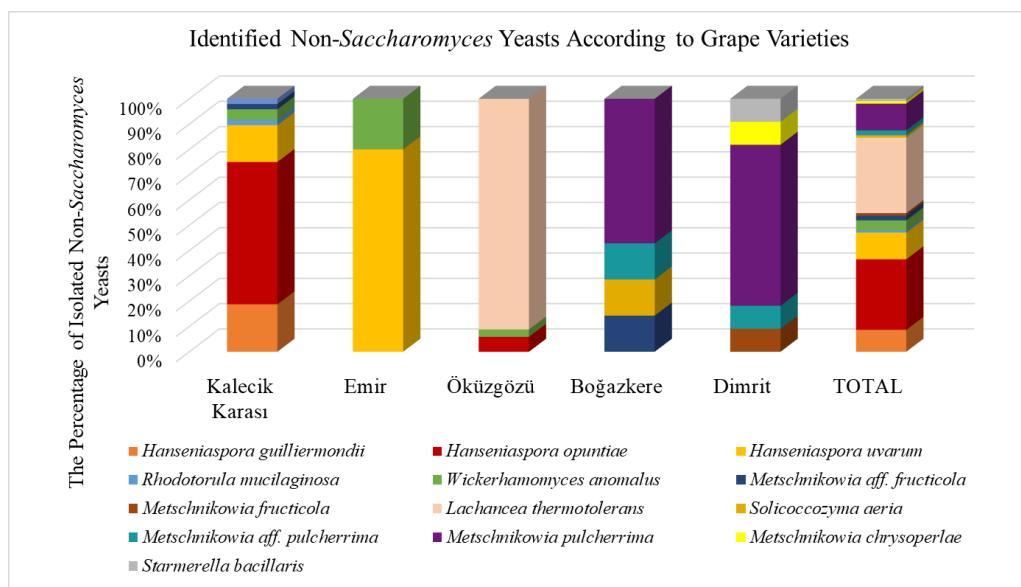


Figure 3.21. Identified Non-Saccharomyces Yeasts According to Grape Varieties

Hanseniaspora uvarum and *Hanseniaspora opuntiae* species were isolated from must and on the fourth day of cold maceration, the second and fourth day of normal maceration while *Hanseniaspora guilliermondii* were detected in the must and the fourth day of maceration. Moreover, *Rhodotorula mucilaginosa* and *Wickerhamomyces anomalus* species were only isolated from the must and *Metschnikowia aff. fructicola* during the fourth day of cold maceration of Kalecik Karası grapes (Figure 3.22).

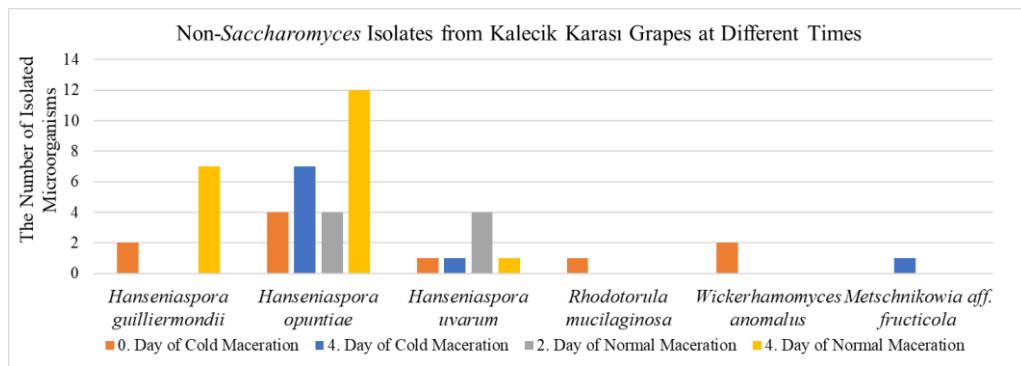


Figure 3.22. Non-Saccharomyces Yeast Isolates from Kalecik Karası Grapes at Different Time

Non-*Saccharomyces* yeasts were only isolated during cold maceration for Öküzgözü, Boğazkere and Dimrit musts. *Lachancea thermotolerans* can be obtained during cold maceration while *Hanseniaspora opuntiae* and *Wickerhamomyces anomalus* were isolated on the fourth day of cold maceration from the Öküzgözü must (Figure 3.23).

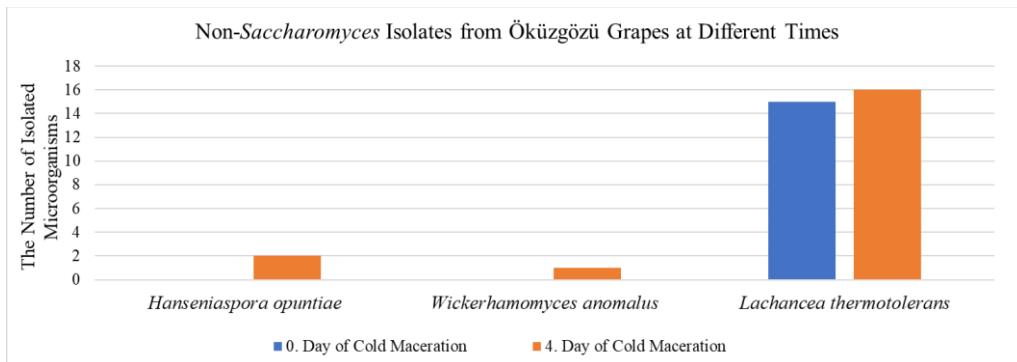


Figure 3.23. Non-*Saccharomyces* Yeast Isolates from Öküzgözü Grapes at Different Time

Metschnikowia pulcherrima species was isolated from must and during the fourth day of cold maceration while *Solicoccozyma aeria* and *Metschnikowia aff. pulcherrima* were only isolated from must grapes of Boğazkere. In addition, *Metschnikowia aff. fructicola* was isolated during the fourth day of cold maceration (Figure 3.24).

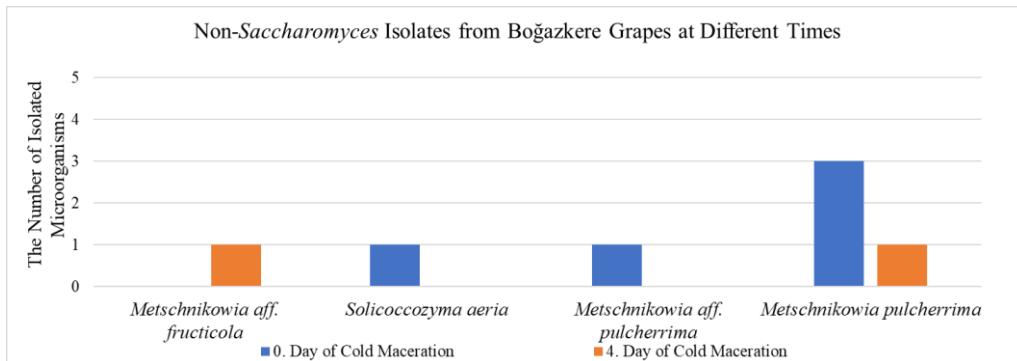


Figure 3.24. Non-*Saccharomyces* Yeast Isolates from Boğazkere Grapes at Different Time

Metschnikowia pulcherrima species were isolated from must and the fourth day of cold maceration of Dimrit grapes. *Metschnikowia aff. pulcherrima*, and *Metschnikowia chrysoperlae* were only isolated from must while *Starmerella bacillaris* during the fourth day of cold maceration (Figure 3.25).

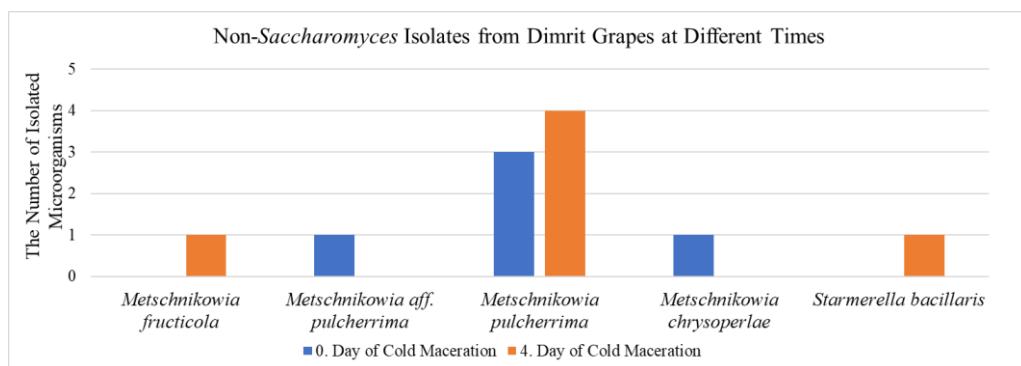


Figure 3.25. Non-Saccharomyces Yeast Isolates from Dimrit Grapes at Different Time

Wickerhamomyces anomalus was isolated after pressing of Emir grapes while *Hanseniaspora uvarum* species were only isolated at the end of first week of fermentation (Figure 3.26).

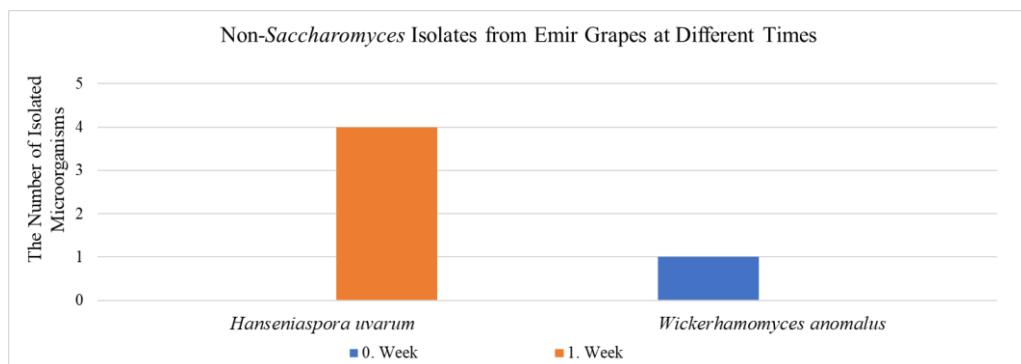


Figure 3.26. Non-Saccharomyces Yeast Isolates from Emir Grapes at Different Time

These microorganisms were generally isolated in cold and/or normal maceration times. Zott et al. (2008) also found non-Saccharomyces species, especially *Hanseniaspora uvarum* and *Starmerella bacillaris* at the end of the cold maceration and early stage of fermentation process. Moreover, Çelik et al. (2017) isolated 8 different non-Saccharomyces yeasts species (*Candida zemplinina*, *Hansenispora guilliermondii*, *Hanseniaspora uvarum*, *Metschnikowia* spp., *Lachancea thermotolerance*, *Pichia kluuyveri*, *Pichia occidentalis*, *Torulaspora delbrueckii*) at the early stage of fermentation in Narince grapes from the Tokat region. In addition,

Nurgel et al. (2005) isolated *Lachancea thermotolerance* and *Kloeckera apiculate* species at the beginning of the fermentation in Emir must. In our study, similarly, 14 non-*Saccharomyces* species were isolated during cold maceration and early stage of fermentation. It is a predictable result because these yeasts are dominant in early stage of fermentation. After increasing ethanol level, these yeasts die, and *Saccharomyces cerevisiae* is a dominant yeast species in the end of fermentation process (Urso et al., 2008).

397 out of 469 isolated microorganisms were identified by molecular or biochemical methods. 72 isolates were not identified due to failure to grow on the agar and/or failure to obtain the ITS results. The numbers of identified non-*Saccharomyces* and *Saccharomyces* yeasts were 104 and 293, respectively (Figure 3.27 and Figure 3.28). 104 non-*Saccharomyces* and 77 out of 293 *Saccharomyces* yeasts were identified by DNA sequencing of ITS region and/or D1/D2 domain. 59 out of 77 *Saccharomyces* strains were isolated from lysine medium. The growth of *Saccharomyces* strains on the Lysine medium has also been previously observed and reported (Ženíšová et al., 2014). 13 *Saccharomyces* strains were isolated from Ethanol Sulfate Agar (ESA), which is a specific agar for *Saccharomyces* (Kish et al., 1983; Heard and Fleet, 1985). The other *Saccharomyces* isolates (221) were also characterized according to biochemical tests and not sequenced due to their nonsuitability for phenotypic properties (low alcohol tolerance and high H₂S production). Finally, 29 *Saccharomyces* isolates were found with high technological properties used as a starter culture.

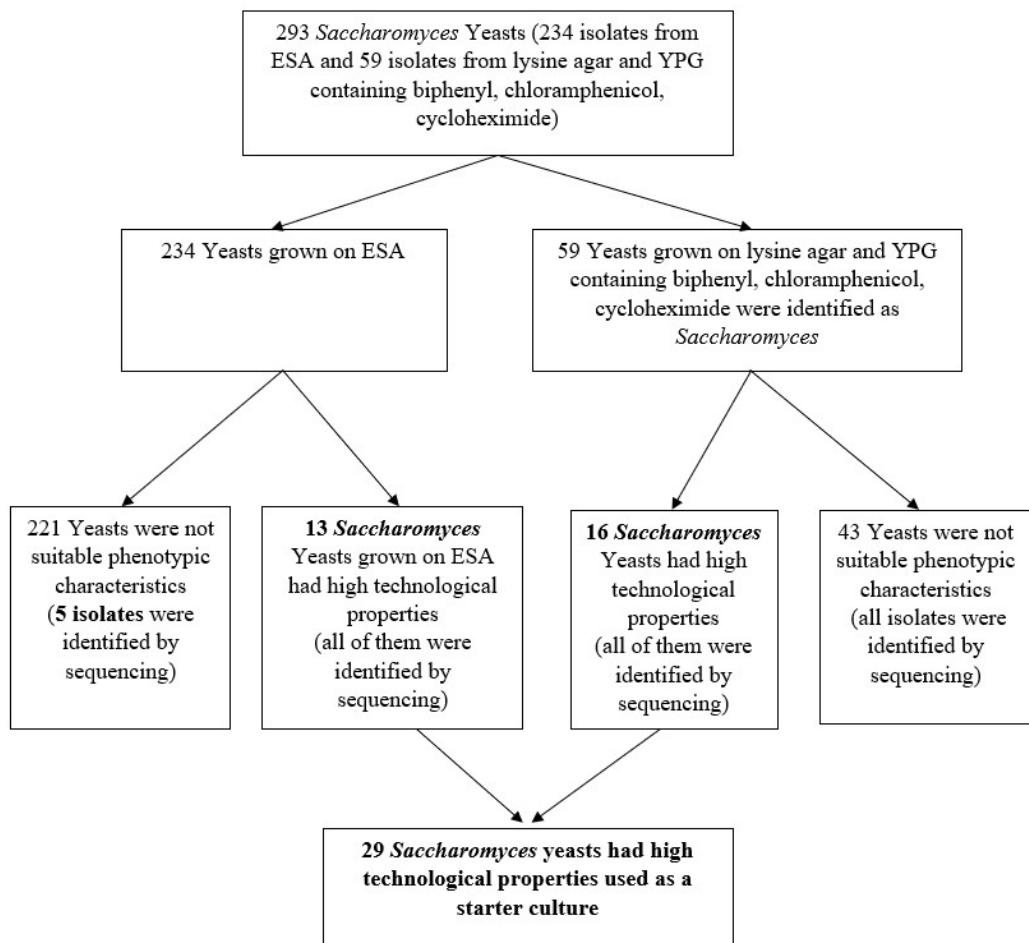


Figure 3.27. The Numbers of Isolated *Saccharomyces* Yeasts in The Culture Collection

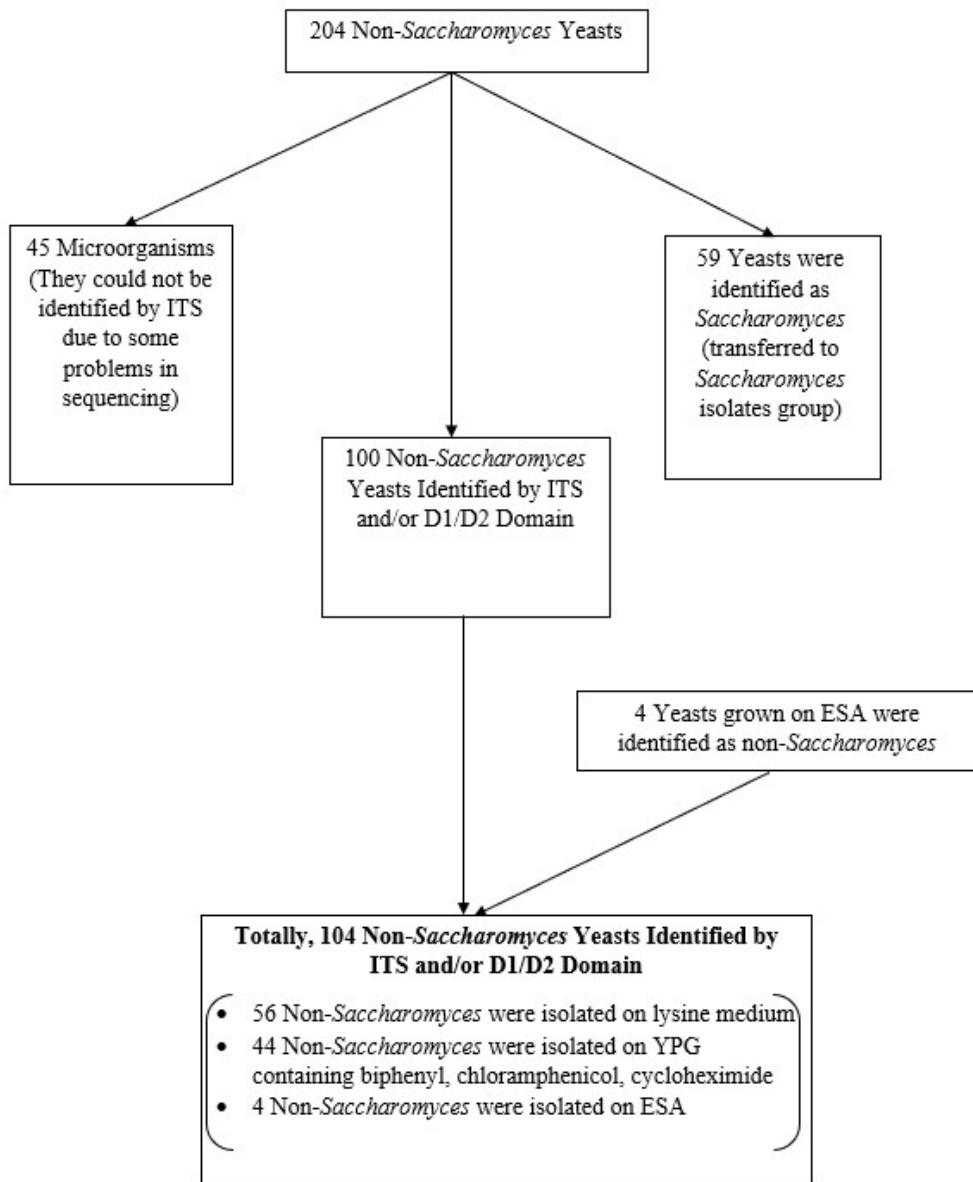


Figure 3.28. The Numbers of Isolated Non-Saccharomyces Yeasts in The Culture Collection

3.6. Phylogenetic Tree

After obtaining BLAST result, the sequences were aligned with ClustalW in MEGA X. After alignment, the phylogenetic trees were created with the Neighbor-Joining and UPGMA Method (unweighted pair group method with arithmetic mean). Distances of pairwise were calculated with the Tajima-Nei Model. The number of bootstrap replications was 100 iterations.

The evolutionary relationships of taxa using Neighbour-Joning Method were shown the following figures (Figure 3.29-3.36) while the phylogenetic trees which created with UPGMA Method were shown in Appendix B.

Phylogenetic trees of each grape varieties were firstly created. After these trees, different species were selected in these phylogenetic trees and then, the other phylogenetic tree with respect to different yeast species and different grape varieties was generated.

According to DNA sequencing results with ITS region, 4 different genera (*Rhodotorula*, *Wickerhamomyces*, *Hanseniaspora* and *Saccharomyces*) were identified in Kalecik Karası's must and wine. One of them was *Rhodotorula mucilaginosa*. 2 strains were defined as *Wickerhamomyces anomalus*. Moreover, 25 strains were found as *Hanseniaspora opuntiae*, while 7 strains were identified as *Hanseniaspora uvarum* and 9 strains were defined as *Hanseniaspora guilliermondii*. In addition, 8 of 53 microorganisms were belonged to *Saccharomyces cerevisiae*. The evolutionary relationship of these isolates was shown in the following phylogenetic tree (Figure 3.29).

It was interesting to observe that clusters of *Hanseniaspora* spp. were not closely related. However, *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* clusters were more closely related to one of those *Hanseniaspora* clusters than the other *Hanseniaspora* clusters.

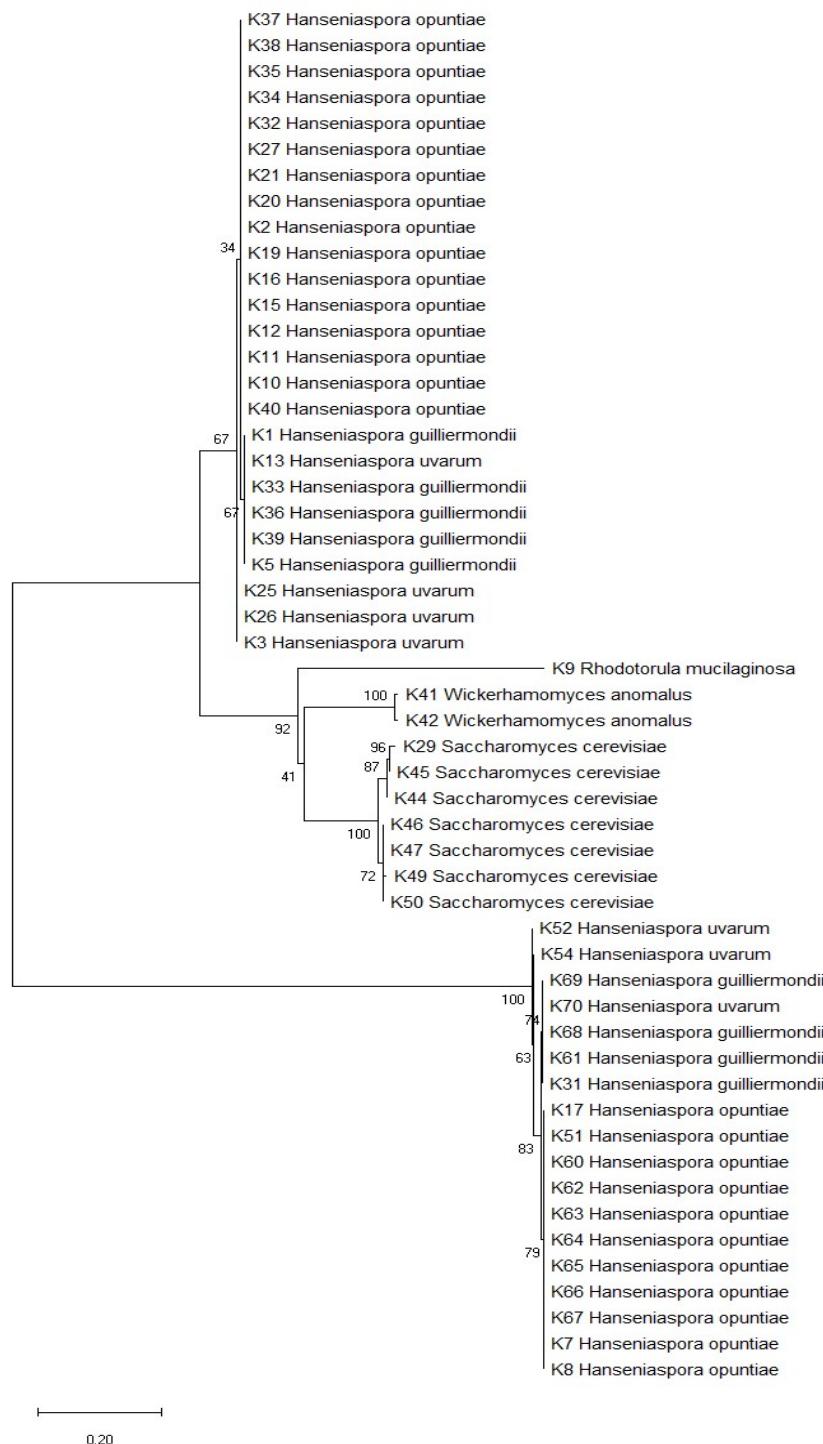


Figure 3.29. Phylogenetic Tree of Non-Saccharomyces and Saccharomyces Yeast in Kalecik Karası Must and Wine with respect to ITS Region. The scale bar indicates substitutions per base pair.

According to DNA sequencing results with ITS region, 3 different genera (*Lachancea*, *Hanseniaspora* and *Saccharomyces*) were identified in Öküzgözü's must and wine. 31 of 36 strains were defined as *Lachancea thermotolerans*. 2 strains were found as *Hanseniaspora opuntiae*, and 3 strains were identified as *Saccharomyces cerevisiae*.

The evolutionary relationship of these isolates was shown in the following phylogenetic tree (Figure 3.30).

It was interesting to observe that *Lachancea thermotolerance* clusters were not closely related to each other. However, *Hanseniaspora opuntia* and *Saccharomyces cerevisiae* clusters were more closely related to one of those *Lachancea* clusters than the other *Lachancea* clusters. According to the alignment of these two *Lachancea* clusters, it was found over 300 base pair differences. On the other hand, PCR products of these strains gave 675 bp in size. It was shown in Figure 3.15-3.16. It also referred to these strains were *Lachancea thermotolerance* (Table 3.10). In addition, *Kluyveromyces marxianus* and *Kluyveromyces lactis* gave 720 bp and 740 bp in size, respectively (Pham et al., 2011). *Lachancea (Kluyveromyces) thermotolerance* gave a more different base pair than *Kluyveromyces marxianus* and *Kluyveromyces lactis* with respect to the size of PCR product although they were the same genus.

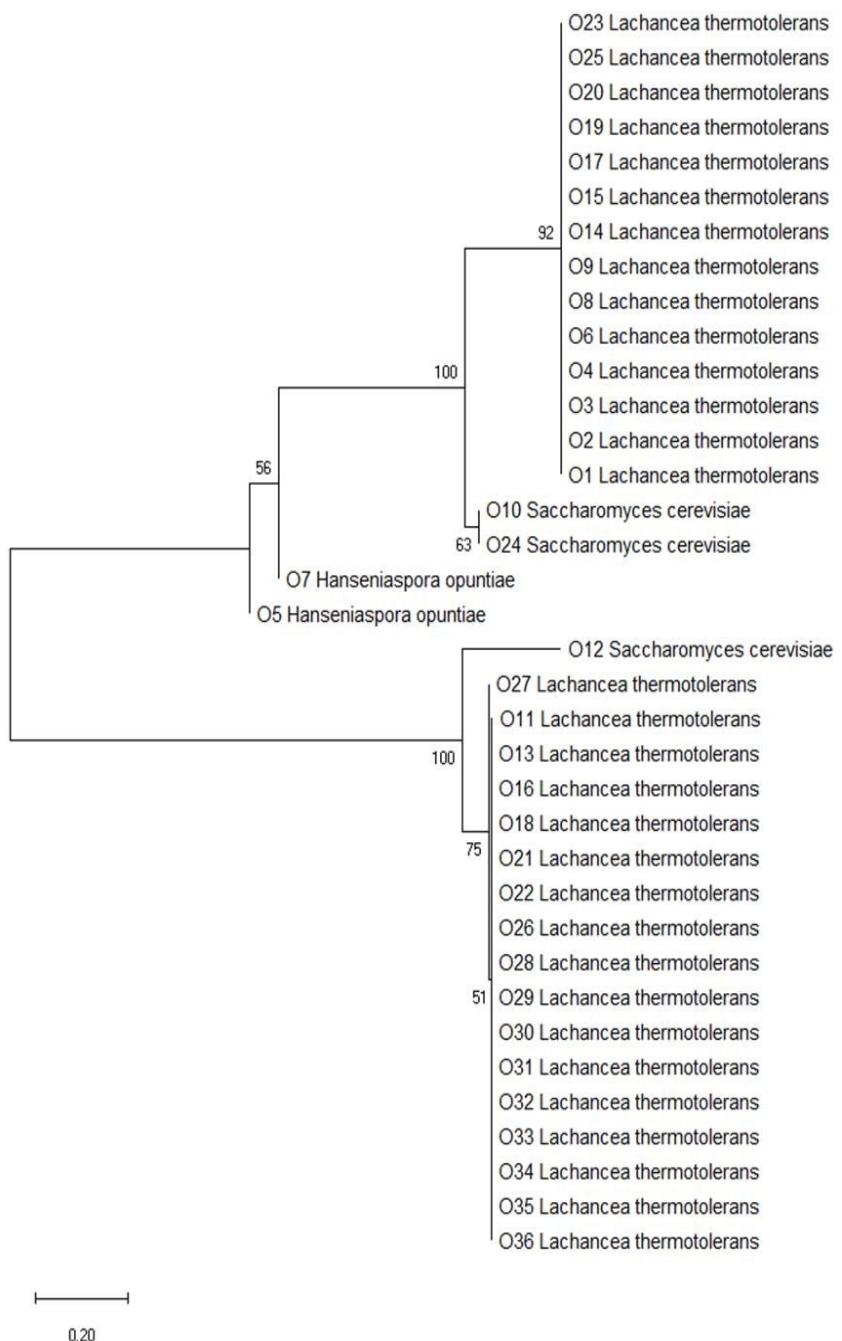


Figure 3.30. Phylogenetic Tree of Non-*Saccharomyces* and *Saccharomyces* Yeast in Öküzgözü Must and Wine with respect to ITS Region. The scale bar indicates substitutions per base pair.

According to DNA sequencing results with ITS region, 3 different genera (*Metschnikowia*, *Solicoccozyma* and *Saccharomyces*) were identified in Boğazkere's must and wine. 5 of them were found as *Metschnikowia pulcherrima* while only one strain was defined as *Solicoccozyma aeria*. In addition, 23 of 28 microorganisms were belonged to *Saccharomyces cerevisiae*. The evolutionary relationship of these isolates was shown in the following phylogenetic tree (Figure 3.31).

Interestingly, one cluster of *Saccharomyces cerevisiae* was not closely related to other *Saccharomyces cerevisiae* clusters due to over 300 base pair differences.

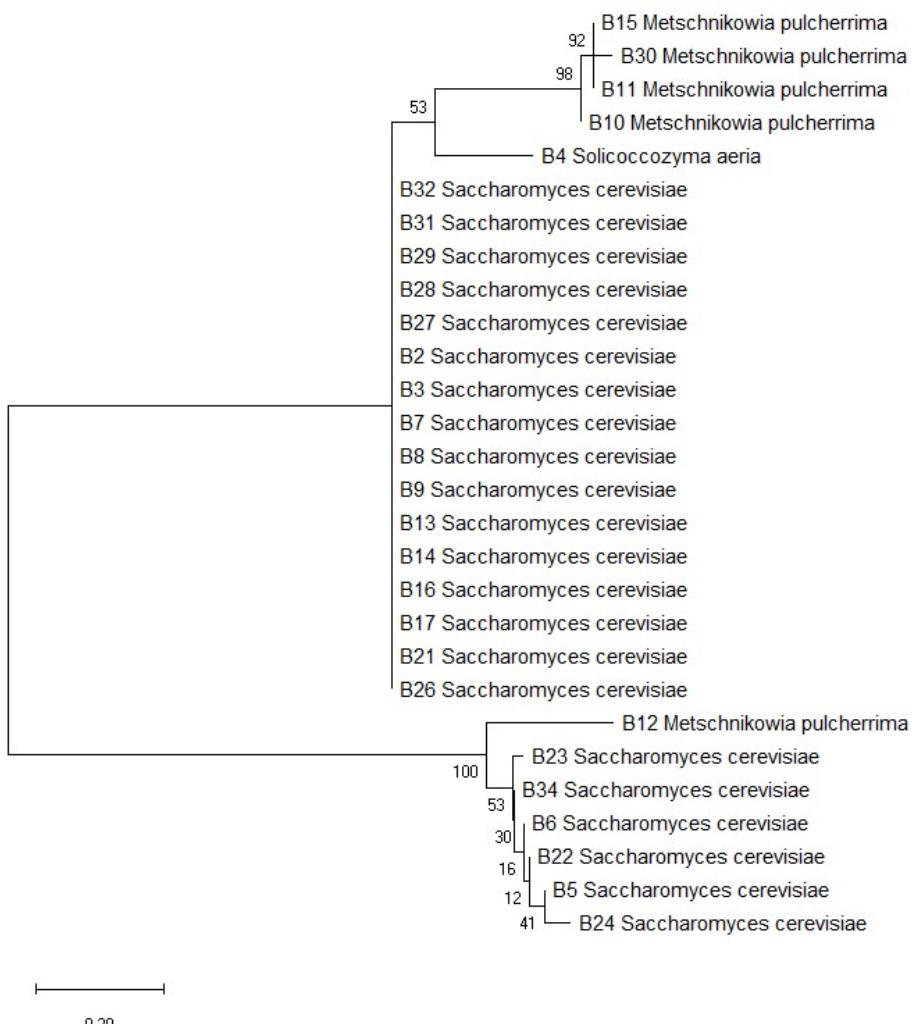


Figure 3.31. Phylogenetic Tree of Non-*Saccharomyces* and *Saccharomyces* Yeast in Boğazkere Must and Wine with respect to ITS Region. The scale bar indicates substitutions per base pair.

According to DNA sequencing results with ITS region, 2 different genera (*Metschnikowia* and *Starmerella*) were identified in Dimrit's must and wine. 5 of them were found as *Metschnikowia pulcherrima* while only one strain was defined as *Metschnikowia chrysoperlae*. In addition, one of them was defined as *Starmerella bacillaris*.

The evolutionary relationship of these isolates was shown in the following phylogenetic tree (Figure 3.32).

As it was expected, *Starmerella bacillaris* was not closely related to *Metschnikowia* spp. while *Metschnikowia pulcherrima* and *Metschnikowia chrysoperlae* had closely related each other.

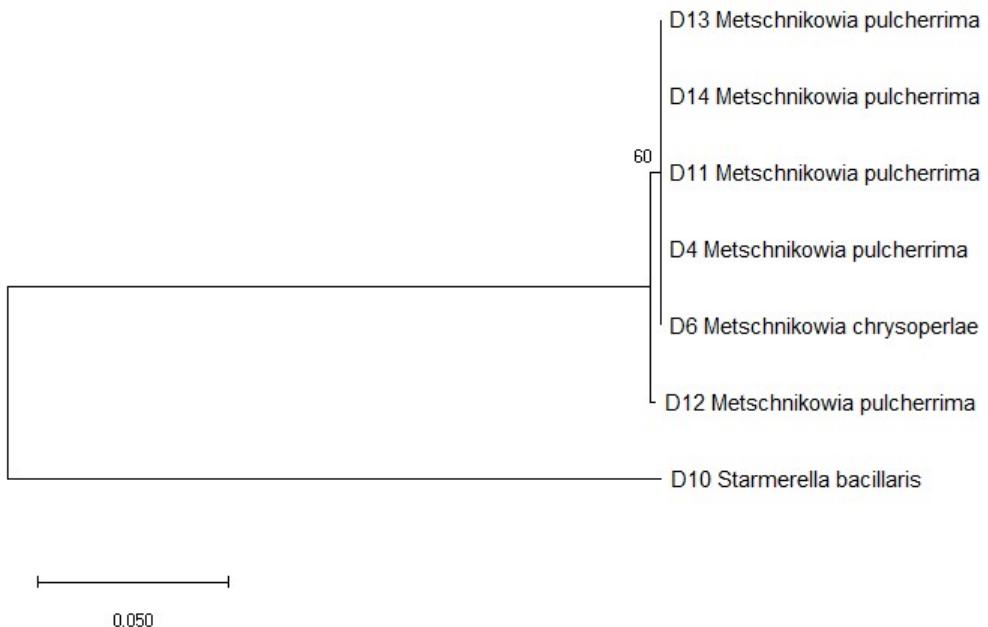


Figure 3.32. Phylogenetic Tree of Non-Saccharomyces and Saccharomyces Yeast in Dimrit Must and Wine with respect to ITS Region. The scale bar indicates substitutions per base pair.

According to DNA sequencing results with ITS region, 3 different genera (*Wickerhamomyces*, *Hanseniaspora* and *Saccharomyces*) were identified in Emir's must and wine. The one of them was *Wickerhamomyces anomalus*. Moreover, 4 strains were found as *Hanseniaspora uvarum*. In addition, rest of them belonged to *Saccharomyces cerevisiae*.

The evolutionary relationship of these isolates was shown in the following phylogenetic tree (Figure 3.33).

Similar unexpected results for members of some species were also noticed. *Saccharomyces cerevisiae* isolates formed two groups which were not closely related due to differences in their base pair alignments. On the other hand, *Saccharomyces* strains within the clusters showed very high similarity.

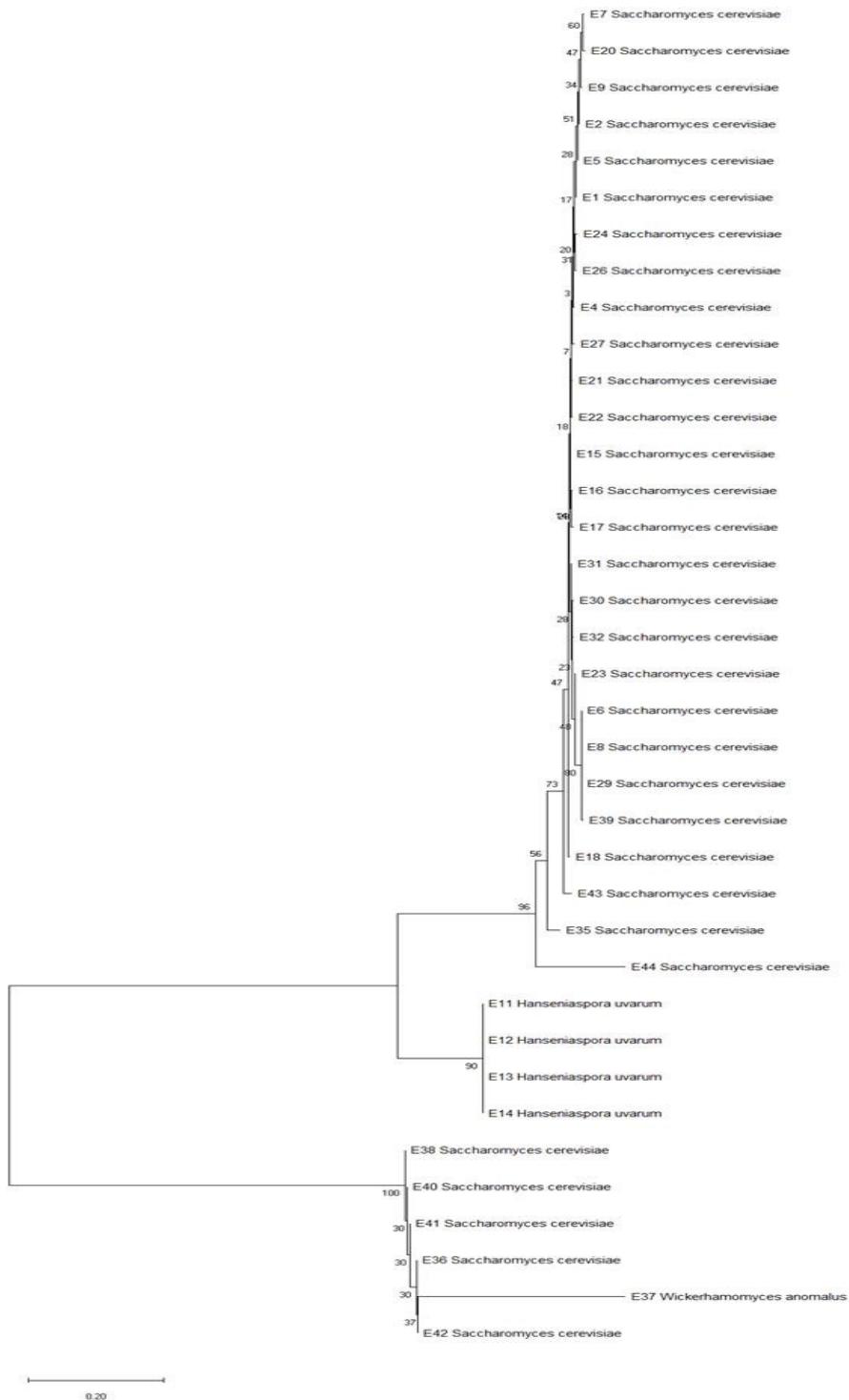


Figure 3.33. Phylogenetic Tree of Non-*Saccharomyces* and *Saccharomyces* Yeast in Emir Must and Wine with respect to ITS Region. The scale bar indicates substitutions per base pair.

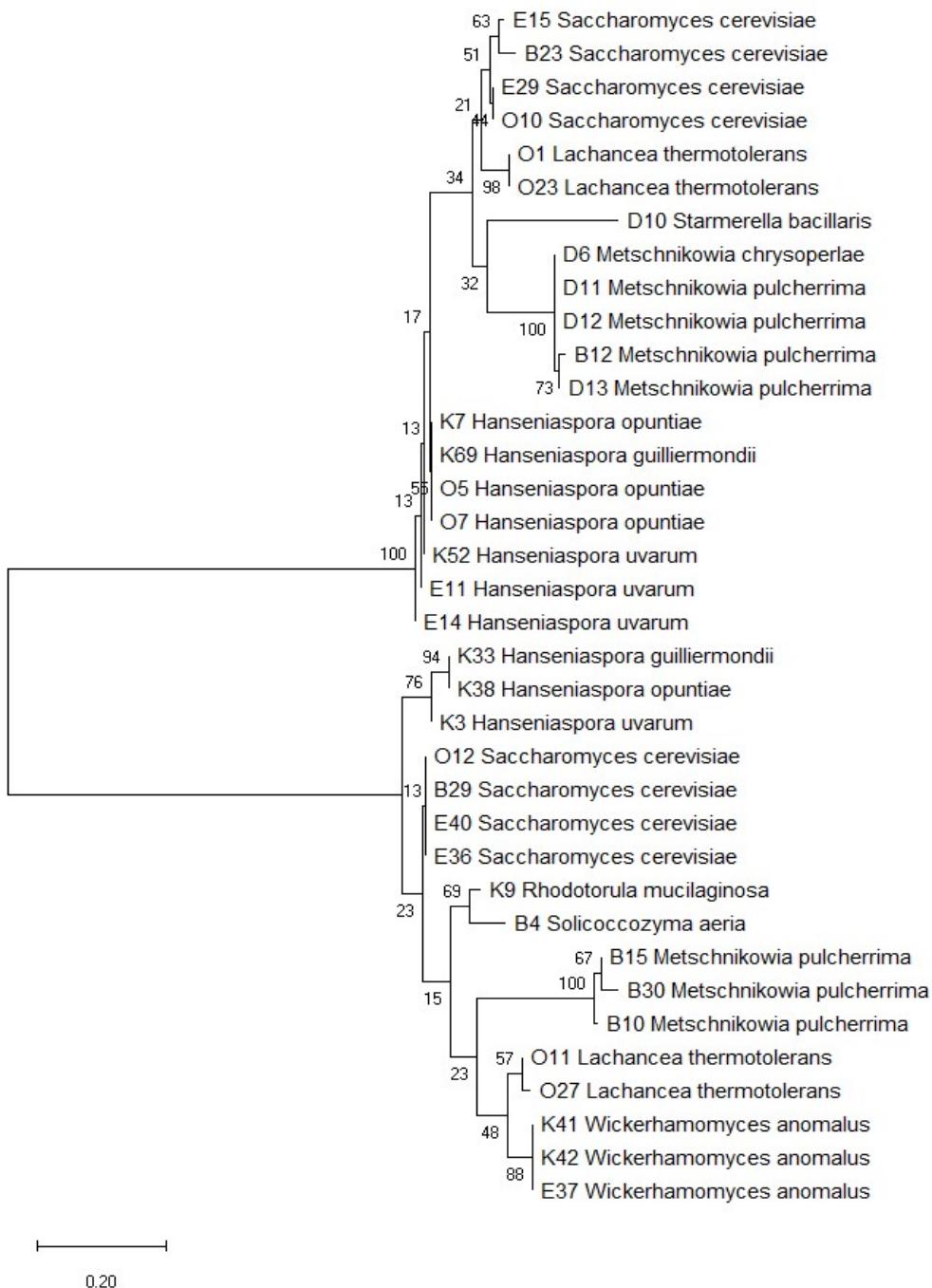


Figure 3.34. Phylogenetic Tree of Selected Non-*Saccharomyces* and *Saccharomyces* Yeast in Five Different Grapes' Musts and Wines with respect to ITS Region. The scale bar indicates substitutions per base pair.

The above phylogenetic tree was created with different selected species and different grape varieties. This showed the evolutionary relationship of each different microorganisms (Figure 3.34).

The similarities or differences of our isolates were determined according to phylogenetic tree. While the similarities of the same species were high, the species belonging to the same genus were clustered with high similarities as expected.

The numbers which were at nodes refer percentage of frequency. It indicated the stability of each branches in the tree.

It was concluded that the isolates of the same species from the same grape types were not related. These strains of the same species have been found in over 300 base pair differences. The difference between the same species concerning sequences of 26S rDNA regions was also found in studies about palm wine yeasts in recent years (Nwaiwu, 2019).

The following phylogenetic trees were created with respect to ITS region and D1/D2 domain of rDNA sequencing, respectively. This showed the evolutionary relationship of each different microorganisms according to ITS region and D1/D2 domain of ribosomal DNA. (Figure 3.35 and Figure 3.36).

According to the phylogenetic trees, *Hanseniaspora* species were found similar with respect to D1/D2 domains of rDNA sequences. Hesham et al. (2014) also revealed that D1/D2 domains of rDNA sequencing was more accurate method than 5.8S rDNA sequencing for identification of the microorganisms.

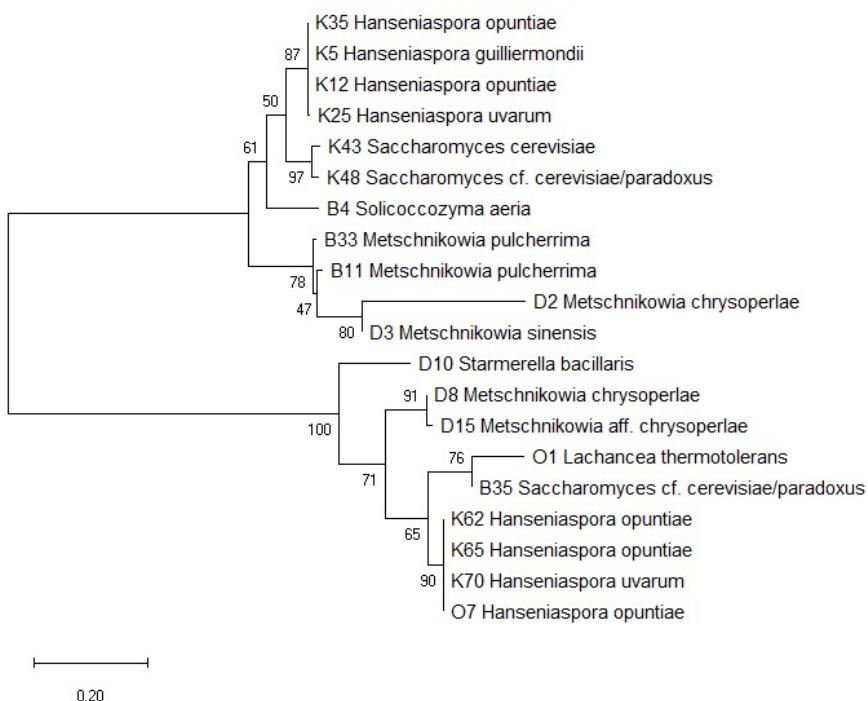


Figure 3.35. Phylogenetic Tree of Some Yeasts with respect to ITS Region. The scale bar indicates substitutions per base pair.

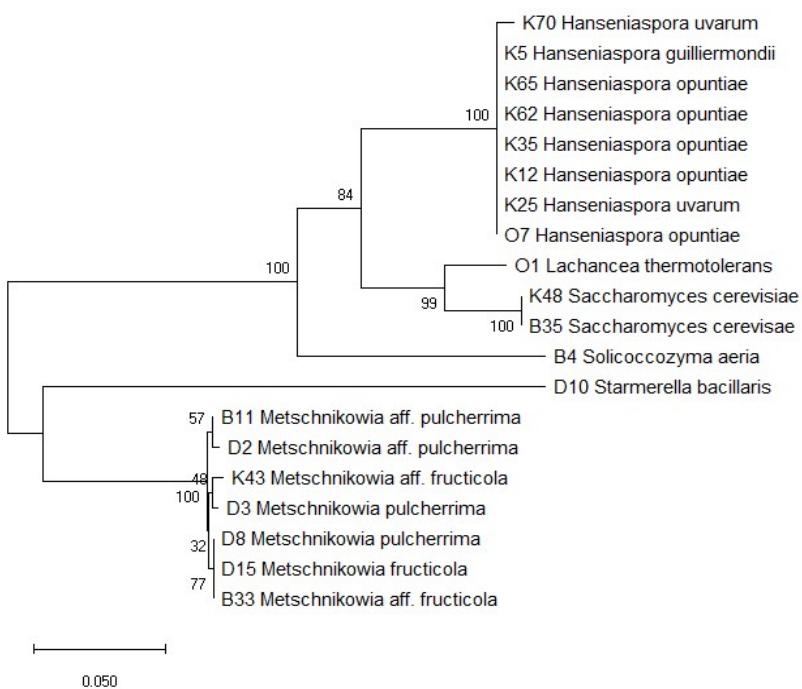


Figure 3.36. Phylogenetic Tree of Some Yeasts with respect to D1/D2 Domain. The scale bar indicates substitutions per base pair.

3.7. Phenotypic Characterization of Non-*Saccharomyces* Yeasts

The alcohol tolerances of non-*Saccharomyces* strains were generally found very low level. This is an expected result (Table 3.13). These strains, which are active during cold and normal maceration, leave the wine production to alcohol tolerant *Saccharomyces* strains (Urso et al., 2008). Non-*Saccharomyces* yeasts having low alcohol tolerance were used as a starter culture for production of wine containing low alcohol content (Jolly, Augustyn, & Pretorius, 2006). Therefore, these isolated non-*Saccharomyces* yeasts can be used as a starter culture for making the wine having low alcohol content. On the other hand, some isolated strains were found high alcohol tolerance. These isolates were *Wickerhamomyces anomalus* (KB 0.CM S5 and OA 4.CM S3), *Hanseniaspora opuntiae* (OA 4.CM NS4) and *Metschnikowia aff. pulcherrima* (DA 0.CM NS2) strains. These non-*Saccharomyces* yeasts can also be used as a starter culture with pure or mixed inoculation of the musts.

Moreover, these non-*Saccharomyces* yeasts highly produced H₂S. This situation was very similar to the literature. Çelik et al. (2017) were found high level H₂S producing non-*Saccharomyces* yeasts.

SO₂ addition was used to inhibit undesirable microorganisms' growth and control oxidation. SO₂ is a highly toxic compound for non-*Saccharomyces* yeasts. Therefore, SO₂ tolerance is a very desirable property for non-*Saccharomyces* yeasts for using as a starter culture of wine (González-Arenzana et al., 2017). Isolated non-*Saccharomyces* yeasts had high SO₂ tolerance in Table 3.13. Only one *Metschnikowia aff. fructicola* strain (KA 4.CM S1) had low SO₂ tolerance in this table. These isolates having high SO₂ tolerance can be used as a starter culture for winemaking.

In addition, phenotypic properties of all non-*Saccharomyces* yeasts were given Appendix D.

Table 3.13. Technological Properties of Non-Saccharomyces Strains

Non-Saccharomyces	Name	Alcohol Tolerance (%)				SO ₂ Tolerance (mg/L)			H ₂ S Production		Identification	
		10	13	15	50	100	150	200			ITS	D1/D2
		-	-	-	+	+	+	+	3	+	nd	
<i>Hanseniaspora guilliermondii</i>	KA 0.CM NS1	-	-	-	+	+	+	+	3	+	nd	
	KA 0.CM NS5	-	-	-	+	+	+	+	4	+	+	
<i>Hanseniaspora opuntiae</i>	KA 0.CM NS2	-	-	-	+	+	+	+	3	+	nd	
	KB 0.CM NS5	vw	vw	vw	+	+	+	+	3	+	nd	
	KA 2.NM NS1	+	-	-	+	+	+	-	2	+	nd	
	KA 4.NM NS5	-	-	-	+	+	+	+	2	+	nd	
<i>Rhodotorula mucilaginosa</i>	KB 0.CM NS4	-	-	-	+	+	+	+	3	+	nd	
<i>Hanseniaspora uvarum</i>	KA 4.CM NS3	+	vw	-	+	+	+	+	3	+	nd	
	KA 2.NM NS4	-	-	-	+	+	+	+	3	+	nd	
<i>Wickerhamomyces anomalus</i>	KA 0.CM S1	-	-	-	+	+	+	+	3	+	nd	
	KB 0.CM S5	+	+	+	+	+	+	+	ng	+	nd	
<i>Metschnikowia aff. fructicola</i>	KA 4.CM S1	-	-	-	-	-	-	-	ng	+	+	
<i>Hanseniaspora uvarum</i>	EA 1.W NS1	-	-	-	+	+	+	+	3	+	nd	
	EA 1.W NS4	vw	vw	vw	+	+	+	+	3	+	nd	
<i>Wickerhamomyces anomalus</i>	EA 0.NM NS8	-	-	-	+	+	+	+	3	+	nd	
<i>Lachancea thermotolerans</i>	OA 0.CM NS1	-	-	-	+	+	+	+	3	+	+	
	OB 4.CM NS4	-	-	-	+	+	+	+	3	+	nd	
<i>Wickerhamomyces anomalus</i>	OA 4CM S3	+	+	w	+	+	+	+	3	nd	+	
<i>Hanseniaspora opuntiae</i>	OA 4.CM NS2	vw	vw	vw	+	+	+	+	3	+	nd	
	OA 4.CM NS4	+	+	+	+	+	+	+	4	+	+	
<i>Metschnikowia pulcherrima</i>	BB 0.CM NS2	w	w	-	+	+	+	+	4	+	nd	
<i>Metschnikowia aff. fructicola</i>	BB 4.CM NS8	-	-	-	+	+	+	+	3	+	+	
<i>Metschnikowia aff. pulcherrima</i>	BB 0.CM NS3	-	-	-	+	+	+	+	5	+	+	
<i>Solicoccozyma aeria</i>	BA 0.CM NS4	-	-	-	+	w	w	w	5	+	+	
<i>Metschnikowia aff. pulcherrima</i>	DA 0.CM NS2	+	+	+	+	+	+	+	ng	+	+	
<i>Metschnikowia sinensis</i> (ITS)	DA 0.CM NS3	-	-	-	+	+	+	+	3	+		
<i>Metschnikowia pulcherrima</i> (D1/D2)										+		
<i>Starmeralla bacillaris</i>	DA 4.CM NS2	w	w	-	+	+	+	+	4	+	+	
<i>Metschnikowia pulcherrima</i>	DA 4.CM NS3	-	-	-	+	+	+	+	2	+	nd	
	DA 4.CM NS4	-	-	-	+	+	+	+	3	+	nd	
<i>Metschnikowia chrysoperlae</i>	DB 0.CM NS2	-	-	-	+	+	+	+	4	+	nd	
<i>Metschnikowia fructicola</i>	DB 4.CM NS3	-	-	-	+	+	+	+	4	+	+	

KA: A Parallel of Wine Made from Kalecik Karası Grape , KB: B Parallel of Wine Made from Kalecik Karası Grape, OA: A Parallel of Wine Made from Öküzgözü Grape , OB: B Parallel of Wine Made from Öküzgözü Grape, BA: A Parallel of Wine Made from Boğazkere Grape, BB: B Parallel of Wine Made from Boğazkere Grape, DA: A Parallel of Wine Made from Dimrit Grape, DB: B Parallel of Wine Made from Dimrit Grape, EA: A Parallel of Wine Made from Emir Grape, EB: B Parallel of Wine Made from Emir Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 2NM: 2. Day of Normal Maceration, 4NM: 4. Day of Normal Maceration, 1W: 1. Week; NS: Non-Saccharomyces ; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; nd: Not determined.

3.8. The Results of Wine Analysis Produced with Our Starter Cultures

Ethanol, total acidity, pH, volatile acidity, free SO₂, total SO₂, reducing sugar, malic acid and citric acid contents of our wines which were produced by adding our starter cultures were given in the Table 3.14.

Table 3.14. The Results of Wine Analysis

Samples	Ethanol (% v/v)	Total Acidity (g/L sulphuric acid)	pH	Volatile Acid (g/L sulphuric acid)	Total SO ₂ (mg/L)	Reducing Sugar (g/L)	Malic Acid (g/L)	Citric Acid (g/L)
E-LT-SC	12,6	2,2	3,8	0,16	221	0	1,42	0,38
E-WA-SC	12,7	2,2	3,8	0,2	119	0	1,42	0,22
E-SC	11,7	2,2	3,6	0,11	255	0	1,48	0,37
O-LT-SC	12,4	5,5	3,2	0,78	15	5,8	0,35	0,13
O-HO-SC	14,1	4,5	3,4	0,85	25	5,3	0,64	0,13
O-HG-SC	12,1	4,3	3,4	0,91	7	9,6	0,57	0,15
O-SC	13	5	3,1	0,39	27	1,2	1,51	0,28

E-LT-SC: Wine made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Emir grapes, E-WA-SC: Wine made by sequential inoculation of our *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* strains in Emir grapes, E-SC: Wine made by inoculation of our *Saccharomyces cerevisiae* strain in Emir grapes, O-LT-SC: Wine made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes, O-HO-SC: Wine made by sequential inoculation of our *Hanseniaspora opuntiae* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes, O-HG-SC: Wine made by sequential inoculation of our *Hanseniaspora guilliermondii* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes, O-SC: Wine made by inoculation of our *Saccharomyces cerevisiae* strain in Öküzgözü grapes.

Alcohol contents of all wines were within the limits according to the Turkish food codex as shown in Table 3.15. Although the total acidities of emir wines were slightly lower than expected, this value was suitable for Öküzgözü wines. It was higher than 2,29 g/L sulphuric acid. Moreover, the wine should be between pH 3 to 3,5. Although pH of Emir wines was slightly above this range, pH of Öküzgözü wines was within this value. Even though the amount of volatile acid was slightly higher in O-HG-SC, these values were still within the limits for all wines.

According to the reducing sugar quantity, wines made from emir grapes fall into the dry wine category, while wines made from Öküzgözü grape fall into the semi-dry category except O-SC. O-SC also fall into the dry wine category.

Although the total amount of SO₂ is slightly higher than the limits for Emir wines, this value decreases with time.

These values were found above and below the expected limits. These values were probably obtained because there is no curative treatment. This topic will be taken into consideration in our next wine trials.

Table 3.15. Limits of Wine Contents (Turkish Food Codex, 2008)

<i>Compounds</i>	<i>Limits</i>
Alcohol	9< and <15 (%v/v)
Total Acidity	>2,29 g/L sulphuric acid
pH	3<and <3,5
Volatile Acid	<0,886 g/L sulphuric acid for white and rose wines <0,985 g/L sulphuric acid for red wines
Total SO ₂	<250 mg/L for white and rose wines (for sugar content more than 5 g/L) <200 mg/L for white and rose wines (for sugar content less than 5 g/L) <200 mg/L for red wines (for sugar content more than 5 g/L) <150 mg/L for red wines (for sugar content less than 5 g/L)
Reducing Sugar (g/L)	<4 g/L for dry wines 4≤ and <12 g/L for semi-dry wine

3.9. The Results of Volatile Compounds and Sensory Analysis of Produced Wines

3.9.1. The Results of Volatile Compound Analysis

Before giving the results of volatile compound for each wine, odour threshold values and odour description of tested each aroma compounds were shown in the following table (Table 3.16) This table was created with respect to literature values.

Table 3.16. Volatile Compounds' Odour Threshold Values and Odour Descriptions

Compound	Odour Threshold ($\mu\text{g/L}$)	Odour Description
Alcohols		
1-Propanol	306000 ⁽³⁾	Ripe fruit, alcohol ⁽³⁾
Isobutyl Alcohol	40000 ⁽¹⁾	Alcohol, winelike, nail polish ⁽¹⁾
1-Butanol	150000 ⁽¹⁾	Medicinal, alcohol ⁽¹⁾
Isoamyl Alcohol	60000 ⁽¹⁾	Whiskey, nail polish ⁽¹⁾
3-Methyl-1-pentanol	50000 ⁽¹⁾	Herbaceous, cocoa ⁽¹⁾
3-Ethoxy-1-propanol	100 ⁽⁴⁾	Fruity ⁽⁴⁾
Methionol	500 ⁽⁴⁾	Cooked vegetable ⁽⁴⁾
Benzyl Alcohol	100000 ⁽¹⁾	Almond ⁽¹⁾
Phenylethyl Alcohol	10000 ⁽¹⁾	Rose, pollen, perfume ⁽¹⁾
3-Methyl-3-butene-1-ol	600 ⁽⁴⁾	Sweet fruity ⁽⁴⁾
1-Pentanol	64000 ⁽⁴⁾	Almond, synthetic, balsamic ⁽⁴⁾
4-Methyl-1-pentanol	5000 ⁽¹⁾	Almond, toasted ⁽¹⁾
2,3-Butanediol	150000 ⁽⁴⁾	Fruity ⁽⁴⁾
1-Heptanol	1000 ⁽¹⁾	Green, sweet ⁽¹⁾
(Z)-3-Hexene-1-ol	400 ⁽¹⁾	Green, cypress ⁽¹⁾
(E)-3-Hexene-1-ol	400 ⁽⁵⁾	Green grass, herb ⁽²⁾
2-(Methylthio)ethanol	250 ⁽⁶⁾	French bean, cauliflower ⁽⁶⁾
2-Hexanol	not found	Herbaceous, medicine ⁽¹²⁾
3-Hexanol	400 ⁽⁹⁾	Flowery, soap ⁽¹²⁾
1-Hexanol	8000 ⁽¹⁾	Green, grass ⁽¹⁾
Acetates		
Isoamyl acetate	30 ⁽¹⁾	Banana, fruity, sweet ⁽¹⁾
Isopropyl acetate	not found	Fruity ⁽⁹⁾
Phenethyl acetate	250 ⁽¹⁾	Pleasant, floral ⁽¹⁾
Esters		
Ethyl lactate	154636 ⁽¹⁾	Lactic, raspberry ⁽¹⁾
Ethyl octanoate	5 ⁽¹⁾	Fruity, pineapple, pear, floral ⁽¹⁾
Ethyl-3-hydroxybutyrate	20000 ⁽³⁾	Green grape, marshmallow ⁽³⁾
Ethyl 9-decenoate	100 ⁽¹⁾	Fruity ⁽¹⁾
Diethyl succinate	6000 ⁽¹⁾	Light fruity, wine ⁽¹⁾
Ethyl butyrate	20 ⁽¹⁾	Strawberry, apple, banana ⁽¹⁾
Ethyl decanoate	200 ⁽¹⁾	Waxy, fruity, rose ⁽¹⁾
Ethyl hexanoate	5 ⁽¹⁾	Flowery, fruity ⁽¹⁾

Table 3.16 (continued)

Compound	Odour Threshold ($\mu\text{g/L}$)	Odour Description
Aldehydes		
Nonanal	2,8 ⁽⁴⁾	Citrusy, floral ⁽⁴⁾
Acids		
Butanoic acid	10000 ⁽⁴⁾	Rancid, cheese ⁽⁴⁾
Hexanoic acid	3000 ⁽¹⁾	Cheese, fatty ⁽¹⁾
Octanoic acid	1000 ⁽¹⁾	Cheese, fatty acid, rancid, harsh ⁽¹⁾
Decanoic acid	10000 ⁽¹⁾	Fatty, unpleasant ⁽¹⁾
Propanoic acid	8100 ⁽¹⁾	Cheese ⁽¹⁾
Nonanoic acid	3000 ⁽⁴⁾	Fatty ⁽⁴⁾
Tetradecanoic acid	10000 ⁽⁴⁾	Not found ⁽⁴⁾
Dodecanoic acid	1000 ⁽¹⁾	Dry, metallic, laurel oil flavour ⁽¹⁾
Pentanoic acid	3000 ⁽⁴⁾	Cheese ⁽⁴⁾
Isobutyric acid	2300 ⁽¹⁾	Cheese, butter, rancid ⁽¹⁾
Heptanoic acid	not found ⁽⁴⁾	Sweaty, cheese ⁽⁴⁾
Acetic acid	1200000 ⁽¹³⁾	Unpleasant ⁽¹³⁾
Isovaleric acid	30 ⁽¹¹⁾	Rancid, cheese, feet, floral ⁽¹¹⁾
Lactones		
γ -butyrolactone	20000 ⁽¹¹⁾	Faint, sweet, caramel ⁽¹¹⁾
Pantolactone	2000 ⁽¹¹⁾	Spicy, caramel ⁽¹¹⁾
Phenols		
2-Methoxy-4-vinylphenol	40 ⁽⁴⁾	Spices, curry ⁽⁴⁾
4-vinyl-phenol	180 ⁽⁹⁾	Pharmaceutical ⁽¹⁰⁾
Hydrocarbons		
Phenol	7100 ⁽⁷⁾	Sickeningly sweet, irritating ⁽⁸⁾
Other Compounds		
Guaiacol	10 ⁽⁴⁾	Smoke, sweet, medicine ⁽⁴⁾
Acetoin	150000 ⁽⁹⁾	Flowery, wet ⁽²⁾
Geraniol	20 ⁽³⁾	Roses, geranium ⁽³⁾
Syringol	570 ⁽⁷⁾	Not found

(1) Wang, X., et. al., 2017

(2) Liu, N., et. al., 2015

(3) Arcari, S., et. al., 2017

(4) Welke, J., et. al., 2014

(5) Tao, Y., et. al., 2010

(6) Moreira, N., et. al., 2010

(7) Parker, M., et. al., 2012

(8) Vries, C., et. al., 2016

(9) Escudero, A., et. al., 2007

(10) Swiegers, J., et. al., 2005

(11) Celik, Z., et. al., 2018

(12) Darici, M., et. al., 2013

(13) Macias, M., et. al., 2012.

3.9.1.1. The Result of Volatile Compound Analysis of White Wine

White wines were made from Emir grapes which were taken from Cappadocia at 2018. One of the white wines was prepared with sequential inoculation with strain OB 4.CM NS4 (*Lachancea thermotolerans*) and strain OB 4.CM S1 (*Saccharomyces cerevisiae*) (E-LT-SC). The other was made with strain OB 4.CM S1 (*Saccharomyces cerevisiae*) (E-SC). The following two diagrams were shown the peaks of volatile compounds in GC-MS chromatogram for E-LT-SC and E-SC, respectively (Figure 3.37 and 3.38).

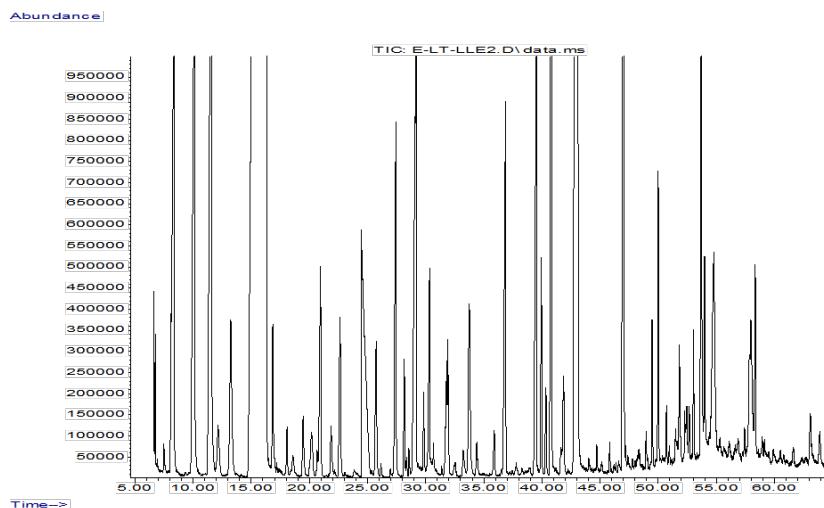


Figure 3.37. GC-MS Chromatogram of E-LT-SC Sample

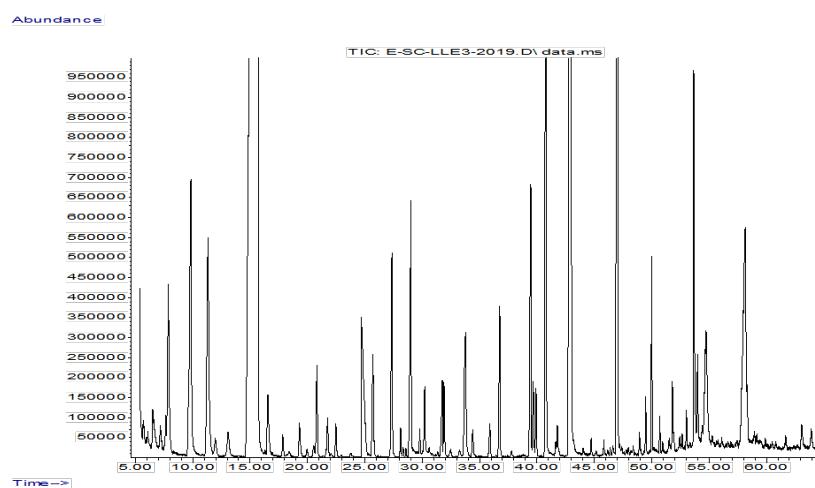


Figure 3.38. GC-MS Chromatogram of E-SC Sample

Moreover, traditional wine (E-A) and commercial wine with using the Laffort starter culture (E-Laff) were made in 2017. These Emir grapes also were taken in Cappadocia.

Table 3.17 was shown volatile compounds of four wine samples (E-A, E-Laff, E-LT-SC, and E-SC). According to GS-MS results, 42 aroma compounds were identified for E-LT-SC and E-SC while 59 volatile compounds were found in E-A and E-Laff. These compounds included alcohols, acetates, esters, aldehydes, acids, lactones, phenols, and other compounds such as tyrosol and acetoin. 46 of them were significantly different among these samples. The total concentrations of aroma compounds were found 150.4 mg/L in E-A, 144.4 mg/L in E-Laff, 131.7 mg/L in E-LT-SC and 113.2 mg/L in E-SC.

The highest amount of volatile compounds was found alcohols, then acids and esters were followed this alignment. Higher alcohols, acids and esters have important roles in giving flavor of wines with respect to amounts and concentration of these compounds (Valero, Moyano, Millan, Medina, & Ortega, 2002).

When concentrations of alcohols were under 300 mg/L, they gave desirable flavor of wine. In contrast, if these values were over 400 mg/L, they attributed negative impacts (Pastor, Huerta, Mateo, & Jime, 2001). The amounts of alcohols were found 140.8 mg/L, 130.9 mg/L, 112.6 mg/L and 95.4 mg/L for E-A, E-Laff, E-LT-SC and E-SC, respectively. In these alcohols, isoamyl alcohol and phenethyl alcohol were found very high level in all of the wine samples while 3-Ethoxy-1-propanol were only identified highly in E-LT-SC and E-SC. Isoamyl alcohol and phenethyl alcohol had suprathreshold OAVs ($OAV>1$) and gave whiskey and rose aroma in these wines, respectively (Table 3.18). Moreover, fruity aroma (3-Ethoxy-1-propanol) was found higher in E-LT-SC wine. Cabaroglu et al. (1997) mentioned that alcohols were an important part of aroma substances in neutral aromatic grape varieties such as Emir.

Table 3.17. Aroma Compounds ($\mu\text{g/L}$) of Emir and Kalecik Karasi Wines with GS-MS Results

<i>Compound</i>	<i>RI</i>	<i>E-A</i>	<i>E-Laff</i>	<i>E-LT-SC</i>	<i>E-SC</i>	<i>K-J3</i>	<i>K-HU</i>	<i>K-SC</i>	<i>K-A</i>
Alcohols									
1-Propanol*	1031	585.94 \pm 30.6 ^E	535.50 \pm 32.59 ^E	6096.35 \pm 252.05 ^A	3168.24 \pm 188.72 ^B	1315.1 \pm 64.92 ^D	1485.67 \pm 146.46 ^D	1709.22 \pm 136.9 ^D	2765.34 \pm 103.52 ^C
Isobutyl Alcohol*	1098	7094.13 \pm 545.87 ^{CD}	5791.22 \pm 456.99 ^{DE}	5249.09 \pm 135.80 ^E	4709.24 \pm 48.74 ^E	8164.57 \pm 613.75 ^C	8473.2 \pm 328.31 ^C	1550.84 \pm 966.7 ^A	12600.61 \pm 474.10 ^B
1-Butanol*	1151	Nd	Nd	505.50 \pm 31.02 ^A	298.61 \pm 17.98 ^B	331.86 \pm 17.25 ^B	138.2 \pm 12.79 ^D	191.70 \pm 13.17 ^C	479.13 \pm 7.32 ^A
Isoamyl Alcohol*	1236	107983.46 \pm 10207.44 ^{AC}	100487.00 \pm 9980.9 ^{BCD}	83097.76 \pm 115.89 ^{DE}	71677.76 \pm 1129.77 ^E	93417.79 \pm 6180.34 ^{CD}	85286.88 \pm 295.02 ^{DE}	123340.20 \pm 10766.2 ^A	113586.80 \pm 3887.80 ^{AB}
3-Methyl-1-pentanol*	1341	74.19 \pm 7.82 ^B	100.39 \pm 9.51 ^A	Nd	Nd	91.38 \pm 6.91 ^A	58.94 \pm 1.80 ^C	45.93 \pm 2.47 ^C	61.43 \pm 4.53 ^{BC}
3-Ethoxy-1-propanol*	1389	38.09 \pm 3.77 ^E	41.06 \pm 3.75 ^E	510.90 \pm 20.70 ^A	185.95 \pm 3.26 ^C	92.19 \pm 6.22 ^D	229.08 \pm 1.0 ^B	52.90 \pm 4.90 ^E	227.37 \pm 9.25 ^B
3-(Ethylthio)-1-propanol*	1802	33.15 \pm 3.13 ^A	23.21 \pm 0.39 ^B	Nd	Nd	Nd	Nd	Nd	Nd
Methionol*	1721	417.14 \pm 18.52 ^C	418.77 \pm 29.46 ^C	94.32 \pm 8.25 ^D	127.47 \pm 4.84 ^D	405.97 \pm 32.63 ^C	731.69 \pm 15.33 ^A	542.25 \pm 45.98 ^B	712.09 \pm 26.68 ^A
Benzyl Alcohol*	1853	42.60 \pm 3.86 ^F	57.01 \pm 4.44 ^{EF}	73.10 \pm 2.07 ^E	54.69 \pm 0.64 ^{EF}	200.40 \pm 19.08 ^B	165.95 \pm 3.27 ^C	139.55 \pm 13.20 ^D	255.13 \pm 8.93 ^A
2-Methyl-2-butene-1-ol*	1332	37.53 \pm 3.51 ^B	34.16 \pm 2.46 ^{BC}	Nd	102.43 \pm 2.58 ^A	37.03 \pm 1.88 ^B	28.09 \pm 1.48 ^C	Nd	Nd
Phenylethyl Alcohol*	1916	21882.72 \pm 759.51 ^C	20826.49 \pm 1767.25 ^C	12913.18 \pm 514.45 ^D	12587.35 \pm 295.67 ^D	21740.58 \pm 510.83 ^C	26839.23 \pm 527.02 ^B	19789.79 \pm 836.44 ^C	30431.02 \pm 637.41 ^A
3-Methyl-3-butene-1-ol*	1250	88.64 \pm 5.39 ^C	111.31 \pm 7.95 ^B	Nd	Nd	90.62 \pm 8.94 ^C	74.34 \pm 5.25 ^{CD}	71.75 \pm 4.45 ^D	134.04 \pm 1.18 ^A
1-Pentanol*	1262	73.91 \pm 6.93 ^{BC}	104.35 \pm 5.62 ^A	Nd	Nd	79.88 \pm 6.15 ^B	63.56 \pm 3.11 ^C	71.68 \pm 2.87 ^{BC}	82.61 \pm 6.48 ^B
4-Methyl-1-pentanol*	1328	35.19 \pm 2.90 ^{AB}	34.33 \pm 2.64 ^{AB}	Nd	Nd	40.77 \pm 3.38 ^A	30.25 \pm 1.32 ^B	Nd	Nd
2,3-Butanediol*	1517	1840.09 \pm 141.79 ^E	1592.14 \pm 34.41 ^E	2554.09 \pm 130.33 ^D	1727.77 \pm 51.04 ^E	4379.08 \pm 409.24 ^B	3347.99 \pm 227.38 ^C	3621.41 \pm 120.30 ^C	5195.34 \pm 209.66 ^A
1-Heptanol*	1421	108.91 \pm 7.31 ^A	45.07 \pm 4.01 ^C	58.23 \pm 5.21 ^C	Nd	88.44 \pm 3.80 ^B	48.14 \pm 3.30 ^C	57.75 \pm 3.35 ^C	51.87 \pm 4.81 ^C
(Z)-3-Hexene-1-ol*	1394	53.36 \pm 4.27 ^B	91.12 \pm 8.92 ^A	Nd	Nd	32.99 \pm 1.98 ^{CD}	22.21 \pm 1.59 ^D	32.64 \pm 2.79 ^D	45.11 \pm 2.68 ^{BC}
(E)-3-Hexene-1-ol*	1384	34.83 \pm 3.27 ^A	34.93 \pm 3.11 ^A	Nd	Nd	37.91 \pm 2.79 ^A	25.41 \pm 0.65 ^B	40.23 \pm 2.42 ^A	39.99 \pm 3.17 ^A

Table 3.17 (continued)

Alcohols	R	E-A	E-Laff	E-LT-SC	E-LT-HU	K-J3	K-HU	K-SC	K-A
2-(Methylthio)ethanol*	1497	40.93±4.26 ^D	39.80±2.30 ^D	63.78±3.58 ^{EC}	46.87±2.66 ^D	75.72±2.38 ^B	94.76±7.13 ^A	66.58±6.37 ^{EC}	62.38±2.90 ^C
1,2-Propanediol*	1583	46.65±2.40 ^C	53.21±3.61 ^C	Nd	Nd	77.44±6.87 ^B	54.70±3.13 ^C	85.12±6.36 ^B	103.07±5.38 ^A
3-Penten-2-ol*	1160	130.82±12.47 ^C	298.33±28.66 ^B	938.13±37.06 ^A	311.43±6.10 ^B	78.97±3.12 ^{CD}	52.14±3.89 ^D	120.10±8.77 ^C	266.97±7.10 ^B
2-Hexanol*	1313	108.76±6.05 ^B	100.90±3.36 ^B	228.55±8.52 ^A	244.79±13.11 ^A	60.06±5.92 ^C	32.84±2.79 ^D	55.44±4.47 ^C	33.77±2.93 ^D
3-Hexanol*	1207	22.14±1.05 ^B	51.91±0.92 ^A	Nd	Nd	Nd	Nd	Nd	Nd
1-Hexanol	1347	Nd	Nd	191.67±3.30 ^A	161.11±13.93 ^A	Nd	Nd	Nd	Nd
2-Phenoxyethanol*	2142	34.42±1.38 ^A	22.43±1.51 ^B	Nd	Nd	Nd	Nd	Nd	Nd
Total		140773.8	130894.64	112574.65	95403.71	130838.75	127283.27	165542.69	167134.07
Acetates									
Isoamyl acetate*	1132	460.66±20.99 ^F	1410.11±25.91 ^D	2749.99±87.16 ^A	1768.53±42.58 ^C	1743.33±80.64 ^C	1452.16±9.78 ^D	2242.31±86.81 ^B	807.86±59.84 ^F
Isopropyl acetate	913	70.87±2.37 ^A	69.56±6.43 ^A	Nd	Nd	Nd	Nd	Nd	Nd
Phenethyl acetate*	1827	82.99±7.39 ^E	151.87±14.80 ^D	290.88±11.20 ^B	146.32±4.04 ^D	273.17±2.35 ^B	330.34±7.50 ^A	227.86±22.65 ^C	226.46±14.20 ^C
Propyl acetate*	953	53.62±5.53 ^A	41.9±3.60 ^B	Nd	Nd	Nd	Nd	Nd	Nd
Isobutyl acetate	1029	Nd	Nd	189.28±12.10	Nd	Nd	Nd	Nd	Nd
Total	668.14	1673.44	3230.1500	1914.85	2016.5	1782.5	2470.17	1034.32	
Esters									
Ethyl lactate*	1363	218.76±21.50 ^E	202.77±19.88 ^E	565.15±18.23 ^{CD}	485.53±24.10 ^{CD}	611.23±19.31 ^{BC}	693.95±14.65 ^B	436.28±41.28 ^D	2059.53±125.75 ^A
Ethyl octanoate*	1412	178.13±13.64 ^D	256.34±12.37 ^C	338.60±7.76 ^B	409.85±1.13 ^A	133.46±6.95 ^E	81.48±12.95 ^F	230.00±12.24 ^C	150.67±6.73 ^{DE}

Table 3.17 (continued)

Esters	<i>R</i>	<i>E-A</i>	<i>E-Loff</i>	<i>E-LT-SC</i>	<i>E-SC</i>	<i>K-J3</i>	<i>K-HU</i>	<i>K-SC</i>	<i>K-A</i>
Ethyl 3-hydroxybutyrate*	1472	90.16±7.27 ^D	86.90±5.75 ^D	182.24±15.36 ^C	96.76±8.53 ^D	206.56±18.12 ^{SC}	249.23±8.90 ^A	218.01±18.89 ^{ABC}	245.64±24.47 ^{AB}
Ethyl 9-deenoate	1712	24.44±0.70 ^A	27.47±2.04 ^A	Nd	Nd	Nd	Nd	Nd	Nd
Diethyl succinate*	1701	44.04±2.40 ^F	64.27±4.58 ^{EF}	74.64±2.48 ^{EF}	76.24±4.24 ^E	228.97±7.68 ^B	142.09±12.56 ^D	198.97±6.40 ^C	259.38±19.83 ^A
Ethyl 4-hydroxybutanoate*	1819	1264.82±69.81 ^{DE}	1480.22±106.25 ^D	1314.64±37.43 ^{DE}	1124.60±52.76 ^E	2922.64±103.64 ^B	2344.84±66.16 ^C	2058.77±202.55 ^C	3684.69±129.87 ^A
Monoethyl succinate*	2350	659.10±41.33 ^E	Nd	1046.13±42.87 ^{DE}	722.32±30.37 ^{DE}	2610.71±248.74 ^A	1157.67±84.94 ^D	1700.59±97.35 ^C	2107.06±177.58 ^B
Ethyl hydrogen succinate	1687	Nd	541.61±39.90	Nd	Nd	Nd	Nd	Nd	Nd
Ethyl butyrate	1044	259.37±3.43 ^A	279.63±23.63 ^A	Nd	Nd	Nd	Nd	Nd	Nd
Ethyl 3-hydroxypropionate*	1587	26.60±1.39 ^C	26.39±1.79 ^C	Nd	Nd	88.55±1.18 ^A	65.98±5.53 ^B	73.37±5.41 ^B	65.18±5.02 ^B
Ethyl decanoate*	1610	40.52±3.11 ^B	55.39±0.81 ^B	463.98±29.26 ^A	496.05±14.03 ^A	Nd	Nd	Nd	Nd
Ethyl 2-hydroxy-3-phenylpropanoate*	2223	Nd	Nd	Nd	Nd	92.33±5.08 ^B	Nd	73.46±6.62 ^B	135.75±11.14 ^A
Methyl 4-hydroxybutanoate*	1802	Nd	Nd	Nd	Nd	20.68±1.03 ^C	28.92±1.65 ^B	16.07±1.34 ^D	56.39±1.68 ^A
Ethyl hexanoate*	1241	469.11±38.99 ^B	449.26±32.27 ^B	444.94±30.92 ^B	579.79±38.80 ^A	248.64±17.46 ^C	209.52±18.45 ^C	396.67±33.92 ^B	249.93±23.04 ^C
<i>Total</i>		3275.05	3470.25	4430.32	3991.14	7163.77	4973.68	5402.19	9014.22
Aldehydes									
Nonanal	1658	922.22±0.00	922.22±0.00	922.22±0.00	922.22±0.00	922.22±0.00	922.22±0.00	922.22±0.00	922.22±0.00

Table 3.17 (continued)

Acids	<i>RI</i>	<i>E-A</i>	<i>E-Laff</i>	<i>E-IT-SC</i>	<i>E-SC</i>	<i>K-J3</i>	<i>K-HU</i>	<i>K-SC</i>	<i>K-A</i>
Butanoic acid*	1604	107.35±8.41 ^C	250.48±22.72 ^B	335.22±27.29 ^A	345.60±9.32 ^A	Nd	Nd	353.34±27.40 ^A	161.70±12.85 ^C
Hexanoic acid*	1832	696.70±38.14 ^D	1741.68±54.76 ^C	2428.84±147.12 ^B	3339.72±72.71 ^A	848.87±239.94 ^D	794.08±3.06 ^D	1528.31±136.78 ^C	556.21±50.67 ^D
Octanoic acid*	2106	1057.21±50.64 ^E	2251.81±58.47 ^C	3300.29±195.00 ^B	3973.74±113.58 ^A	527.04±3.10 ^F	548.38±22.41 ^F	1317.59±69.03 ^P	433.89±35.17 ^F
Decanoic acid*	2231	127.71±10.38 ^A	138.02±9.34 ^A	132.54±8.44 ^A	78.99±6.42 ^B	Nd	Nd	77.44±5.59 ^B	Nd
Propanoic acid*	1508	43.21±3.44 ^D	53.86±3.07 ^D	106.87±3.01 ^B	58.65±0.27 ^D	134.87±12.60 ^A	99.56±5.07 ^{BC}	89.04±6.67 ^C	124.89±5.71 ^A
Nonanoic acid*	2157	28.21±2.66 ^D	49.46±3.38 ^C	171.29±8.93 ^A	109.98±4.13 ^B	43.71±3.92 ^C	8.99±0.44 ^E	49.80±1.64 ^C	22.81±0.73 ^D
Tetradecanoic acid*	2717	39.64±3.69 ^B	69.45±3.13 ^A	Nd	Nd	Nd	Nd	Nd	Nd
Dodecanoic acid*	2479	56.71±3.53 ^B	108.84±9.01 ^A	Nd	Nd	Nd	Nd	Nd	Nd
Pentanoic acid*	1689	104.49±9.15 ^E	374.67±6.30 ^D	Nd	Nd	623.99±24.75 ^B	405.91±4.83 ^{CD}	804.82±58.30 ^A	463.38±43.73 ^C
Isobutyric acid*	1579	72.21±2.66 ^E	229.29±15.09 ^D	197.95±16.20 ^D	119.39±1.35 ^{DE}	491.97±44.85 ^B	360.52±10.55 ^C	885.63±75.51 ^A	389.59±29.94 ^C
Heptanoic acid*	1934	Nd	Nd	86.06±4.65 ^A	Nd	70.31±3.63 ^{BC}	62.65±4.21 ^C	30.37±0.93 ^D	74.43±1.94 ^B
Acetic acid*	1403	632.16±26.70 ^D	541.41±1.49 ^D	1662.24±58.48 ^A	1317.76±78.73 ^C	1522.34±103.41 ^{BC}	691.19±20.50 ^D	1379.79±130.75 ^C	3414.62±56.48 ^A
Diethyl dl-malate*	2053	Nd	Nd	Nd	Nd	35.80±2.19 ^A	29.20±2.13 ^B	27.01±1.67 ^B	36.58±2.27 ^A
<i>Total</i>		2985.6	5808.97	8421.3	9343.83	4298.9	3000.48	6543.14	5678.1
Lactones									
γ -butyrolactone*	1592	283.19±25.34 ^E	339.38±21.98 ^E	522.28±29.07 ^{CD}	578.69±4.20 ^C	1216.72±30.83 ^A	1074.37±20.60 ^B	495.74±41.53 ^D	1175.99±30.32 ^A
Pantolactone*	2034	62.26±6.14 ^{CDE}	50.40±0.91 ^E	72.87±5.18 ^{BC}	54.41±4.55 ^{DE}	65.24±6.42 ^{BCD}	64.53±3.69 ^{BCD}	86.67±2.77 ^A	77.34±3.87 ^{AB}
<i>Total</i>		345.45	389.78	595.5	633.1	1281.96	1138.9	582.41	1233.33

Table 3.17 (continued)

Phenols	RI	E-A	E-Laff	E-LT-SC	E-SC	K-13	K-HU	K-SC	K-A
2-Methoxy-4-vinylphenol*	2168	142.37±9.09 ^C	176.56±5.31 ^B	283.40±18.50 ^A	290.51±5.90 ^A	79.03±1.40 ^D	52.17±4.49 ^E	286.06±0.67 ^A	153.11±7.83 ^C
4-vinyl-phenol*	2334	Nd	Nd	329.95±28.57 ^A	254.27±12.56 ^B	Nd	Nd	212.27±6.16 ^B	100.22±4.13 ^C
Total	142.37	176.56	613.35	544.78	79.03	52.17	498.33	253.33	
Hydrocarbons									
Phenol*	1947	24.18±1.77 ^A	12.16±0.81 ^B	Nd	Nd	Nd	Nd	Nd	Nd
Other Compounds									
Tyrosol*	2965	1874.52±36.44 ^B	1723.01±51.70 ^C	1666.29±25.31 ^C	1208.80±9.31 ^D	722.41±66.06 ^F	906.56±18.25 ^E	1656.07±60.20 ^C	2217.50±33.10 ^A
Soleron*	2096	35.13±3.36 ^C	22.47±0.57 ^C	Nd	81.83±7.33 ^B	34.52±3.10 ^C	82.17±5.29 ^B	97.06±8.48 ^A	
Guaiacol*	1840	63.04±4.82 ^B	57.14±1.00 ^B	Nd	61.30±1.36 ^B	37.13±3.36 ^C	57.41±5.72 ^B	94.69±4.78 ^A	
Acetoin*	1291	190.28±10.74 ^C	175.87±15.96 ^C	168.09±0.18 ^C	178.74±7.77 ^C	200.67±19.09 ^C	270.36±26.70 ^B	190.79±14.51 ^C	1292.28±26.83 ^A
Geraniol*	1844	Nd	Nd	Nd	Nd	24.01±1.64 ^B	25.28±1.423 ^B	36.58±3.65 ^A	25.89±1.60 ^B
Syringol*	2854	Nd	Nd	Nd	Nd	61.30±5.04 ^C	37.53±1.21 ^D	163.60±11.08 ^B	190.71±0.51 ^A
Total	2162.97	1978.49	1834.38	1387.54	1151.52	1311.38	2186.62	3918.13	
<i>TOTAL</i>	150376.94	144404.29	131699.3	113218.95	146830.43	13542.38	183225.55	183235.5	

E-A: Emir traditional wine. E-Laff: Emir commercial wine. E-LT-SC: Emir wine sample inoculated with strain OB 4.CM NS4 (*Lachancea thermotolerans*) and strain OB 4.CM S1 (*Saccharomyces cerevisiae*). E-SC: Emir wine sample inoculated with strain OB 4.CM S1 (*Saccharomyces cerevisiae*). K-13: Kalecik Karasi wine sample inoculated with strain 13 (*Saccharomyces cerevisiae*). K-HU: Kalecik Karasi wine sample inoculated with *Hanseniaspora warrenii*. K-SC: Kalecik Karasi wine sample inoculated with commercial *Saccharomyces cerevisiae* (Chr. Hansen). K-A: Traditional wine made by Eliaz Seydi Monir. RI: retention indices. *: significantly different ($P < 0.05$). A,B,C,D,E,F: refer significantly different. Nd: not determined.

3.27 mg/L, 3.47 mg/L, 4.43 mg/L and 3.99 mg/L of esters were found in E-A, E-Laff, E-LT-SC and E-SC, respectively. Ethyl 4-hydroxybutanoate, monoethyl succinate, ethyl lactate, ethyl decanoate, ethyl hexanoate, and ethyl octanoate were found high amount in these wines. They had important roles for young wines and occurred in maceration time. In addition, these aromas played critical roles for giving fruity flavor in the wines (Sellie, et. al, 2004; Herraiz, et. al., 1991) Ethyl octanoate, ethyl decanoate, and ethyl hexanoate had suprathreshold OAVs and gave fruity aroma. Especially, these aroma compounds were found at a higher level in E-LT-SC and E-SC than E-A and E-Laff. The amount of ethyl decanoate was found at 10 times high in E-LT-SC and E-SC in comparison with E-A and E-Laff. Erten & Tanguler (2010) also stated the similar result obtained in the Emir wine sample which produced in the mixed inoculation of *Williopsis saturnus* and *Saccharomyces cerevisiae* (OAV>1).

Table 3.18. Suprathreshold Compounds for E-A, E-Laff, E-LT-SC and E-SC of Emir Wines

Compound	RI	Odour Threshold ($\mu\text{g}/\text{L}$)	Odour Activity Values (OAV)	Odour Description
Alcohols				
Isoamyl Alcohol	1236	60000	>1	Whiskey, nail polish ⁽¹⁾
3-Ethoxy-1-propanol	1389	100	>1	Fruity ⁽²⁾
Phenylethyl Alcohol	1916	10000	>1	Rose, pollen, perfume ⁽¹⁾
Acetates				
Isoamyl acetate	1132	30	>1	Banana, fruity, sweet ⁽¹⁾
Phenethyl acetate	1827	250	>1	Pleasant, floral ⁽¹⁾
Esters				
Ethyl octanoate	1412	5	>1	Fruity, pineapple, pear, floral ⁽¹⁾
Ethyl decanoate	1610	200	>1	Waxy, fruity, rose ⁽¹⁾
Ethyl hexanoate	1241	5	>1	Flowery, fruity ⁽¹⁾
Aldehydes				
Nonanal	1658	2.8	>1	Citrusy, floral ⁽⁴⁾
Acids				
Hexanoic acid	1832	3000	>1	Cheese, fatty ⁽¹⁾
Octanoic acid	2106	1000	>1	Cheese, fatty acid, rancid, harsh ⁽¹⁾
Phenols				
2-Methoxy-4-vinylphenol	2168	40	>1	Spices, curry ⁽²⁾
4-vinyl-phenol	2334	180	>1	Pharmaceutical ⁽³⁾

E-LT-SC: Emir wine sample inoculated with strain OB 4.CM NS4 (*L. thermotolerans*) and strain OB 4.CM S1 (*S. cerevisiae*), E-SC: Emir wine sample inoculated with strain OB 4CM S1 (*S. cerevisiae*). RI: retention indices, OAV: Odour activity values calculated by dividing found concentration by threshold of the compound.

(1) Wang, X., et. al., 2017; (2) Welke, J., et. al., 2014; (3) Swiegers, J., et. al., 2005; (4) Celik, Z., et. al., 2018.

The amounts of acetates were found as 0.67 mg/L for E-A, 1.67 mg/L for E-Laff, 3.23 mg/L for E-LT-SC and 1.91 mg/L for E-SC. Although banana flavor was found in all wine samples, the amount of isoamyl acetate was higher in E-LT-SC than the others. Isoamyl acetate differ significantly among the samples of wines. In this study, these amounts of isoamyl acetate were 1.41, 2.75, and 1.77 mg/L for E-Laff, E-LT-SC, and E-SC, respectively. Zohre & Erten (2002) also mentioned that the amounts of isoamyl acetate in the mixed inoculation of *Saccharomyces cerevisiae/Candida pulcherrima* and *Saccharomyces cerevisiae/Kloeckera apiculata/Candida pulcherrima* were found 1.29 and 1.16 mg/L in the Emir wine samples, respectively. In addition, phenethyl acetate was significantly different and gave pleasant aroma in E-LT-SC. However, this aroma did not have the suprathreshold OAV in E-SC, E-A and E-Laff (OAV<1).

The amount of fatty acids was higher in E-SC than E-LT-SC, E-A and E-Laff. Especially, the concentration of hexanoic and octanoic acids were quite higher than the other compounds and significantly different among the Emir wine samples. These acids gave cheese aroma in the wines. This cheese aroma was noticed remarkably in E-SC.

The concentration of phenols was more considerable in E-LT-SC than E-SC. While 2-methoxy-4-vinylphenol gave curry spice aroma, 4-vinyl-phenol contributed pharmaceutical in both the wines. On the hand, 4-vinyl-phenol were not detected in E-A and E-Laff while 2-methoxy-4-vinylphenol was found lower level in E-A and E-Laff.

The other compounds such as tyrosol, soleron, guaiacol and acetoin were also detected. The concentration level of acetoin was obtained very low for odour in all these wine samples and it was not significantly different among these samples. Guaiacol was found detectable level in E-A and E-Laff. This aroma gave smoke odour in the wines. On the other hand, the odour threshold value of tyrosol and soleron were not found in the literature.

3.9.1.2. The Result of Volatile Compound Analysis of Red Wine

Red wines were made from Kalecik Karası grapes which were taken from Ankara at 2017. One of the red wines was prepared with strain 1 (*Hanseniaspora uvarum*) (K-HU). The other was made with strain 13 (*Saccharomyces cerevisiae*) (K-13). In order to compare, the other wines were traditional wine (K-A) made by Elnaz Seyid Monir and commercial wine made with using commercial Chr. Hansen starter culture (*Saccharomyces cerevisiae*) (K-SC).

The following three diagrams were shown the peaks of volatile compounds in GC-MS chromatogram for K-13, K-HU and K-SC, respectively (Figure 3.39-3.41).

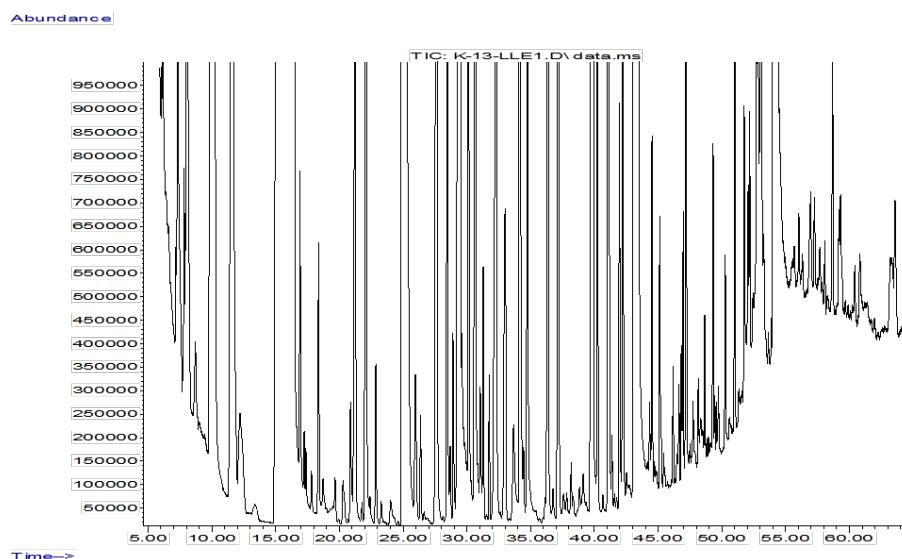


Figure 3.39. GC-MS Chromatogram of K-13 Sample

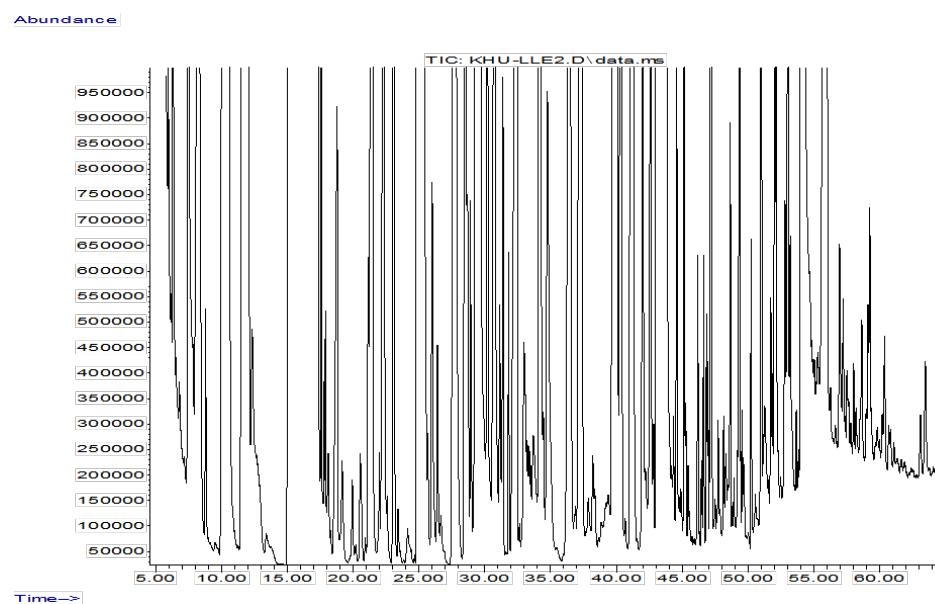


Figure 3.40. GC-MS Chromatogram of K-HU Sample

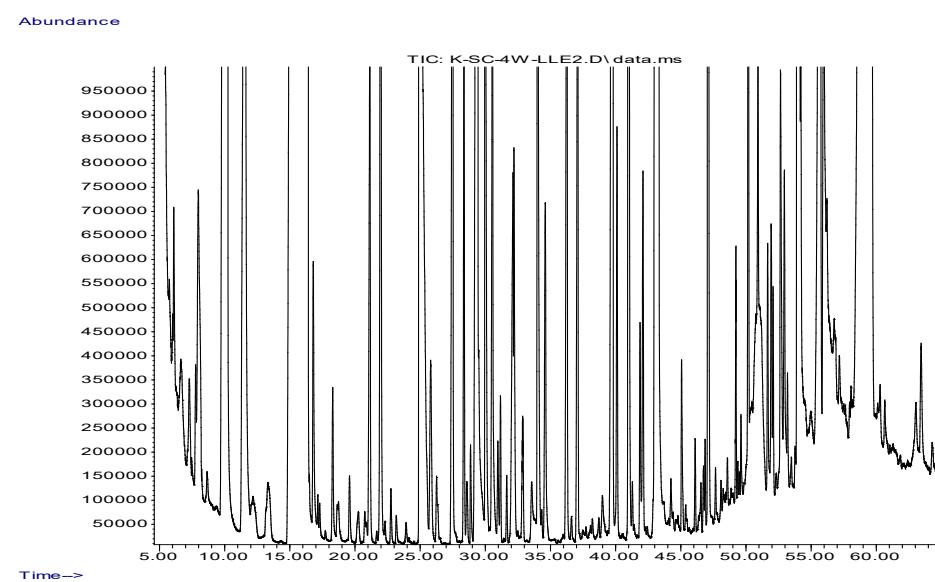


Figure 3.41. GC-MS Chromatogram of K-SC Sample

According to GS-MS results, 55 aroma compounds were identified for K-13, K-HU, K-SC and K-A (Table 3.17). These compounds included alcohols, acetates, esters, aldehydes, acids, lactones, phenols, and other compounds such as tyrosol, soleron, guaiacol, acetoin, geraniol and syringol. 52 of them were significantly different for these samples. The total concentrations of aroma compounds were found 146.83

mg/L in K-13, 139.54 mg/L in K-HU, 183.23 mg/L in K-SC and 188.28 mg/L in K-A. High alcohols and esters are the most abundant aroma in the wines. In previous studies, similar results have been reported in Dona Branca, Treixadura and Loureira wines (Falqué, Fernández, & Dubourdieu, 2002), Pinot Noir wines (Girard, Yuksel, Cliff, Delaquis, & Reynolds, 2001), Merlot and Cabernet Sauvignon wines (Kotseridis & Baumes, 2000).

The amounts of alcohols were found 130.84 mg/L, 127.28 mg/L, 165.54 mg/L and 167.13 mg/L for K-13, K-HU, K-SC and K-A respectively. In these alcohols, isobutyl alcohol, isoamyl alcohol, phenethyl alcohol and 2,3-butanediol were found very high level in these three wines. Isoamyl alcohol is an aroma compound that plays a role in the formation of the flavour of Kalecik karası wine (Sincar, 2010). Isoamyl alcohol and phenethyl alcohol had suprathreshold OAVs and gave whiskey and rose aroma, respectively in these wine samples (Table 3.19). In the previous studies, phenethyl alcohol, gave the rose aroma, was also reported to be important aroma compounds in Pinot Noir wine and Riesling wine. Moreover, fruity aroma (3-Ethoxy-1-propanol) was found higher in K-HU wine, and it was significantly different than K-13 and K-SC samples. In addition, odour of cooked vegetables was detected highly in K-HU, K-A and K-SC samples ($OAV>1$).

Esters contributes fruity and floral aroma in young wines (Rapp & Mandery, 1986). 7.16 mg/L, 4.97 mg/L, 5.40 mg/L 9.01 mg/L of esters were found in K-13, K-HU, K-SC and K-A, respectively. Ethyl 4-hydroxybutanoate and monoethyl succinate were found high amount and significantly differ among these wines. However, the odour threshold values of these aroma compounds were not found in the literature. According to OAV's, ethyl octanoate, and ethyl hexanoate affected aromas of these wines. They gave fruity aroma. Sincar (2010) also obtained a similar result in Kalecik karası wines with the application of cold maceration.

The amounts of acetates were found as 2.02 mg/L for K-13, 1.78 mg/L for K-HU, 2.47 mg/L for K-SC, 1.03 mg/L for K-A. Isoamyl acetate gave banana aroma in these wines. On the other hand, pleasant aroma (phenethyl acetate) enhanced remarkably in K-HU and K-13.

The concentrations of fatty acids were 4.30 mg/L, 3.00 mg/L, 6.54 mg/L and 5.68 in K-13, K-HU, K-SC, and K-A, respectively. Even though fatty acids may tend to have an adverse effect on wine flavor profile, they participate in the equilibrium reaction through the formation of this ester (Wang, Capone, Wilkinson, & Jeffery, 2016). Acetic acid, hexanoic acid, octanoic acid, isobutyric acid were found higher level in these wines with respect to total concentration.

The concentration of phenols was more considerable in K-SC than K-13, K-HU and K-A. 2-methoxy-4-vinylphenol gave curry spice aroma in these wines. Especially, 2-methoxy-4-vinylphenol enhanced remarkably in K-SC. Moreover, 4-vinyl-phenol was found as high odour value in K-SC (OAV>1). This gave pharmaceutical aroma in this wine.

The other compounds such as tyrosol, soleron, guaiacol, acetoin, geraniol, and syringol were also detected. Guaiacol and geraniol had suprathreshold OAVs (OAV>1). While guaiacol gave smoke flavors, geraniol contributes rose aroma for these wines. Geraniol was also a remarkable compound that was identified in orange wine and apricot wine (Kocabey, 2013).

Table 3.19. Suprathreshold Compounds for K-A, K-13, K-HU and K-SC in Wines of Kalecik Karası

Compound	RI	Odour Threshold ($\mu\text{g/L}$)	Odour Activity Values	Odour Description
Alcohols				
Isoamyl Alcohol	1236	60000	>1	Whiskey, nail polish ⁽¹⁾
3-Ethoxy-1-propanol	1389	100	>1	Fruity ⁽³⁾
Methionol	1721	500	>1	Cooked vegetable ⁽³⁾
Phenylethyl Alcohol	1916	10000	>1	Rose, pollen, perfume ⁽¹⁾
Acetates				
Isoamyl acetate	1132	30	>1	Banana, fruity, sweet ⁽¹⁾
Phenethyl acetate	1827	250	>1	Pleasant, floral ⁽¹⁾
Esters				
Ethyl octanoate	1412	5	>1	Fruity, pineapple, pear, floral ⁽¹⁾
Ethyl hexanoate	1241	5	>1	Flowery, fruity ⁽¹⁾
Aldehydes				
Nonanal	1658	2.8	>1	Citrusy, floral (3)
Phenols				
2-Methoxy-4-vinylphenol	2168	40	>1	Spices, curry ⁽³⁾
Other Compounds				
Guaiacol	1840	10	>1	Smoke, sweet, medicine ⁽³⁾
Geraniol	1844	20	>1	Roses, geranium ⁽²⁾

K-13: Kalecik karası wine sample inoculated with strain 13 (*Saccharomyces cerevisiae*), K-HU: Kalecik karası wine sample inoculated with *Hanseniaspora uvarum* K-SC: Kalecik karası wine sample inoculated with commercial *Saccharomyces cerevisiae* (Chr. Hansen), K-A: Traditional wine made by Elnaz Seyid Monir RI: retention indices, OAV: Odour activity values calculated by dividing found concentration by threshold of the compound.

(1)Wang, X., et. al., 2017; (2) Arcari, S., et. al., 2017; (3) Welke, J., et. al., 2014; (4) Celik, Z., et. al., 2018

3.9.2. The Results of Sensory Analysis of Wine

Sensory analysis was made by the degustators. They evaluated the wines by giving the points. These points change between 1 (weak) to 5 (strong).

The sensory evaluation of Emir wines was based on colour, flavour, fullness, sweetness, acidness, bitterness, astringency, final astringency and general impression characteristics. Figure 3.42 showed that E-LT-SC, made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Emir grapes, was found more desirable than others with respect to flavour and general impression.

On the other hand, E-WA-SC, made by sequential inoculation of our *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* strains in Emir grapes, was liked with sweetness.

Gobbi et al. (2013) also reported that LT-SC sequential inoculation enhanced desirable aroma in white wines. Spicy notes and acidness were found significantly different in LT-SC wine.

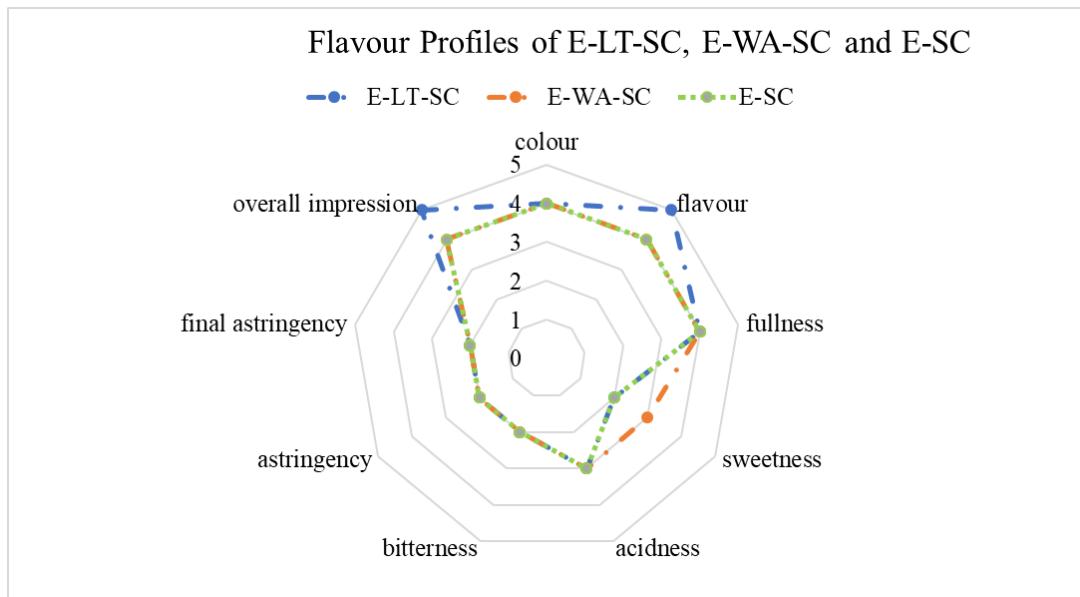


Figure 3.42. Flavour Profiles of E-LT-SC, E-WA-SC and E-SC

Figure 3.43 showed that traditional wines (E-A and E-B) were found more desirable than the commercial wine with respect to flavour and fullness, but they have not changed the overall impression score.

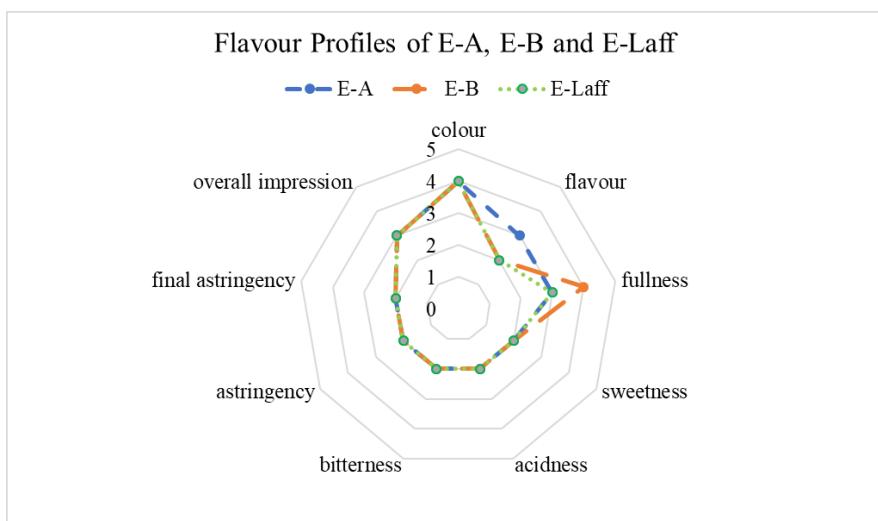


Figure 3.43. Flavour Profiles of E-A, E-B and E-Laff

Figure 3.44 showed that K-13, made by inoculation of our *Saccharomyces cerevisiae* strain in Kalecik Karası grapes, was found more desirable than others according to colour, flavour, fullness, and overall impression.

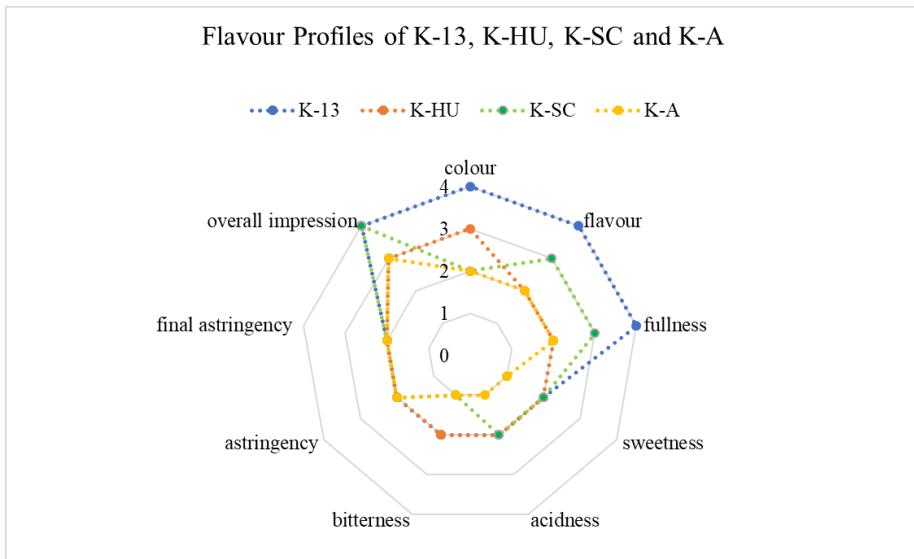


Figure 3.44. Flavour Profiles of K-13, K-HU, K-SC, and K-A

Figure 3.45 showed that O-LT-SC, made by sequential inoculation of our *Lachancea thermotolerans* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes, was found more desirable than others according to flavour, fullness, and overall impression. In

addition, acidity of O-HG-SC, made by sequential inoculation of our *Hanseniaspora guilliermondii* and *Saccharomyces cerevisiae* strains in Öküzgözü grapes was found higher than others.

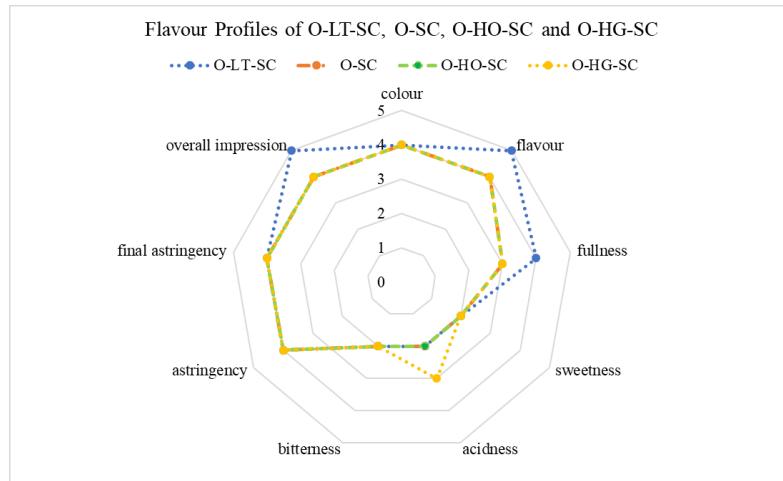


Figure 3.45. Flavour Profiles of O-LT-SC, O-SC, O-HO-SC, and O-HG-SC

Figure 3.46 showed that there was no significant difference between traditional and commercial wines of Öküzgözü grapes except flavour.

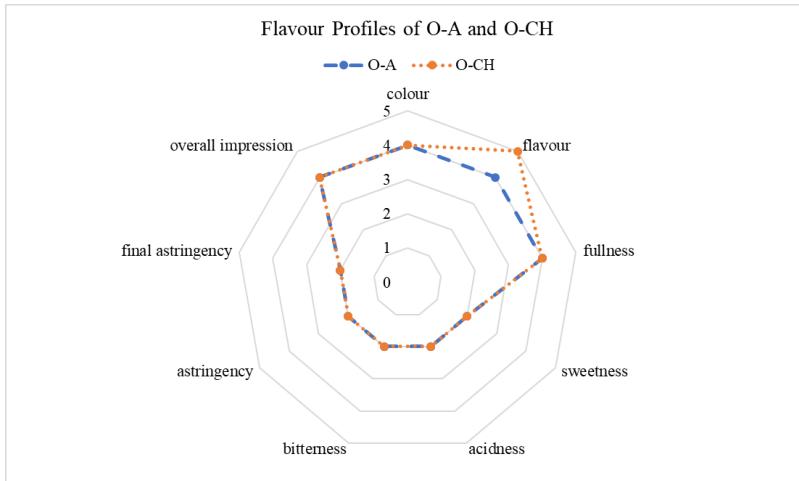


Figure 3.46. Flavour Profiles of O-A and O-CH

CHAPTER 4

CONCLUSION AND RECOMMENDATIONS

In this study, traditional wines were made from 5 type of grapes (Kalecik Karası, Öküzgözü, Boğazkere, Dimrit, and Emir) from 3 different states (Ankara, Elazığ, and Cappadocia) in Turkey. In these musts and wines, totally, 397 microorganisms were isolated. Strains which were selected according to phenotypic properties were identified by molecular (DNA sequencing for ITS region and/or D1/D2 domain) and biochemical methods (carbohydrate fermentation test). The numbers of isolated non-*Saccharomyces* and *Saccharomyces* yeasts in the culture collection were 104 and 293, respectively (Figure 4.1).

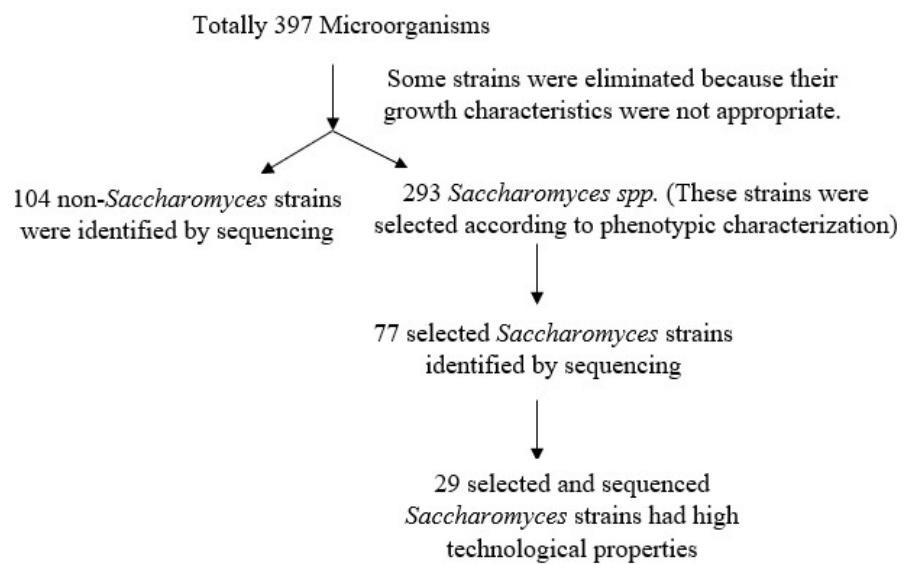


Figure 4.1. The Numbers of Isolated Non-*Saccharomyces* and *Saccharomyces* Yeasts in the Culture Collection

In this study, 16 different species were obtained as *Hanseniaspora guilliermondii*, *Hanseniaspora opuntiae*, *Hanseniaspora uvarum*, *Rhodotorula mucilaginosa*, *Wickerhamomyces anomalus*, *Metschnikowia aff. fructicola*, *Metschnikowia fructicola*, *Lachancea thermotolerans*, *Solicoccozyma aeria*, *Metschnikowia aff. pulcherrima*, *Metschnikowia pulcherrima*, *Metschnikowia sinensis*, *Metschnikowia chrysoperlae*, *Starmerella bacillaris*, *Saccharomyces cerevisiae* and *Saccharomyces cerevisiae/paradoxus*. However, *Metschnikowia sinensis* was identified as *Metschnikowia pulcherrima* and *Saccharomyces cerevisiae/paradoxus* as *Saccharomyces cerevisiae* by D1/D2 sequencing.

Phenotypic characterization was performed for isolated and identified strains. Firstly, alcohol tolerance and H₂S production of *Saccharomyces* yeasts were tested and having a high alcohol tolerance and low H₂S producing isolates were selected for each grape variety. SO₂, pH, temperature tolerance and carbohydrate fermentation abilities of selected isolates were also tested. According to these properties, the selection table was created (Table 3.17). General properties of these selected yeasts were 13% and above 13% alcohol tolerance, low H₂S production (1 or 2), resistance to pH (3, 4 and 6) (except DA 4. CM S3 strain), growth at 28°C and 37°C, fermentation of sucrose and maltose, dissimilation of maltose and lactose. These *Saccharomyces* isolates with high technological properties for wine production were selected as starter cultures.

In addition, non-*Saccharomyces* isolates were generally found low alcohol tolerance (below 10%), high SO₂ tolerance (200 mg/L), and intermediate and high H₂S production (3 and above 3). On the other hand, some of them were found high alcohol tolerance (15%) such as OA 4.CM S3 (*Wickerhamomyces anomalus*), OA 4.CM NS4 (*Hanseniaspora opuntiae*), KB 0.CM NS5 (*Wickerhamomyces anomalus*), DA 0.CM NS2 (*Metschnikowia aff. pulcherrima*). In addition, KA 2.NM NS1 (*Hanseniaspora opuntiae*), KA 4.NM NS5 (*Hanseniaspora opuntiae*) and DA 4.CM NS3 (*Metschnikowia pulcherrima*) produced low level of H₂S (2).

White and red wines were made using some isolates in our culture collection as a starter culture in order to analyze volatile compounds and flavour profiles. According to flavour profiles, Emir and Öküzgözü wines with *Lachancea thermotolerans* strain were found successful by degustators. Our *Lachancea thermotolerans* strain was randomly chosen from Öküzgözü grapes and inoculated sequentially with our *Saccharomyces cerevisiae* as starter cultures to produce wine. The total aroma concentrations were found higher in E-LT-SC (131.7 mg/L) than E-SC (113.2 mg/L) with respect to GS-MS results. 3-Ethoxy-1-propanol (fruity), isoamyl acetate (banana) and phenylethyl acetate (pleasant aroma) were higher in E-LT-SC wine while ethyl octanoate (pineapple), ethyl hexanoate (flowery), and hexanoic acid (cheese odour) were found highly in E-SC.

K-13 was found more successful than others by the degustators while the total concentrations of aroma compounds were 146.83 mg/L in K-13, 139.54 mg/L in K-HU and 183.23 mg/L in K-SC. 3-Ethoxy-1-propanol (fruity), methionol (cooked vegetable) and phenethyl acetate (pleasant) were found higher in K-HU, while guaiacol (smoke) was high level in K-13. On the other hand, isoamyl alcohol (whiskey aroma), phenylethyl alcohol (rose), isoamyl acetate (banana), ethyl octanoate (pineapple), ethyl hexanoate (flowery), 2-methoxy-4-vinylphenol (curry spice), and geraniol (rose) were found in all wine's samples ($OAV > 1$).

This study revealed native non-*Saccharomyces* and *Saccharomyces* yeast populations in Turkey. Selected strains of starter cultures for authentic wine productions can be used to improve aroma and quality of native wines.

REFERENCES

- Andorrà, I., Berradre, M., Rozès, N., Mas, A., Guillamón, J. M., & Esteve-Zarzoso, B. (2010). Effect of pure and mixed cultures of the main wine yeast species on grape must fermentations. *European Food Research and Technology*, 231(2), 215–224. <https://doi.org/10.1007/s00217-010-1272-0>
- Arbefeuille, S., Harris, A., & Ferrieri, P. (2017). Comparison of sequencing the D2 region of the large subunit ribosomal RNA gene (MicroSEQ ®) versus the internal transcribed spacer (ITS) regions using two public databases for identification of common and uncommon clinically relevant fungal species. *Journal of Microbiological Methods*, 140, 40–46. <https://doi.org/10.1016/j.mimet.2017.06.015>
- Arcari, S. G., Caliari, V., Sganzerla, M., & Godoy, H. T. (2017). Volatile composition of Merlot red wine and its contribution to the aroma: Optimization and validation of analytical method. *Talanta*, 174, 752–766. doi:10.1016/j.talanta.2017.06.074
- Assis, M. O., Pereira, A., Santos, C., Rosa, C. A., Eugênia, M., & Mamede, D. O. (2014). Impact of a Non- *Saccharomyces* Yeast Isolated in the Equatorial Region in the Acceptance of Wine Aroma. *Food and Nutrition Sciences*, (April), 759–769. <https://doi.org/10.4236/fns.2014.59086>
- Azzolini, M., Tosi, E., Lorenzini, M., Finato, F., & Zapparoli, G. (2015). Contribution to the aroma of white wines by controlled *Torulaspora delbrueckii* cultures in association with *Saccharomyces cerevisiae*. *World Journal of Microbiology and Biotechnology*, 31(2), 277–293. <https://doi.org/10.1007/s11274-014-1774-1>
- Bağder, S. (2008). Türkiye'de Değişik Şarap Bölgelerinden İzole Edilmiş Şarap Mayalarının Teknolojik Özellikleri. (*Master Thesis*) Ankara University Ankara, Turkey. <https://doi.org/10.1017/CBO9781107415324.004>
- Baker, J. & Clarke, J. R. (2012). Wine flavour. *Wine Flavour Chemistry, Second Edition*. <https://doi.org/10.1002/9780470995594>
- Balkan Pazar* (n.d.). Retrieved from http://www.balkanpazar.org/tr_sarap.asp

- Baleiras-Couto, M. M., & Eiras-Dias, J. E. (2006). Detection and identification of grape varieties in must and wine using nuclear and chloroplast microsatellite markers. *Analytica Chimica Acta*, 563(1-2 SPEC. ISS.), 283–291. <https://doi.org/10.1016/j.aca.2005.09.076>
- Barnett J.A., Payne R.W. and Yarrow D. (2000) *Yeasts: Characteristics and Identification*, 3rd edn. Cambridge University Press Cambridge, UK
- Beckner, M. E., Carlin, S., Jacobson, D., Weighill, D., Divol, B., Conterno, L., Vrhovsek, U. (2015). Early fermentation volatile metabolite profile of non-*Saccharomyces* yeasts in red and white grape must : A targeted approach. *LWT - Food Science and Technology*, 64(1), 412–422. <https://doi.org/10.1016/j.lwt.2015.05.018>
- Boekhout, T., & Robert, V. (2014). *Yeasts in food: Beneficial and detrimental aspects*. London: Elsevier.
- Belloch, C., Barrio, E., Garcia, M. D., & Querol, A. (1998). Phylogenetic reconstruction of the yeast genus *Kluyveromyces* : Restriction map analysis of the 5.8S rRNA gene and the two ribosomal internal transcribed spacers. *Systematic and Applied Microbiology*, 21, 266–273. [https://doi.org/10.1016/S0723-2020\(98\)80032-5](https://doi.org/10.1016/S0723-2020(98)80032-5)
- Bouchilloux, P., Darriet, P., & Henry, R. (1998). Identification of volatile and powerful odorous thiols in bordeaux red wine varieties. *Journal of Agricultural and Food Chemistry*, 8561(97), 3095–3099. <https://doi.org/10.1021/jf971027d>
- Bowers, J., Boursiquot, J.M., This, P., Chu, K., Johansson, H., & Meredith, C. (1999). Historical genetics: the parentage of Chardonnay, Gamay, and other wine grapes of Northeastern France, *Ed. Science*, 285, 1562-1565. Retrieved from <http://prodinra.inra.fr/record/56459>
- Cabaroglu, T., Canbas, A., Baumes, R., Bayonove, C., Lepoutre, J. P., & Günata, Z. (1997). Aroma composition of a white wine of *Vitis vinifera* L. cv. Emir as affected by skin contact. *Journal of Food Science*, 62(4), 680–683. <https://doi.org/10.1111/j.1365-2621.1997.tb15434.x>
- Cañas, P. M. I., García, A. T. P., & Romero, E. G. (2011). Enhancement of flavour properties in wines using sequential inoculations of non-*Saccharomyces* (*Hansenula* and *Torulaspora*) and *Saccharomyces* yeast. *VITIS - Journal of Grapevine Research*, 50(4), 177–182. Retrieved from <http://pub.jki.bund.de/index.php/VITIS/article/view/4088>

- Capece, A., Romaniello, R., Siesto, G., Pietrafesa, R., Massari, C., Poeta, C., & Romano, P. (2010). Selection of indigenous *Saccharomyces cerevisiae* strains for Nero d'Avola wine and evaluation of selected starter implantation in pilot fermentation. *International Journal of Food Microbiology*, 144(1), 187–192. <https://doi.org/10.1016/j.ijfoodmicro.2010.09.009>
- Cappello, M. S., Bleve, G., Grieco, F., Dellaglio, F., & Zacheo, G. (2004). Characterization of *Saccharomyces cerevisiae* strains isolated from must of grape grown in experimental vineyard. *Journal of Applied Microbiology*, 97(6), 1274–1280. <https://doi.org/10.1111/j.1365-2672.2004.02412.x>
- Carrau, F. M., Medina, K., Boido, E., Farina, L., Gaggero, C., Dellacassa, E., Henschke, P. A. (2005). De novo synthesis of monoterpenes by *Saccharomyces cerevisiae* wine yeasts. *FEMS Microbiology Letters*, 243, 107–115. <https://doi.org/10.1016/j.femsle.2004.11.050>
- Chavan, P., Mane, S., Kulkarni, G., Shaikh, S., Ghormade, V., Nerkar, D. P., Deshpande, M. V. (2009). Natural yeast flora of different varieties of grapes used for wine making in India. *Food Microbiology*, 26(8), 801–808. <https://doi.org/10.1016/j.fm.2009.05.005>
- Christoph, M. E., & Thomas, H.-K. (2001). Identification of *Brettanomyces/Dekkera* species based on polymorphism in the rRNA internal transcribed spacer region. *American Journal of Enology and Viticulture*, 52(3), 241–246.
- Ciani, M. and Faticanti, F. (1999). Selective sugar consumption by apiculate yeasts. *Lett. Appl. Microbiol.* 28: 203–206.
- Ciani, M., & Ferraro, L. (1998). Combined use of immobilized *Candida stellata* cells and *Saccharomyces cerevisiae* to improve the quality of wines. *Journal of Applied Microbiology*, 85(2), 247–254. <https://doi.org/10.1046/j.1365-2672.1998.00485.x>
- Ciani, Maurizio, Beco, L., & Comitini, F. (2006). Fermentation behaviour and metabolic interactions of multistarter wine yeast fermentations. *International Journal of Food Microbiology*, 108(2), 239–245. <https://doi.org/10.1016/j.ijfoodmicro.2005.11.012>
- Clemente-Jimenez, J. M., Mingorance-Cazorla, L., Martínez-Rodríguez, S., Las Heras-Vázquez, F. J., & Rodríguez-Vico, F. (2005). Influence of sequential yeast mixtures on wine fermentation. *International Journal of Food Microbiology*, 98(3), 301–308. <https://doi.org/10.1016/j.ijfoodmicro.2004.06.007>

- Clemente-Jimenez, Josefa María, Mingorance-Cazorla, L., Martínez-Rodríguez, S., Las Heras-Vázquez, F. J., & Rodríguez-Vico, F. (2004). Molecular characterization and oenological properties of wine yeasts isolated during spontaneous fermentation of six varieties of grape must. *Food Microbiology*, 21(2), 149–155. [https://doi.org/10.1016/S0740-0020\(03\)00063-7](https://doi.org/10.1016/S0740-0020(03)00063-7)
- Cocolin, L., Pepe, V., Comitini, F., Comi, G., & Ciani, M. (2004). Enological and genetic traits of *Saccharomyces cerevisiae* isolated from former and modern wineries. *FEMS Yeast Research*, 5(3), 237–245. <https://doi.org/10.1016/j.femsyr.2004.08.005>
- Comitini, F., Gobbi, M., Domizio, P., Romani, C., Lencioni, L., Mannazzu, I., & Ciani, M. (2011). Selected non-*Saccharomyces* wine yeasts in controlled multistarter fermentations with *Saccharomyces cerevisiae*. *Food Microbiology*, 28(5), 873–882. <https://doi.org/10.1016/j.fm.2010.12.001>
- Cordero-bueso, G., Mangieri, N., Maghradze, D., Foschino, R., Valdetara, F., Cantoral, J. M., & Vigentini, I. (2017). Wild grape-associated yeasts as promising biocontrol agents against *Vitis vinifera* fungal pathogens, 8(November). <https://doi.org/10.3389/fmicb.2017.02025>
- Çelik, Z. D., Erten, H., Darıcı, M., & Cabaroğlu, T. (2017). Molecular characterization and technological properties of wine yeasts isolated during spontaneous fermentation of *Vitis vinifera* L.cv. Narince grape must grown in ancient wine making area Tokat, Anatolia. *BIO Web of Conferences*, 9, 02017. <https://doi.org/10.1051/bioconf/20170902017>
- D'Agata, I. (2014) Native wine grapes of Italy. Ed. *University of California Press*.
- Dias, L., Dias, S., Sancho T., Stender, Querol H., A., Malfeito-Ferreira M., and Loureiro, V. (2003). Identification of yeasts isolated from wine-related environments and capable of producing 4-ethylphenol. *Food Microbiol.* 20: 567–574.
- Don, B. (2006). *Handbook of Enology Volume 1 The Microbiology of Wine and Vinifications* 2 nd Edition (Vol. 1).
- Ebeler, S. E., & Thorngate, J. H. (2009). *Wine chemistry and flavor : Looking into the Crystal Glass*, 8098–8108. <https://doi.org/10.1021/jf9000555>

- Englezos, V., Rantsiou, K., Torchio, F., Rolle, L., Gerbi, V., & Cocolin, L. (2015). Exploitation of the non-*Saccharomyces* yeast *Starmerella bacillaris* (synonym *Candida zemplinina*) in wine fermentation: Physiological and molecular characterizations. *International Journal of Food Microbiology*, 199, 33–40. <https://doi.org/10.1016/j.ijfoodmicro.2015.01.009>
- Erten, H. (2002). Relations between elevated temperatures and fermentation behaviour of *Kloeckera apiculata* and *Saccharomyces cerevisiae* associated with winemaking in mixed cultures. *World Journal of Microbiology and Biotechnology*, 18(4), 377–382. <https://doi.org/10.1023/a:1015221406411>
- Erten, H., & Tanguler, H. (2010). Influence of *Williopsis saturnus* yeasts in combination with *Saccharomyces cerevisiae* on wine fermentation. *Letters in Applied Microbiology*, 50(5), 474–479. <https://doi.org/10.1111/j.1472-765X.2010.02822.x>
- Escudero, A., Campo, E., Fariña, L., Cacho, J., & Ferreira, V. (2007). Analytical characterization of the aroma of five premium red wines. Insights into the role of odor families and the concept of fruitiness of wines. *Journal of Agricultural and Food Chemistry*, 55(11), 4501–4510. doi:10.1021/jf0636418
- Esteve-Zarzoso, B., Belloch, C., Uruburu, F., & Querol, A. (2009). Identification of yeasts by RFLP analysis of the 5.8S rRNA gene and the two ribosomal internal transcribed spacers. *International Journal of Systematic Bacteriology*, 49(1), 329–337. <https://doi.org/10.1099/00207713-49-1-329>
- Esteve-Zarzoso, B., Peris-Toran, M. J., Garcia-Maiquez, E., Uruburu, F., & Querol, A. (2001). Yeast population dynamics during the fermentation and biological aging of sherry wines, 67(5), 2056–2061. <https://doi.org/10.1128/AEM.67.5.2056>
- Fang, Y., & Qian, M. C. (2016). Development of C₁₃-norisoprenoids , carotenoids and other volatile compounds in *Vitis vinifera L . Cv . Pinot noir* grapes. *Food Chemistry*, 192, 633–641. <https://doi.org/10.1016/j.foodchem.2015.07.050>
- FAO (2012). Retrieved from <http://www.fao.org/home/en/>
- Fazzalari, F. A. (Ed.). (1978). Compilation of odor and taste threshold values data. doi: 10.1520/ds48a-eb

- Fernandez-Espinar, M. T., Esteve-Zarzoso, B., Querol, A., & Barrio, E. (2000). RFLP analysis of the ribosomal internal transcribed spacers and the 5.8S rRNA gene region of the genus *Saccharomyces*: a fast method for species identification and the differentiation of flor yeasts. *Kl*, 78, 87–97. <https://doi.org/10.1023/B>
- Fernandez-Espinar, M. T., Llopis, S., Querol, A., & Barrio, E. (2011). Molecular Identification and Characterization of Wine Yeasts. <https://doi.org/10.1016/B978-0-12-375021-1.10005-0>
- Fleet, G. H. (2003). Yeast interactions and wine flavour. *International Journal of Food Microbiology*, 86(1–2), 11–22. [https://doi.org/10.1016/S0168-1605\(03\)00245-9](https://doi.org/10.1016/S0168-1605(03)00245-9)
- Fuglsang, K. C., & Edwards, C. G. (2007). *Wine microbiology: Practical applications and procedures*. <https://doi.org/10.1007/978-0-387-33349-6>
- Gaillardin C. and Heslot H. (1987) *Lalevure*. LaRecherche, 188 (18) 586–596.
- Galet, P. (2015) Dictionnaire encyclopédique des cépages et de leurs synonymes. *Ed. Libre&Solidaire*.
- Galet, P. (1990) Cépages et vignobles de France. *Ed. Ministère de la Recherche et de la Technologie*, Vol II.
- Garavaglia, J., Schneider, R. de C. de S., Camargo Mendes, S. D., Welke, J. E., Zini, C. A., Caramão, E. B., & Valente, P. (2015). Evaluation of *Zygosaccharomyces bailii* BCV 08 as a co-starter in wine fermentation for the improvement of ethyl esters production. *Microbiological Research*, 173, 59–65. <https://doi.org/10.1016/j.micres.2015.02.002>
- García, M., Esteve-zarzoso, B., Crespo, J., Cabellos, J. M., & Arroyo, T. (2017). Yeast monitoring of wine mixed or sequential fermentations made by native strains from D.O.“Vinos de Madrid ” using real-time quantitative PCR, 8(December), 1–15. <https://doi.org/10.3389/fmicb.2017.02520>
- Ghosh, S. (2017). Metagenomic screening of cell wall hydrolases , their anti-fungal activities and potential role in wine fermentation. (March 2015). <https://doi.org/10.13140/RG.2.2.28479.71846>
- Girard, B., Yuksel, D., Cliff, M. A., Delaquis, P., & Reynolds, A. G. (2001). Vinification effects on the sensory, colour and GC profiles of Pinot noir wines from British Columbia. *Food Research International*, 34(6), 483–499. [https://doi.org/10.1016/S0963-9969\(00\)00177-0](https://doi.org/10.1016/S0963-9969(00)00177-0)

- Gobbi, M., Comitini, F., Domizio, P., Romani, C., Lencioni, L., Mannazzu, I., & Ciani, M. (2013). *Lachancea thermotolerans* and *Saccharomyces cerevisiae* in simultaneous and sequential co-fermentation: A strategy to enhance acidity and improve the overall quality of wine. *Food Microbiology*, 33(2), 271–281. <https://doi.org/10.1016/j.fm.2012.10.004>
- Goldhawk, B., Kahlon, M., Lotto, J., & Deeg, C. M. (2016). Yeasts from greenhouse grapes show less phenotypic and genetic diversity than yeasts from vineyard grapes when isolated from grape crush cultured in liquid media, 2(June), 8–15.
- González-Arenzana, L., Garijo, P., Berlanas, C., López-Alfaro, I., López, R., Santamaría, P., & Gutiérrez, A. R. (2017). Genetic and phenotypic intraspecific variability of non-*Saccharomyces* yeasts populations from La Rioja winegrowing region (Spain). *Journal of Applied Microbiology*, 122(2), 378–388. <https://doi.org/10.1111/jam.13341>
- Guillamón, J. M., Sabaté, J., Barrio, E., Cano, J., & Querol, A. (1998). Rapid identification of wine yeast species based on RFLP analysis of the ribosomal internal transcribed spacer (ITS) region. *Archives of Microbiology*, 169(5), 387–392. <https://doi.org/10.1007/s002030050587>
- Guimaraes, T. M., Moriel, D. G., Machado, I. P., Fadel Picheth, C. M. T., Bonfim, T. (2006). Isolation and characterization of *Saccharomyces cerevisiae* strains of winery interest. *Revista Brasileira de Ciencias Farmaceuticas*, 42(1), 119–126. <https://doi.org/10.1590/S1516-93322006000100013>
- Henick-Kling, T., Edinger, W., Daniel, P., & Monk, P. (1998). Selective effects of sulfur dioxide and yeast starter culture addition on indigenous yeast populations and sensory characteristics of wine. *Journal of Applied Microbiology*, 84, 865–876. <https://doi.org/10.1046/j.1365-2672.1998.00423.x>
- Herraiz, T., Reglero, G., Martin-Alvarez, P. J., Herraiz, M., & Cabezudo, M. D. (1991). Identification of aroma components of Spanish ‘Verdejo’ wine. *Journal of the Science of Food and Agriculture*, 55(1), 103-116. doi:10.1002/jsfa.2740550111
- Hesham, A. E., & Hashem, M. M. (2011). Molecular genetic identification of yeast strains isolated from egyptian soils for solubilization of inorganic phosphates and growth promotion of corn plants. *J. Microbial. Biotechnol.*, 21(November 2010), 55–61. <https://doi.org/10.4014/jmb.1006.06045>

- Hesham, A. E. L., Wambui, V., Ogola J.O., H., & Maina, J. M. (2014). Phylogenetic analysis of isolated biofuel yeasts based on 5.8S-ITS rDNA and D1/D2 26S rDNA sequences. *Journal of Genetic Engineering and Biotechnology*, 12(1), 37–43. <https://doi.org/10.1016/j.jgeb.2014.01.001>
- Hidalgo, L. (2002) Tratado de Viticultura General. *Ed. Mundiprensa*.
- Hierro, N., González, Á., Mas, A., & Guillamón, J. M. (2006). Diversity and evolution of non-*Saccharomyces* yeast populations during wine fermentation: Effect of grape ripeness and cold maceration. *FEMS Yeast Research*, 6(1), 102–111. <https://doi.org/10.1111/j.1567-1364.2005.00014.x>
- Hu, K., Zhu, X. L., Mu, H., Ma, Y., Ullah, N., & Tao, Y. S. (2016). A novel extracellular glycosidase activity from *Rhodotorula mucilaginosa*: ITS application potential in wine aroma enhancement. *Letters in Applied Microbiology*, 62(2), 169–176. <https://doi.org/10.1111/lam.12527>
- Hyma, K. E., Saerens, S. M., Verstrepen, K. J., & Fay, J. C. (2011). Divergence in wine characteristics produced by wild and domesticated strains of *Saccharomyces cerevisiae*. *FEMS Yeast Research*, 11(7), 540-551. doi:10.1111/j.1567-1364.2011.00746.x
- International Organisation of Vine and Wine.* (2017). Retrieved from <http://www.oiv.int/>
- Jolly, N. P., Augustyn, O. P. H., & Pretorius, I. S. (2006). The role and use of non-*Saccharomyces* yeasts in wine production. *South African Journal of Enology & Viticulture*, 27(1). <https://doi.org/10.21548/27-1-1475>
- Karabat, S. (2014). *Dünya ve Türkiye Bağcılığı*. Retrieved from <http://apelasyon.com/Yazi/33-dunya-ve-turkiye-bagciliği>
- Kim, D. H., Hong, Y. A., & Park, H. D. (2008). Co-fermentation of grape must by *Issatchenka orientalis* and *Saccharomyces cerevisiae* reduces the malic acid content in wine. *Biotechnology Letters*, 30(9), 1633–1638. <https://doi.org/10.1007/s10529-008-9726-1>
- King, A., & Dickinson, J. R. (2000). Biotransformation of monoterpane alcohols by *Saccharomyces cerevisiae*, *Torulaspora delbrueckii* and *Kluyveromyces lactis*. *Yeast*, (16), 499–506.
- Kish, S., Sharf, R., & Margalith, P. (1983). A note on a selective medium for wine yeasts. *Journal of Applied Microbiology*, 55(1), 177-179.

- Kocabey, N. (2013). Arapgir'de Yetiştirilen Karaoğlan ve Aşık Beyazı Üzümlerinin ve Bu Üzümlerden Elde Edilen Şarapların Fenol Bileşiklerinin ve Aroma Maddelerinin Belirlenmesi. (*Master Thesis*) Inonu University, Malatya, Turkey
- Kotseridis, Y., & Baumes, R. (2000). Identification of impact odorants in Bordeaux red grape juice, in the commercial yeast used for its fermentation, and in the produced wine. *Journal of Agricultural and Food Chemistry*, 48(2), 400–406. <https://doi.org/10.1021/jf990565i>
- Kurtzman, C. P. (2011). Discussion of teleomorphic and anamorphic ascomycetous yeasts and yeast-like taxa. *The Yeasts* (Vol. 2). Elsevier B.V. <https://doi.org/10.1016/B978-0-444-52149-1.00013-6>
- Kurtzman, C. P., Fell, J. W., Boekhout, T., & Robert, V. (n.d.). Methods for Isolation, Phenotypic Characterization and Maintenance of Yeasts. *The Yeasts, A Taxonomic Study*. Elsevier B.V. <https://doi.org/10.1016/B978-0-444-52149-1.00007-0>
- Kurtzman, C. P., & Robnett, C. J. (1998). Identification and phylogeny of ascomycetous yeasts from analysis of nuclear large subunit (26S) ribosomal DNA partial sequences. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 73(4), 331–371. <https://doi.org/10.1023/A:1001761008817>
- Lacombe, T., (2012) Contribution à l'étude de l'histoire évolutive de la vigne cultivée (*Vitis vinifera L.*) par l'analyse de la diversité génétique neutre et de gènes d'intérêt, Montpellier, France.
- Lambrechts, M., & Pretorius, I. S. (2000). Yeast and its importance to wine aroma-a review. *South African Journal of Enology & Viticulture*, (21), 97–129.
- Li, S. S., Cheng, C., Li, Z., Chen, J. Y., Yan, B., Han, B. Z., & Reeves, M. (2010). Yeast species associated with wine grapes in China. *International Journal of Food Microbiology*, 138(1–2), 85–90. <https://doi.org/10.1016/j.ijfoodmicro.2010.01.009>
- Liu, N., Song, Y., Dang, G., Ye, D., Gong, X., & Liu, Y. (2015). Effect of wine closures on the aroma properties of chardonnay wines after four years of storage. *South African Journal of Enology and Viticulture*, 36(3). doi:10.21548/36-3-963
- Maul, E., & Topfer, R., Vitis International Variety Catalogue (VIVC): A cultivar database referenced by genetic profiles and morphology, BIO Web of conferences Vol. 5, Article No. 01009, 2015

- Macías, M., Manso, A., Orellana, C., Velasco, H., Caballero, R., & Chamizo, J. (2012). Acetic acid detection threshold in synthetic wine samples of a portable electronic nose sensors, 13(1), 208-220. doi:10.3390/s130100208
- Mateo, J. J., & Jimenez, M. (2000). Monoterpenes in grape juice and wines. *J. Chromatogr. A*, 881, 557–567
- Medina, K., Boido, E., Fariña, L., Gioia, O., Gomez, M. E., Barquet, M., Carrau, F. (2013). Increased flavour diversity of Chardonnay wines by spontaneous fermentation and co-fermentation with *Hanseniaspora vineae*. *Food Chemistry*, 141(3), 2513–2521. <https://doi.org/10.1016/j.foodchem.2013.04.056>
- Meredith, C., & Boursiquot, J.M. (2008) Origins and importance of Syrah around the world, in: Proceedings of the International Syrah Symposium, 17-20. Presented at the International Syrah Symposium, Lyon (France). Retrieved from <http://prodinra.inra.fr/record/184458>
- Mendoza, L. M., De Nadra, M. C. M., & Farias, M. E. (2007). Kinetics and metabolic behavior of a composite culture of *Kloeckera apiculata* and *Saccharomyces cerevisiae* wine related strains. *Biotechnology Letters*, (29), 1057–1063. <https://doi.org/10.1007/s10529-007-9355-0>
- Meyer, S.A., Payne, R.W. and Yarrow, D. (1998) *Candida* Berkout. In: The Yeasts. C.P. Kurtzman and J.W. Fell (Eds.), 4th edition, Chapter 64, pp. 454–573. Elsevier, New York, NY.
- Moreira, N., Mendes, F., Guedes de Pinho, P., Hogg, T., & Vasconcelos, I. (2008). Heavy sulphur compounds, higher alcohols and esters production profile of *Hanseniaspora uvarum* and *Hanseniaspora guilliermondii* grown as pure and mixed cultures in grape must. *International Journal of Food Microbiology*, 124(3), 231–238. <https://doi.org/10.1016/j.ijfoodmicro.2008.03.025>
- Moreira, Nathalie, Mendes, F., Hogg, T., & Vasconcelos, I. (2005). Alcohols, esters and heavy sulphur compounds production by pure and mixed cultures of apiculate wine yeasts. *International Journal of Food Microbiology*, 103(3), 285–294. <https://doi.org/10.1016/j.ijfoodmicro.2004.12.029>
- Moreira, N., Pinho, P. G., Santos, C., & Vasconcelos, I. (2010). Volatile sulphur compounds composition of monovarietal white wines. *Food Chemistry*, 123(4), 1198-1203. doi:10.1016/j.foodchem.2010.05.086

- Morinaga, K., (2001) Grape production in Japan, in: Papademetriou, M.K. & Dent, F.J., *Grape production in the Asia-Pacific region*, Ed. FAO, pp. 38-52.
- Nwaiwu, O. (2019). Phylogeny of three palmwine yeasts genera. recent advances in phylogenetics. doi:10.5772/intechopen.79958
- Nelsen M.P. (2018) Primer maps. <http://sites.google.com/site/mpnelsen/primer-maps>.
- Nikolaou, E., Soufleros, E. H., Bouloumpasi, E., & Tzanetakis, N. (2006). Selection of indigenous *Saccharomyces cerevisiae* strains according to their oenological characteristics and vinification results. *Food Microbiology*, 23(2), 205–211. <https://doi.org/10.1016/j.fm.2005.03.004>
- Nurgel, C., Erten, H., Canbas, A., Cabaroglu, T., & Sellı, S. (2005). Yeast flora during the fermentation of wines made from *Vitis vinifera L. cv. Emir* and *Kalecik Karası* grown in Anatolia. *World Journal of Microbiology and Biotechnology*, 21(6–7), 1187–1194. <https://doi.org/10.1007/s11274-005-1106-6>
- Padilla, B., Gil, J. V., & Manzanares, P. (2016). Past and future of non-*Saccharomyces* yeasts: From spoilage microorganisms to biotechnological tools for improving wine aroma complexity. *Frontiers in Microbiology*, 7(MAR), 1–20. <https://doi.org/10.3389/fmicb.2016.00411>
- Pallmann, C. L., Brown, J. A., Olineka, T. L., Cocolin, L., Mills, D. A., & Bisson, L. F. (2001). Use of WL medium to profile native flora fermentations. *Am. J. Enol. Vitic.*, 52(3), 198-203
- Parker, M., Osidacz, P., Baldock, G. A., Hayasaka, Y., Black, C. A., Pardon, K. H., . . . Francis, I. L. (2012). Contribution of Several Volatile Phenols and Their Glycoconjugates to Smoke-Related Sensory Properties of Red Wine. *Journal of Agricultural and Food Chemistry*, 60(10), 2629-2637. doi:10.1021/jf2040548
- Pastor, A., Huerta, T., Mateo, J. J., & Jimé, M. (2001). Yeast starter cultures affecting wine fermentation and volatiles, 34.
- Pham, T., Wimalasena, T., Box, W., G., Koivuranta, K., Storgards, E., Smart, K. A., & Gibson, B. R. (2011). Evaluation of ITS PCR and RFLP for differentiation and identification of brewing yeast and brewery “wild” yeast contaminants. *Journal of the Institute of Brewing*, 117(4), 556–568. <https://doi.org/10.1002/j.2050-0416.2011.tb00504.x>

- Pina, C., Santos, C., Couto, J. A., & Hogg, T. (2004). Ethanol tolerance of five non-*Saccharomyces* wine yeasts in comparison with a strain of *Saccharomyces cerevisiae* - Influence of different culture conditions. *Food Microbiology*, 21(4), 439–447. <https://doi.org/10.1016/j.fm.2003.10.009>
- Pretorius, I. S. (n.d.). The tailoring of grapevine cultivars and wine yeast strains for a market-directed and quality-focused wine industry : Novel approaches to the ancient art of winemaking. *Yeast*.
- Rantsiou, K., Dolci, P., Giacosa, S., Torchio, F., Tofalo, R., Torriani, S., Cocolin, L. (2012). *Candida zemplinina* can reduce acetic acid produced by *Saccharomyces cerevisiae* in sweet wine fermentations. *Applied and Environmental Microbiology*, 78(6), 1987–1994. <https://doi.org/10.1128/aem.06768-11>
- Rapp, A., & Mandery, H. (1986). Wine aroma. *Experientia*, 42(8), 873–884.
- Rapp, A., & Versini, G. (1995). Influence of nitrogen compounds in grapes on aroma compounds of wines, 1659–1694.
- Rodriguez, M. E., Lopes, C. A., Van Broock, M., Valles, S., Ramon, D., & Caballero, A. C. (2004). Screening and typing of Patagonian wine yeasts for glycosidase activities, 84–95. <https://doi.org/10.1046/j.1365-2672.2003.02032.x>
- Rojas, V., Gil, J. V., Piñaga, F., & Manzanares, P. (2003). Acetate ester formation in wine by mixed cultures in laboratory fermentations. *International Journal of Food Microbiology*, 86(1–2), 181–188. [https://doi.org/10.1016/S0168-1605\(03\)00255-1](https://doi.org/10.1016/S0168-1605(03)00255-1)
- Romancino, D. P., Maio, S. Di, Muriella, R., & Oliva, D. (2008). Analysis of non-*Saccharomyces* yeast populations isolated from grape musts from Sicily (Italy), 105, 2248–2254. <https://doi.org/10.1111/j.1365-2672.2008.03894.x>
- Romanelli, A. M., Fu, J., Herrera, M. L., & Wickes, B. L. (2014). A universal DNA extraction and PCR amplification method for fungal rDNA sequence-based identification. *Mycoses*, 57(10), 612–622. <https://doi.org/10.1111/myc.12208>
- Romano, P., Fiore, C., Paraggio, M., Caruso, M., & Capece, A. (2003). Function of yeast species and strains in wine flavour. *International Journal of Food Microbiology*, 86, 169–180. [https://doi.org/10.1016/S0168-1605\(03\)00290-3](https://doi.org/10.1016/S0168-1605(03)00290-3)

- Romano, P., Suzzi, G., Comi, G., Zironi, R., & Maifreni, M. (1997). Glycerol and other fermentation products of apiculate wine yeasts. *Journal of Applied Microbiology*, 82(5), 615–618. <https://doi.org/10.1111/j.1365-2672.1997.tb02870.x>
- Ru, M., Bernal-grande, M. C., Cordero-bueso, G., & Hughes-herrera, D. (2017). A microtiter plate assay as a reliable method to assure the identification and classification of the veil-forming yeasts during sherry wines ageing. <https://doi.org/10.3390/fermentation3040058>
- Selli, S., Cabaroglu, T., Canbas, A., Erten, H., Nurgel, C., Lepoutre, J., & Gunata, Z. (2004). Volatile composition of red wine from cv. Kalecik Karası grown in central Anatolia. *Food Chemistry*, 85(2), 207-213. doi:10.1016/j.foodchem.2003.06.008
- Sidhu, D., Lund, J., Kotseridis, Y., Saucier, C., Sidhu, D., Lund, J., & Kotseridis, Y. (2015). Methoxypyrazine analysis and influence of viticultural and enological procedures on their levels in grapes , musts , and wines *Crit. Rev. Food Sci. Nutr.*, 485–502. <https://doi.org/10.1080/10408398.2012.658587>
- Sincar, Ö. (2010). Kalecik karası üzümlerinden kırmızı şarap üretiminde soğuk maserasyon uygulamasının aroma ve antosianin bileşikleri üzerine etkileri. (*Master Thesis*) Cukurova University, Adana, Turkey
- Smith, M.Th. 1998b. Brettanomyces Kufferath and van Laer. In: The Yeasts. C.P. Kurtzman and J.W. Fell (Eds.), 4th edition, Chapter 63, pp. 450–453. Elsevier, New York, NY
- Snow, P.G. and Gallander, J.F. (1979) Deacidification of white table wines through partial fermentation with *Schizosaccharomyces pombe*. *Am. J. Enol. Vitic.* 30, 45-48.
- Swiegers, J. H., & Pretorius, I. S. (2005). Yeast modulation of wine flavor. *Advances in Applied Microbiology*, 57(SUPPL. A), 131–175. [https://doi.org/10.1016/S0065-2164\(05\)57005-9](https://doi.org/10.1016/S0065-2164(05)57005-9)
- Tao, Y., & Zhang, L. (2010). Intensity prediction of typical aroma characters of cabernet sauvignon wine in Changli County (China). *LWT - Food Science and Technology*, 43(10), 1550-1556. doi:10.1016/j.lwt.2010.06.003
- Thomas, D.S. & Davenport R.R. (1985). *Zygosaccharomyces bailii* a profile of characteristics and spoilage activities. *Food Microbiol.* 2: 157–169.

- Tofalo, R., Chaves-López, C., Di Fabio, F., Schirone, M., Felis, G. E., Torriani, S. (2009). Molecular identification and osmotolerant profile of wine yeasts that ferment a high sugar grape must. *International Journal of Food Microbiology*, 130(3), 179-187.
- Toro, M. E., & Vazquez, F. (2002). Fermentation behaviour of controlled mixed and sequential cultures of *Candida cantarellii* and *Saccharomyces cerevisiae* wine yeasts. *World Journal of Microbiology & Biotechnology*, 18, 347–354. <https://doi.org/10.1023/A:1015242818473>
- Tristezza, M., Tufariello, M., Capozzi, V., Spano, G., Mita, G., & Grieco, F. (2016). The oenological potential of *Hanseniaspora uvarum* in simultaneous and sequential co-fermentation with *Saccharomyces cerevisiae* for industrial wine production. *Frontiers in Microbiology*, 7(MAY), 1–14. <https://doi.org/10.3389/fmicb.2016.00670>
- Türk Gıda Kodeksi Şarap Tebliği* (NO: 2008/67). Retrieved from http://www.gidamo.org.tr/mevzuat/mevzuat_detay.php?kod=76
- Urso, R., Rantsiou, K., Dolci, P., Rolle, L., Comi, G., & Cocolin, L. (2008). Yeast biodiversity and dynamics during sweet wine production as determined by molecular methods. <https://doi.org/10.1111/j.1567-1364.2008.00364.x>
- Ükelgi, N. (2011). Isolation, identification and characterization of wine yeast species from grapes of three different vineyards in Turkey. (*Master Thesis*) Sabancı University, Istanbul, Turkey.
- Valero, E., Moyano, L., Millan, M. C., Medina, M., & Ortega, J. M. (2002). Higher alcohols and esters production by *Saccharomyces cerevisiae*. Influence of the initial oxygenation of the grape must. *Food Chemistry*, 78(1), 57–61. [https://doi.org/10.1016/S0308-8146\(01\)00361-2](https://doi.org/10.1016/S0308-8146(01)00361-2)
- Van der Aa Kühle, A., & Jespersen, L. (1998). Detection and identification of wild yeasts in lager breweries. *International Journal of Food Microbiology*, 43(3), 205-213.
- Van der Walt, J.P. and Van Kerken, A. E. (1961). The wine yeasts of the Cape. Part V. Studies on the occurrence of *Brettanomyces intermedius* and *Brettanomyces schanderlii*. *Antonie van Leeuwenhoek* 27: 81–90.
- Van Dijken, J.P., and Harder, W.(1974) Optimal conditions for the enrichment and isolation of methanol-assimilating yeasts. *J.Gen. Microbiol.* 84, 409-411

- Vaughan-Martini, A. and Martini, A. (1998) Determination of ethanol production, In: The Yeasts, A Taxonomic Study. Kurtzman, C.P. and Fell, J.W. (eds), Elsevier, pp. 107, Amsterdam.
- Viana, F., Gil, J. V., Genovés, S., Vallés, S., & Manzanares, P. (2008). Rational selection of non-*Saccharomyces* wine yeasts for mixed starters based on ester formation and enological traits. *Food Microbiology*, 25(6), 778–785. <https://doi.org/10.1016/j.fm.2008.04.015>
- Villa-Carvajal, M., Querol, A., & Belloch, C. (2006). Identification of species in the genus *Pichia* by restriction of the internal transcribed spacers (ITS1 and ITS2) and the 5.8S ribosomal DNA gene. *Antonie van Leeuwenhoek, International Journal of General and Molecular Microbiology*, 90(2), 171–181. <https://doi.org/10.1007/s10482-006-9071-0>
- Vries, C. D., Buica, A., Brand, J., & Mckay, M. (2016). The impact of smoke from vegetation fires on sensory characteristics of cabernet sauvignon wines made from affected grapes. *South African Journal of Enology and Viticulture*, 37(1). doi:10.21548/37-1-755
- Wang, J., Capone, D. L., Wilkinson, K. L., & Jeffery, D. W. (2016). Chemical and sensory profiles of rosé wines from Australia. *Food Chemistry*, 196, 682–693. <https://doi.org/10.1016/j.foodchem.2015.09.111>
- Wang, X. C., Li, A. H., Dizy, M., Ullah, N., Sun, W. X., & Tao, Y. S. (2017). Evaluation of aroma enhancement for “Ecolly” dry white wines by mixed inoculation of selected *Rhodotorula mucilaginosa* and *Saccharomyces cerevisiae*. *Food Chemistry*, 228, 550–559. <https://doi.org/10.1016/j.foodchem.2017.01.113>
- Welke, J. E., Zanus, M., Lazzarotto, M., & Zini, C. A. (2014). Quantitative analysis of headspace volatile compounds using comprehensive two-dimensional gas chromatography and their contribution to the aroma of Chardonnay wine. *Food Research International*, 59, 85–99. doi:10.1016/j.foodres.2014.02.002
- Wines of Turkey. (2019). Retrieved from <http://www.winesofturkey.org/>
- Ye, M., Yue, T., & Yuan, Y. (2014). Effects of sequential mixed cultures of *Wickerhamomyces anomalus* and *Saccharomyces cerevisiae* on apple cider fermentation. *FEMS Yeast Research*, 14(6), 873–882. <https://doi.org/10.1111/1567-1364.12175>

Zalacain, A., Marin, J., Alonso, G. L., & Salinas, M. R. (2007). Analysis of wine primary aroma compounds by stir bar sorptive extraction. *Talanta*, 71, 1610–1615. <https://doi.org/10.1016/j.talanta.2006.07.051>

Zanol, G. C., Baleiras-Couto, M. M., & Duarte, F. L. (2010). Restriction profiles of 26S rDNA as a molecular approach for wine yeasts identification. *Ciencia e Tecnica Vitivinicola*, 25(2), 75–85.

Ženišová, K., Chovanová, K., Chebeňová-Turcovská, V., Godálová, Z., Kraková, L., Kuchta, T., Brežná, B. (2014). Mapping of wine yeast and fungal diversity in the Small Carpathian wine-producing region (Slovakia): evaluation of phenotypic, genotypic and culture-independent approaches. *Annals of Microbiology*, 64(4), 1819–1828. <https://doi.org/10.1007/s13213-014-0827-x>

Zohre, D. E., & Erten, H. (2002). The influence of *Kloeckera apiculata* and *Candida pulcherrima* yeasts on wine fermentation. *Process Biochemistry*, 38(3), 319–324. [https://doi.org/10.1016/S0032-9592\(02\)00086-9](https://doi.org/10.1016/S0032-9592(02)00086-9)

Zott, K., Miot-Sertier, C., Claisse, O., Lonvaud-Funel, A., & Masneuf-Pomarede, I. (2008). Dynamics and diversity of non-*Saccharomyces* yeasts during the early stages in winemaking. *International Journal of Food Microbiology*, 125(2), 197–203. <https://doi.org/10.1016/j.ijfoodmicro.2008.04.001>

APPENDICES

A. DNA Sequencing of Isolated Yeasts

Table A.1. DNA Sequencing of Isolated Yeasts

Name	BLAST Result	Similarity	Region	Sequence
KA 0.CM NS1 (K1)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	GGAAAGGATCATTAGATTGAATTATCATTTGCTCGAGTTCTAGTTAG ATCTTTTACATAATGTGTATCTTATTGAAGATGTGCGCTTAATTGCGC TGCTTTTTAAAGTGTGCGAGTAGAAGTAATCTTGTGAATCTCAGTC ACGTTTACACACATTGGAGTTTTACTTTAATTAAATTCTTCTGCTT GAATCGAAAGGTTCAAGGCAAAAAACAAACAAACAATTTATTAT TATAATTTTAAACTAAACCAAATTCTAACGGAAATTTTAAAATAA TTAAAACCTTCAACACGGATCTCTGGTTCTCGCATCGATGAAGAAC GTAGCGAATTGCGATAAGTAATGTGAATTGCAAGATACTCGTGAATCATT GAATTTTGAACGCACATTGCCCTTGAGCATTCTCAAGGGCATGCC GTTTGAGGCTCATTTCTTCAAGGAAATTGAGATAATTTTATTGGTTGTG GGCAGACTCAGGGTTAGCTGAAGATTGTTCAATCTTTTA ATTCAACACTTAGCTCTTGGAGACGCTGTTCTCGCTGTGATGTATT TAAGAATTATTGCTTTACTTTACAAGGGAAATGGTAATGTACCTAGGC AAGGGTTGCTTAATTACATCAAGTTGACCTCAA
KA 0.CM NS2 (K2)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATTAATTAAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCAACACG GATCTCTGGTCTCGCATCGATGAAGAACGTAGCGAATTGCGATAAGT AATGTGAATTGCGATACTCGTGAATCATTAATTGAACGGCACATT CGCCCTTGAGCATTCTCAAGGGCATGCCCTGTTGAGCGTATTCCTTC TCAAAGATAATTATTATTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTTAATTCAACACTTAGCTCTT TGGAGACGCTGTCTCGCTGTGATGTATTATGGATTATTGTTACT TTACAAGGGAAATGGTAATGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTTGACCTA
KA 0.CM NS3 (K3)	<i>Hanseniaspora uvarum</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATTAATTAAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCAACACG GATCTCTGGTCTCGCATCGATGAAGAACGTAGCGAATTGCGATAAGT AATGTGAATTGCGATACTCGTGAATCATTAATTGAACGGCACATT CGCCCTTGAGCATTCTCAAGGGCATGCCCTGTTGAGCGTATTCCTTC TCAAAGATAATTATTATTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTTAATTCAACACTTAGCTCTT TGGAGACGCTGTCTCGCTGTGATGTATTATGGATTATTGTTACT TTACAAGGGAAATGGTAACGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTTGACCTA
KA 0.CM NS5 (K5)	<i>Hanseniaspora guilliermondii</i>	100%	D1/D2 Domain	CGCGAGGTGAAGCGGTAAGCTCAAATTGAAATCTGGTACTTCACTG GCCGAGTTGATTTGAGAATTGCTTTGATTAAGGCTTGTCTATG TTCTTGGAACAGGACGTAGAGGGTGAGAATCCGTTGGCAGG ATACCTTTCTCTGTAAGACTTCTGAGAGTCGAGTTGGGAGAATG CAGCTCAAAGTGGTGTAAATTCCATCTAAAGCTAAATATTGGCGAGA GACCGATAGCGAACAGTACGTGATGGAAAGATGAAAAGACTTGA AAAGAGAGTAAAAAGTACGTGAAATTGTAAGGAAAGGGCATTG ATCAGACATGGTTTGCATGCACTCGCTCTCGTGGCTGGCCT CTCAAAAATTTCACTGGGCCAACATCGTTCTGGCAGCAGGATAATCA TTAAGAATGTAGTACCTCGTAGTGTATAGCTTATTGGAATACTGCT AGCTGGGATTGAGGACTGCGCTCGGAAGGATGTTGCAATGGTTA ATGCCGCC
		100%	ITS	CGGAAGGATCATTAGATTGAATTATCATTTGCTCGAGTTCTAGTTA GATCTTTACATAATGTGTATCTTATTGAAGATGTGCGCTTAATTGCG CTGCTTTAAAGTGTGCGAGTAGAAGTAATCTTGTGAATCTCAGTC AACGTTTACACACATTGGAGTTTTACTTTAATTAAATTCTTCTGCTT TGAATCGAAAGGTTCAAGGGAAAAAACAAACAAACAATTTATTAA ATTATAATTAAACTAAACCAAATTCTAACGGAAATTAAAT AATTAAAACCTCAACACGGATCTCTGGTCTCGCATCGATGAAGA ACGTAGCGAATTGCGATAAGTAATGTGAATTGCAAGATACTCGTGAATCA TTGAAATTGGAGACGCAATTGCCCTTGAGCATTCTCAAGGGCATGC CTGTTGAGGCTATTCTCTCAAAGATAATTTTATTGGTTG TGGCGATACTCAGGGTTAGCTGAATTGAAGATTGTTCAATTGTT TAATTCAACACTTAGCTCTTGGAGACGCTGTTCTCGCTGTGATGTATT TATGAATTATTGCTTTACTTTACAAGGGAAATGGTAATGTACCTAGG CAAAGGGTTGCTTAATTACATCAAGTTGACCTCA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 0.CM NS2 (K7)	<i>Hanseniaspora opuntiae</i>	100%	ITS	GTCAACTTGATGAATATTAAGCAACCCCTTGCTTAAGGTACATTACC ATTTCCTGTAAGTAAACGAATAAATCCATAAAATACATCACAGCGA GAACAGCGTCTCCAAGAAGCTAACCTGAGTATGCCACAACCAAAAAAT CAGTCTCCAATTCAAGCTAACCTGAGTATGCCACAACCAAAAAAT AAAATTATCTTTGAGAAGGAAATGACGCTAAACAGGCATGCCCTT GAGAATGCTCAAGGGCGAATGTGCGTTAAAAATTCAATGATTCAACG AGTATCTGAATTTCACATTACTTATCGAATTGCTACGTTCTCATCGA TGCAGAACCAGAGATCGTTGTTGAAAGTTAAATTATTTAAAT TTCCGTTAGGAATTGGTTAGTTAAATTATAATAAAATAAAAT TGTGTTGTTGTTGGCTGATTCAAAGCAGAAAGAA TTAAATTAAAGTAAAA
KB 0.CM NS3 (K8)	<i>Hanseniaspora opuntiae</i>	100%	ITS	GTCAACTTGATGAATATTAAGCAACCCCTTGCTTAAGGTACATTACC ATTTCCTGTAAGTAAACGAATAAATCCATAAAATACATCACAGCGA GAACAGCGTCTCCAAGAAGCTAACCTGAGTATGCCACAACCAAAAAAT CAGTCTCCAATTCAAGCTAACCTGAGTATGCCACAACCAAAAAAT AAAATTATCTTTGAGAAGGAAATGACGCTAAACAGGCATGCCCTT GAGAATGCTCAAGGGCGAATGTGCGTTAAAAATTCAATGATTCAACG AGTATCTGAATTTCACATTACTTATCGAATTGCTACGTTCTCATCGA TGCAGAACCAGAGATCGTTGTTGAAAGTTAAATTATTTAAAT TTCCGTTAGGAATTGGTTAGTTAAATTATAATAAAATAAAAT TGTGTTGTTGTTGGCTGATTCAAAGCAGAAAGAA TTAAATTAAAGTAAAA
KB 0.CM NS4 (K9)	<i>Rhodotorula mucilaginosa</i>	100%	ITS	TCCGTAGGGTAACCTCGCGAAGGATCATTAGTGAATAGGACGTCC AACTTAACCTGGAGTCCGAACCTCACTTCTAACCTGTGATTGTTT GGGATAGTAACCTCGCAAGAGGGCGAACCTCTATTCACTTATAAACAC AAAGTCTATGATGTTAAATTATAACAAATAAAACTTCAACAA CGGATCTTGGCTCTCGCATCGATGAAGAACGCAAGCGAAATCGGATAA GTAATGTGAATTGAGAATTCTGATGAATCATCGAATCTTGAACCGACC TTGCCTCATGGTATTCCGGAGCATGCTGTTGAGTGTATGAA ACTTCAACCCCTCTTCTTAATGATGAGGAGGTGTTGGTTCTGAG CGCTGCTGGCTTAGGGCTAGCTGGCTAATGCATTAGCATTCCGC AATGCAACTTCGGATTGACTTGGCTAATAGACTATCGCTGAGGAATT CTAGTCTTCGGACTAGAGCCGGTTGGTTAAAGGAAGCTCTAACAG AATGTCACATTAAAGATTAGATCTCAA
KB 0.CM NS5 (K10)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTAATTAAATTCTTCTGTTGAATCGAAAGGTCAAGGC AAAAAACAAACACAAACAAACATTATTATTATAATTAAACTTAAACTAAA CCAAAATTCTAACGGAAATTAAATAATTAAACTTCAACAAACG GATCTCTTGGTTCTCGCATCGATGAAGAACGTAAGCGATTGCGATAAGT AATGTGAATTGAGCAGATACTCGTGAATCATTGAATTGGAAACGCACATT CGCCCTTGAGCATTCTCAAGGGCATGCTGTTGAGCGTCAATTCTTC TCAAAAGATAATTATTATTATTGGTTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCTGAGTCTTAAATTCAACACTAGCTT TGGAGACGCTGTTCTCGCTGTGATGTATTGATTGATTCTGTTACT TTACAAGGGAAATGTAATGTACCTTAGGCAAAGGGTTGTTAAAT TCATCAAGTTGACCTCAA
KA 4.CM NS1 (K11)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTAATTAAATTCTTCTGTTGAATCGAAAGGTCAAGGC AAAAAACAAACACAAACAAACATTATTATTATAATTAAACTTAAACTAAA CCAAAATTCTAACGGAAATTAAATAATTAAACTTCAACAAACG GATCTCTTGGTTCTCGCATCGATGAAGAACGTAAGCGATTGCGATAAGT AATGTGAATTGAGCAGATACTCGTGAATCATTGAATTGGAAACGCACATT CGCCCTTGAGCATTCTCAAGGGCATGCTGTTGAGCGTCAATTCTTC TCAAAAGATAATTATTATTATTGGTTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCTGAGTCTTAAATTCAACACTAGCTT TGGAGACGCTGTTCTCGCTGTGATGTATTGATTGATTCTGTTACT TTACAAGGGAAATGTAATGTACCTTAGGCAAAGGGTTGTTAAAT TCATCAAG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KA 4.CM NS2 (K12)	<i>Hanseniaspora opuntiae</i>	100%	D1/D2 Domain	CTTAGTACGGCGAGTGAAGCGGTAAAAGCTCAAATTGAAATCTGGTAC TTCACTGCCCCGAGTTAATTGAGAATTGCTTTGATTAGTCCT GTCTATGTTCTTGGAACAGGACGTCATAGAGGGTGAGAATCCCGTTG GCGAGGATACTTTCTGTAAAGACTTTCAAGAGTCGAGTTGTTG GGAATGCAGCTAAAGTGGTGGTAATTCCATCTAAAGCTAAATATTG CGAGAGACCGATAGCGAACAGTACAGTGTGGAAAGATGAAAAGA ACTTGAAAAGAGAGTGAAAAAGTACCGTGAATTGTTGAAAGGGAAAG GCATTGATCAGACATGGTTTTGCATGCACTCGCTCTCGTGGCT TGGGCCTCTAAAATTCACTGGGCCAACATCAATTCTGGCAGTAGGA TAAATCATTAAGAATGTAGCTACCTCGTAGTGTATAGCTTATTGAA TACTGCTAGCTGGGATTGGAGACTGCGCTCGGCAAGGATGTTGCA ATGGTTAAATGCCCGCTGTGAAACACGGACC
	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTAATTAACTCTGCTTGAATCGAAAGGTTCAAGGC AAAAAAACAACACAAACAAATTATTATTAAACTTAAACTAAA CCAAAATTCTAACGGAAATTAAATAATTAAACTTAAACAAAC GATCTTGGTCTCGCATCGATGAAGAACGCTAGCGAATTGCGATAAGT AATGTGAATTGAGATACTCGTGAATCATTAATTGAAATTGCAAC CGGCCCTTGAGCATTCTAACGGCATGCTTGTGGCGTATTCTTC TCAAAGATAATTTTATTTTGGTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCTAGCTTTAACTCAACACTAGCTT TGGAGACGCTGTCTCGCTGTGATGTATTATGGATTATTCGTTTACT TCACAAGGAAATGTAATGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTTGACCTCAAATC
KA 4.CM NS3 (K13)	<i>Hanseniaspora uvarum</i>	100%	ITS	ACCTGGGAAGGATCATTAGATTGAATTATCATTTGCTGAGTTCTA GTTTAGATCTTACAATAATGTATCTTATTGAAGATGTCGCTTA ATTGCGCTGTTTAAAGTGTGCGAGTAGAAGTAATCTGCTGTAATC TCAGTCACGTTACACATGGAGTTTTACTTTAATTAAATCTTT CTGCTTGAATCGAAAGGTTCAAGGCAAAAACAAACAAACAAATT TATTATTATAATTAAACTAAACAAAATTCTAACGGAAATT AAAATAATTAAACTTCAACACGGATCTTGGTCTCGCATCGAT GAAGAACGTAGCGAATTGCGATAAGTAATGTGAATTGAGATACTCGT GAATCATTAATTGGAAACGCACATTGCGCCCTTGAGCATTCTCAAGG GCATGCCCTGGTGGCGATACTCAGGGTTAGCTGAATTGAAGATGTTCA ATCTTTTAATTCAACACTAGCTTTGGAGACGCTTCTCGCT GATGTATTGAAATTATCGTTTACTTACAAGGAAATGTAATGT ACCTTAGGCAAAGGGTTGCTTTAATATTCAAGTTGACCTCAA
KA 4.CM NS5 (K15)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTAATTAACTCTGCTTGAATCGAAAGGTTCAAGG AAAAAAACAACACAAACAAATTATTAAATTAAATTAAACTAA ACAAAATTCTAACGGAAATTAAATAATTAAACTTAAACAAAC GGATCTTGGTCTCGCATCGATGAAGAACGCTAGCGAATTGCGATAAG TAATGTGAATTGCGAGATACTCGTGAATCATTAATTGAAACGACAT TGGCCCTTGAGCATTCTAACGGCATGCTTGTGGCGTATTCTT CTCAAAGATAATTTTATTGGTGTGGCGATACTCAGGGTTAG CTGAAATTGGAGACTGTTCTAGCTTTTAATTCAACACTAGCTT TGGAGACGCTGTCTCGCTGTGATGTATTATGGATTATCGTTTAC TTCAAGGAAATGTAATGTACCTAGGCAAAGGGTTGCTTTAATA TTCATCAAGTTGACCTCAAATC
KB 4.CM NS1 (K16)	<i>Hanseniaspora opuntiae</i>	100%	ITS	GGAAGGATCATTAGATTGAATTATCATTTGCTGAGTTCTGTTAGA TCTTACAATAATGTATCTTATTGGAGATGTCGCTTAATTGCGCT GCTTCATTAGAGTGTGCGAGTAGAAGTAGTCTGCTTGAATCTCAGTC ACGTTACACACATTGGAGTTTTACTTTAATTAAATTCTGCTT ATCGAAAGGTTCAAGGCAAAAACAAACAAACAAATTAAAT ATAATTAAACTAAACAAATTCTAACGGAAATTAAATAAT TAAAATTCTAACACGGATCTTGGTCTCGCATCGATGAAGAACG TAGCGAATTGCGATAAGTAATGTGAATTGAGATACTCGTGAATCAT AATTGGAGCAGCATTGCGCCCTTGAGCATTCTCAAGGGCATGCG TTGAGCGTCAATTCTCTCAAAGATAATTAAATTGGTGTGG GCGATACTCAGGGTTAGCTGAAATTGGAGACTGTTCTGAGTATT GGATTATTGTTTACTTACAAGGAAATGTAATGTACCTAGGCA AAGGGTTGCTTTAATATTCAAGTT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 4.CM NS2 (K17)	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACATTCCCTTGAAAGTAAAACGAATAAATCATAAATACATCACAGCGAGAACAGCGTCTCAAAGAAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTTCAGCTAACCCGTAGATATGCCAACACCAAAAAATAA AAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGATTACGAGT ATCTGCAATTTCACATTACTTATCGCAATTGCTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTGTTGAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAAATTATAATAAAAATAAAATTGT TTGTGTTGTTTTGCCTGAAACCTTCGATTCAAAGCAGAAAGAATTA AATTAAAGTAAAAAA
KB 4.CM NS4 (K19)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCACAAACG GATCTTGTCTCGCATCGATGAAGAACGTTAGCGAATTGCGATAAGT AATGTGAATTGAGATACTCGTGAATCATTGAATTGGAAACGCACATT GCGCCCTTGAGCATTCTCAAGGGCATGCCGTGTTGAGCGTCATTCCCTC TCAAAGATAATTTTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTAAATTCAACACTTAGCTTCTT TGGAGACGCTGTCTCGCTGTGATGTATTGATTTCGTTTACT TTACAAGGGAAATGTAATGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTTGACCTCAAAT
KB 4.CM NS5 (K20)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCACAAACG GATCTTGTCTCGCATCGATGAAGAACGTTAGCGAATTGCGATAAGT AATGTGAATTGAGATACTCGTGAATCATTGAATTGGAAACGCACATT GCGCCCTTGAGCATTCTCAAGGGCATGCCGTGTTGAGCGTCATTCCCTC TCAAAGATAATTTTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTAAATTCAACACTTAGCTTCTT TGGAGACGCTGTCTCGCTGTGATGTATTGATTTCGTTTACT TTACAAGGGAAATGTAATGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTTGACCTCAAAT
KA 2.NM NS1 (K21)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCACAAACG GATCTTGTCTCGCATCGATGAAGAACGTTAGCGAATTGCGATAAGT AATGTGAATTGAGATACTCGTGAATCATTGAATTGGAAACGCACATT GCGCCCTTGAGCATTCTCAAGGGCATGCCGTGTTGAGCGTCATTCCCTC TCAAAGATAATTTTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTAAATTCAACACTTAGCTTCTT TGGAGACGCTGTCTCGCTGTGATGTATTGATTTCGTTTACT TTACAAGGGAAATGTAATGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGT
KA 2.NM NS9 (K25)	<i>Hanseniaspora uvarum</i>	100%	D1/D2 Domain	AGCGGTAAAGCTCAAATTGAAATCTGGTACTTCAGTGCCGAGTT TAATTGTAGAATTGCTTGTGATTAGTCCTGCTATGTCCTGGAA CAGGACGTCATAGAGGGTGAAGATCCGTTGGCGAGGATACTTCT CTGTAAGACTTTGAGAGCTCAAGCTAAATGGCCAGAGACCGATAGC TGTTGGTAATTCCATCTAAAGCTAAATGGCCAGAGACCGATAGC GAACAAGTACGTGATGAAAGATGAAAAGAACTTGAGAGAGT AAAAGTACGTGAAATTGTAAGGGAGGGCATTTGATCACAGCATG GTGTTTTGACACTCGCTCTGTTGGCTGTTGGCTCTCAAATT TCACTGGGCCAACATCAATTCTGGCAGCAGGATAAATCATTAAGATGT AGCTACT
	<i>Hanseniaspora uvarum</i>	100%	ITS	TTTTTACTTTAATTAAATTCTTCTGCTTGAATCGAAAGGTTCAAGGC AAAAACAAACACAAACAATTTATTATAATTAAACTAAA CAAAATTCTAACGGAAATTAAAATAATTAAAACCTTCACAAACG GATCTTGTCTCGCATCGATGAAGAACGTTAGCGAATTGCGATAAGT AATGTGAATTGAGATACTCGTGAATCATTGAATTGGAAACGCACATT GCGCCCTTGAGCATTCTCAAGGGCATGCCGTGTTGAGCGTCATTCCCTC TCAAAGATAATTATTGGTTGTGGCGATACTCAGGGTTAGC TTGAAATTGGAGACTGTTCACTTTAAATTCAACACTTAGCTTCTT TGGAGACGCTGTCTCGCTGTGATGTATTGATTTCGTTTACT TTACAAGGGAAATGTAACGTACCTAGGCAAAGGGTTGCTTTAATAT TCATCAAGTT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 2.NM NS2 (K26)	<i>Hanseniaspora uvarum</i>	99%	ITS	TGCGGAAGGATCATTAGATTGAATTATCATTGGCTCGAGTTCTGTT AGATCTTTACAATAATGTATCTTATTGAAGATGTGCGCTTAATGCG GCTGCTTCTTAGAGTGTGCGAGTGAAGTAGTCTTGCTTGAATCTCAGT CAACGCTCACACATTGGAGTTTTACTTTAATTAAATTCTTCGCTT TGAATCGAAAGGTTCAAGGCAAAAACAAACACAAACAATTTTATT ATTATAATTTTAAACTAAACCAAATTCTAACCGAAATTAAAAT AATTAAAACCTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGA ACGTAGCGAATTGCGATAAGTAATGTGAATTGCAGATACTCGTGAATCA TTGAATTGGACGCACATTGCCCTGAGCATTCTCAGGGCATGC CTGTTGAGCGTCATTCCCTCAAAGATAATTATTATTGGTT GTGGGCAGACTCAGGGTAGCTTGAATTGGAGACTGTTCTGAGTCTT TTAAATTCAACACTAGCTTGGAGACGCTGTTCTGCTGTGATGTA TTTATGGATTATCGTTTACTTACAAGGAAATGGTAACGTACNCT TAGGCAAAGGGTTGCTTTAATTATTCATCAAGT
KB 2.NM NS4 (K27)	<i>Hanseniaspora opuntiae</i>	100%	ITS	GCGGAAGGATCATTAGATTGAATTATCATTGGCTCGAGTTCTGTTA GATCTTTACAATAATGTATCTTATTGGAGATGTGCGCTTAATGCG CTGCTTCTTAGAGTGTGCGAGTGAAGTAGTCTTGCTTGAATCTCAGT CAACGTTACACATTGGAGTTTTACTTTAATTAAATTCTTCGCTT TGAATCGAAAGGTTCAAGGCAAAAACAAACACAAACAATTTTATT ATTATAATTTTAAACTAAACCAAATTCTAACCGAAATTAAAAT AATTAAAACCTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGA ACGTAGCGAATTGCGATAAGTAATGTGAATTGCAGATACTCGTGAATCA TTGAATTGGACGCACATTGCCCTGAGCATTCTCAGGGCATGC CTGTTGAGCGTCATTCCCTCAAAGATAATTATTATTGGTT TGGGGCAGACTCAGGGTAGCTTGAATTGGAGACTGTTCTGAGTCTT TAATTCAACACTAGCTTGGAGACGCTGTTCTGCTGTGATGTTA TATGGATTATCGTTTACTTACAAGGAAATGGTAATGTACCTTAGG CAAAGGGTTGCTTTAATTATTCATCAAGT
KB 2.NM NS8 (K29)	<i>Saccharomyces cerevisiae</i>	98%	ITS	GGATCATTAAAGAAATTATAATTGGAAAATGGATTTTGGTTGG CAAGAGCATGAGAGTTTACTGGGAGAGAAGACAGAGATGGAGAGT CCAGCCGGGCTGCGCTTAAGTGCCTGCTTAGGTTGTAAGTTT CTTCTGCTATTCAAACGGTGAGAGATTCTGTGTTTGTATAGGA CAATTAAAACGTTCAATACAACACACTGTGGAGTTTCAATCTTGCA AACTTTCTTGGCATTGAGCAATCGAGAACAGGGGCCAGAGGTAACNAACA CAAACAATTATTATTCAATTAAATTGGTCAAACAAAGAATTGG TAACTGGAAATTAAAATTTAAACTTCAACAAACGGATTGGG TTCTCGCATCGATGAAGAACGAGCAGAAATGCAGATACTGTGAATT GCAGAATTCCGTGAATCTGAATCTTGAACGCCCTTGCGCCCTTG GTATTCCGGGGCATGCGCTGGTGGAGCGTCATTCCCTCAAACCTT TGTTGGTAGTGAGTGAATTGGAGCTTGAATTGCTGGCCCTTTCATTGGATGTTTCAAAAGAGGGTTCTTGCCTGCTGAGG TATAATGCAAGTACGGCTGTTTAGGTTTACCAACTGCGCTAATCTT TTTTACTGAGCGTATTGGAACGTTATCGATAAGAAGAGAGCGCTAG CGAACAAATGTTCAAAG
KA 4.NM NS2 (K31)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	GGTCAACTTGTGAATTATAAAAGCAACCCTTGCCCTAACGGTACATTAC CATTCCCTTGTAAAGTAAACGAATAATTCTAAATACATCACAGC AGAACAGCGTCTCAAAGAAGCTAAGTGTGAATTAAAAAGATGAA ACAATTCTCAATTCAAGCTAACCCCTGAGTATGCCAACACAAAAAA TAAAAAAATTATTTGAGAAGGAAATGACGCTAACAGGCATGCCCT TGAGAATGCTCAAGGGCGCAATGCGCTTCAAAATTCATGATTCAAG AGTATCTGCAATTCAATTCTGCAATTGCTAACGTTCTCATCGA TGCGAGAACCAAGAGATCGCTTGTGAAAGTTAAATTATTAAAAT TTCCGTTAGGAATTGGTTAGTTAAAAAATTATAAAAATTTAAAAT TGTTGGTAGTTGGCTTGCCTTGAACCTTCGATTCAAAGCAGAAAGAA TAAATTAAAGTAAAAAAACTCCAATGTGTAACGTTGACTGAGATT CAAGCAAGATTACTTCACTGCGACACTTAAAAAGCAGCGAACATA CGCACATCTCAATAAGATAACACATTATGTAAAAGATCTAAACTA GAACTCGAGCAACAATGATAATTCAATCTAATGATCCTCCGAGGTC
KA 4.NM NS4 (K32)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAAATTCTCTGCTTGAATCGAACGGTCAAGGC AAAAACAAACACAAACAATTTTATTATAATTTTAAACTAAA CAAAATTCTAACGGAATTAAAATAATTAAACTTCAACACG GATCTCTGGTCTCGCATCGATGAAGAACGTTAGCGAATTGCGATAAGT AATGTGAATTGCGATACTCGTGAATCATGAAATTGCGATAAGT GCGCCCTTGAGCATTCTCAAGGGCATGCCCTGGTGGAGCGTCATTCCCTC TCAAAAGATAATTTTATTGGTTGTGGCGATACTCAGGGTAGC TTGAAATTGGAGACTGTTCTGAGTCTTTAATTCAACACTAGCTTCTT TGGAGACGCTGTTCTCGCTGTGATGTTATGGATTATTGCTTACT TTACAAGGAAATGGTAATGTACCTTACGGCAAAGGGTTGCTTTAATAT TCATCAAG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KA 4.NM NS6 (K33)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	CGCGAAGGATCATTAGATTGAATTATCATTGTCGAGTTCTAGTTT AGATCTTTACAATAATGTGTATCTTATTGAAGATGTGCGCTTAATTGC GCTGCTTTAAAGTGTGCGAGTAGAAGTAATCTTGCTTGAATCTCAGT CAACGTTACACACATTGGAGTTTTACTTTAATTAAATTCTCTGCT TTGAATCGAAAGGTTCAAGGCAAAAACAACACAAACAATTATT ATTATAATTTTAAACTAACCAAATTCTAACGGAAACGGAAATT AATTAAAACCTTCAACAAACGGATCTTGGTCTCGCATCGATGAAGA ACGTAGCGAATTGCATAAGTAATGTAATTGCAGATACTCGTGAATCA TTGAATTTTGAACGCACATTGCCCTGAGCATTCTCAAGGGCATGC CTGTTGAGGCTCATTTCTTCAAAGATAATTTTATTGGTTG TGGGCGATACTCAGGGTTAGCTTGAAGATTGTTCAATTCTT TAATTCAACACTTAGCTTGGAGACGCTGTTCTCGTGTGATGTATT TATGAATTIATCGTTTACTTACAAGGAAATGTAATGTACCTAGG CAAAGGGTGCTTTAATATTCAAGTTGACCTCAA
KA 4.NM NS8 (K34)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTAATTAAATTCTCTGCTTGAATCGAAAGGTTCAAGGC AAAAAACAAACACAAACAAATTATTATTATAATTAAATT CCAAAATTCTAACGGAAATTAAATTAAATTAAATTCTAACAAACG GATCTTGTCTCGCATCGATGAAGAACGTAACGCAATTGCGATAAGT AATGTGAATTGCAGATACTCGTGAATCATTGAATTGCGACATT GCCGCTTGAAGCATTCTCAAGGGCATGCCTGTTGAGCGTCATT TCAAAGATAATTTTATTGGTGTGGCGATACTCAGGGTTAG TTGAAATTGGAGACTGTTCACTTTTAAATTCAACACTTAGCTT TGGAGACGCTGTTCTCGCTGTGATGTATTATGGATTATTGTT TTACAAGGAAATGTAATGTACCTAGGAAAGGTTGCTTTAAT TCATCAAGTTGACCTCAA
KA 4.NM NS10 (K35)	<i>Hanseniaspora opuntiae</i>	100%	D1/D2 Domain	ACGGCGAGTGAAGCGTAAAGCTCAAATTGAATCTGGTACTTCAG TCCCCGAGTTGTAATTGTAGATTGCTTGTATTAGTCCTTGTCTAT GTTCTTGAACAGGACGTCATAGAGGGTGAAGATCCCGTTGGCAG GATACCTTCTCTGTAAGACTTTTCAAGAGTCGAGTTGGAA GCAGCTCAAAGTGGGTGTAATTCCATCTAAAGCTAAATATTGGCGAG AGACCGATAGCGAACAGTACAGTGAAAGATGAAAAGAACATTG AAAAGAGAGTGAAGGAAAGTACGTGAAATTGTTGAAGGGAAAGGCATT GATCAGACATGGTTTTGATGCACTCGCCTCTCGTGGCTGGGCC TCTCAAAAATTCACTGGCCAACATCAATTCTGGCAGTAGGATAAAC ATTAGAAATGTAGCTACCTCGGTAGTTAGCTTATTGAAACTG TAGCTGGATTGAGGACTGCGCTCGCAAGGATGTTGCATAATGGTT AAATGCCGCCGTCTGAA
	<i>Hanseniaspora opuntiae</i>	99%	ITS	CATTGGAGTTTTACTTAATTAAATTCTCTGCTTGAATCGAAAGG TTCAAGGCAAAAACAACACAAACAAATTATTATTATAATT AAACTAAACCAAAATTCTAACGGAAATTAAATTAAATTAAACTTT CAACAACGGATCTTGTCTCGCATCGATGAAGAACGTAACG CGATAAGTAATGTAATTGCGAGACTCTGTGAATCTGAAATT CGCACATTGCCCTTGAGCATTCTCAAGGGCATGCCTGTTGAGCGTC ATTCTCTCTCAAAGATAATTTTATTGGTGTGGCGATACTC AGGGTAGCTTGAATTGGAGACTGTTCACTGTTGAGTATT TAGCTTCTGGAGACGCTGTTCTCGCTGTGATGTATTATGGATT CGTTTACTTACAAGGAAATGTAATGTAACCTAGGAAAGGGT TGCTTTAAATATTCA
KB 4.NM NS1 (K36)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	ACCTGCGGAAGGATCATTAGATTGAATTATCATTGTCGAGTTCTA GTTTAGATCTTACAATAATGTGTATCTTATTGAAGATGTGCGCTTA ATTGCGCTGTTTAAAGTGTGCGAGTAGAAGTAATCTGCTTGAATC TCAGTCACGTTACACACATTGGAGTTTTACTTTAATTAAATTCTT CTGCTTGAATCGAAAGGTTCAAGGCAAAAACAACACAAATT TATTATTATAATTAAACTAAACCAAAATTCTAACGGAAATT AAAATAATTAAACCTCAACACGGATCTTGGTCTCGCATCGAT GAAGAACGTAACGCGAATTGCGATAAGTAATGTAATTGCGAGATACTCGT GAATCATTGAATTGTAACGCACATTGCCCTGAGCATTCTCAAGG GCATGCCTGTTGAGCGTCACTTCTCAAAGATAATTTTATT TTGGTTGTGGCGATACTCAGGGTTAGCTTGAATTGAAGATTGTTCA ATCTTATTGATGTTATTGAAATTGTTACTTACAAGGAAATGTAATG ACCTTACAGGAAAGGGTGCTTTAATATTCAAGTTGACCTCAAATC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 4.NM NS3 (K37)	<i>Hanseniaspora opuntiae</i>	99%	ITS	TTGGAGTTTTTACTTTAATTAACTTCTGCTTGAAATCGAAAGGTTCA CAAGGCACAAAACAACAAACAATTTTATTATAATTTTAA ACTAAACCAAATTCTAACCGGAATTAAATAATTAAAACCTTCAC ACAACGGATCTGGTCTCGCATCGATGAAGAACGTAGCGAATTCG ATAAGTAATGTGAATTGCAGATACTCGAATCATTAATTGAAACG CACATTGCGCCCTTGAGCATTCTCAAGGGCATGCCGTTGAGCGTC TTCTCTCAAAAGATAATTTTATTGGTTGAGCAGTCTTCAGTCTTAA GCTTCTGGAGACGCTGTCGCTGTGATGTATTGATTGATTTCG TTTACTTTACTAAGGGAAATGGAATGTACCTTAGGCAAAGGGTTGC TTTAATATTCA
KB 4.NM NS5 (K38)	<i>Hanseniaspora opuntiae</i>	100%	ITS	GGAGTTTTACTTTAATTAACTTCTGCTTGAAATCGAAAGGTTCA AGGCAAAAAACAAACACAAACAATTTTATTATAATTTTAAAC TAAACCAAAATTCTAACCGGAATTAAATAATTAAAACCTTCAC AACGGATCTGGTCTCGCATCGATGAAGAACGTAGCGAATTGCGAT AGTAATGTGAATTGCAGATACTCGAATCATTAATTGAAACG CATTGCGCCCTTGAGCATTCTCAAGGGCATGCCGTTGAGCGTCATT CTTCTCAAAAGATAATTTTATTGGTTGAGCAGTCTTCAGTCTTAA TAGCTGAAATTGGAGACTGTTCACTGCTGTGATGTATTGATTGATT TCTTGGAGACGCTGTCGCTGTGATGTATTGATTGATTGATTGCTT TACTTACAAGGGAAATGGAATGTACCTTAGGCAAAGGGTTGCTTTA ATATTCA
KB 4.NM NS7 (K39)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	TCCGTAGGTGAACCTGCGGAAGGATCATTAGATTGAAATTATTCA CTCGAGTTCTAGTTTAGATCTTACAATAATGTGATCTTATTGAAG ATGTGCGCTTAAATTGCGCTGTTTAAAGTGTGCAAGTAGAAGTAAT CTTGCCTGAATCTCAGTCACGTTACACACATTGGAGTTTTTACTT AATTAAATTCTCTGCTTGAATCGAAAGGTTCAAGGCAAAAACAAA CACAAACAATTTTATTATAATTAAACTAAACCAAAATTCT AACGGAAATTAAAATAATTAAAACCTTCACAAACGGATCTGGT TCTCGCATCGATGAAGAACGTAGCGAATTGCGATAAGTAATGTGAATTG CAGACTCTCGTAATCTGAATTGAACTTGAACGCACATTGCGCCTTGTAG CATTCTCAAGGGCATGCCGTTGAGCGTCATTCTCAAAAGATA ATTTTTATTGGTTGAGCAGTCTTCAGGTTAGCTTGAATTGA AGATTGTTCAATCTTTTAATTCAACACTAGCTTGGAGACGCT GTTCTCGCTGTGATGTATTGATTGAAATTATTGCTTACTTACAAGGG AATGGAATGTACCTTAGGCAAAGGGTTGCTTTAATATTCAAGT
KB 4.NM NS9 (K40)	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTTTTACTTTAATTAACTTCTGCTTGAAATCGAAAGGTTCAAGGC AAAAAACAAACACAAACAATTTTATTATAATTTTAAACTAA CCAAAATTCTAACCGGAATTAAAATAATTAAAACCTTCACAAACG GATCTCTGGTCTCGCATCGATGAAGAACGTAGCGAATTGCGATAAGT AATGTGAATTGCACTCGATGAAATTGCGATAAGTAATGTGAATTG CGGCCCTGAGCATTCTCAAGGGCATGCCGTTGAGCGTCATTCTC TCAAAGATAATTTTATTGGTTGAGCAGTCTTCAGGTTAGCTTGAATT TGAAATTGGAGACTGTTCACTGCTTAACTCAACACTAGCTTCT TGGAGACGCTTCTCGCTGTGATGTATTGATTGATTTCGTTACT TTACAAGGGAAATGGAATGTACCTTAGGCAAAGGGTTGCTTTAATAT TCATCAAGT
KA 0.CM S1 (K41)	<i>Wickerhamomyces anomalus</i>	100%	ITS	GCGGAAGGATCATTATAGTATTCTATTGCCAGCGCTTAATTGCGGGCG ATAAACCTTACACACATTGCTAGTTTTGAACCTTGCTTGGGGT GAGCCTGGCTACTGCCAACAGGCTAACACATTTTTAATGTTAA AACCTTAAACCAATAGTCATGAAATTAAACAAAATTAAACCTTC AAAACCTTCACAAACGGATCTTGGTCTCGCAACGATGAAGAACGCA CGGAATCGCATACTGATTGCAATTGCGAGATTTCGTGAATCATCGAA CTTTGAACGCACATTGCAACCTCTGGTATTCCAGAGGGTATGCC GAGCGTCATTCTCTCAAAACCTTCGGGTTGGTATTGAGTGTACT GTCAAGGGTTAACTGAAATAATTGATTAGGTTCTCCAACCTGTT GGCAGGTTAGAAGTATTAGGCTGGCTAACACAATAAACTAA GTTGACCTC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 0.CM S5 (K42)	<i>Wickerhamomyces anomalus</i>	100%	ITS	CGGAAGGATCATTAGTATTCTATTGCCAGCGCTTAATTGCGCGGC TAAACCTTACACACATTGTCTAGTTTTGAACTTGCCTTGGGTGCAT CAGCCTAGCTGCGTGCCTAAAGGTCAAACACATTTTAATGTTAAAAA CCTTTAACCAATAGTCATGAAAATTTCACAAAATTAAAATCTCAA AACTTCACAACACGGATCTCTGGTTCTCGAACGATGAAGAACGCAGC GAATATGCGATACGTATTGTGAATTGCAAGATTTCTGTGAATCATCGAATC TTGAAACGCCATTGACCCCTCTGGTATTCCAGAGGGTATGCCTTTG AGCGTCATTCTCTCAAACCTCGGGTTGGTATTGAGTGTACTCTG TCAAGGGTTAACTGAAATATTGACTTAGCAAGAGTGTACTAATAAGCA GTCTTCTGAATAATGTATTAGGTTCTCCAACCGTTATCAGCTAG GCAGGTTAGAAGTATTAGGCTCGGTTAACAAACAATAAA
KA 4.CM S1 (K43)	<i>Metschnikowia aff. fructicola</i>	99%	D1/D2 Domain	ACGGCGAGTGAAGCGCAAAGCTCAAATTGAAATCCCCGGGAATT GTAATTGAAAGAGATTGGTCCGGCGGGGGTAAAGTCCACTGGA AAGTGGCGCACAGAGGGTACAGCCCGTGAAACCCCTCAAAGCCCT CATCCCAGATCTCAAGAGTCGAGTTGGGAATGCAGCTAAAGTG GGTGTAAATCCATCTAAAGCTAAATACCGGCAGAGACCGATAGCG ACAAGTACAGTGAATGAAAGATGAAAGCACTTGAAAAGAGAGTGA AAAAGTACGTGAAATTGTTGAAAGGGAGGGCTGCAAGCAGACACTT AACTGGGCAGCATCGGGCGGGAAACAAAACCACGGGGAAATGT ACCTTCGAGGATTATAACCCCGTCTCAATTCTGTGCCCCGAGGC CTGCAATCTAAAGATGCTGGCGTAATGGTTGCAAGTCGCCGTCTGAA CCACGGA
Not identified			ITS	CAATCGGGGCCAGAGGTAAACAAACAAACAAATTATTATTCTTA AATTGTCAAAACAAGAATTGTAACTGAAATTAAAATATT AAAACCTTCACACGGATTGTTGGTCTCGCATCGATGAAGAACGC AGCAGAAATGCGATACGTAATGTGAATTGCAAGATTCTGTGAATCATCGA ATCTTGAAACGACATTGCGCCCTTGATTCGGGGCATGCGTGT TGAGCGTCATTCTTCAAAACCTTTGTTGTAGTGTAGTGTACTCT TGAGGTTAACTGAAATTGCTGGCTTCTATTGGATGTTTTTCCA AAGAGAGGTTCTTGCCTGCTGAGGTATAATGCAAGTACGGTGT TAGGTTTACCAACTGCGCTAATCTTTTACTGAGCGTATTGAA CGTTATCGATAAGAAGAGAGCGTCAAGGCAGAACATGTT
KB 4.CM S5 (K44)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TTAAAGAAATTAAATAATTGAAAATGGATTTTTGTTTGGCAAGAG CATGAGAGCTTTACTGGCAAGAAGACAAGAGATGGAGAGTCCAGCC GGGCTGCGCTTAAGTGCCTGGCTAGGCTTGTAAAGTTCTTCTT GCTATTCCAACCGTGAGAGATTGTTGCTTGTATAGGACAATT AAACCGTTCAATAACACACTGTGGAGTTTCATATTGCAACTTT TCTTTGGCATTGAGCAATCGGGGCCAGAGGTAACAAACACAAAC AATTATTATTATTCAAAACAAGAATTGTAAC GGAATTAAATATTAAACCTTCAACACGGATTGTTGTTCTC GCATCGATGAAGAACGCAGCGAAATGCGATACGTAATGTGAATTGCAAG AATTCCGTGAATCGAATCTTGAACGCACATTGCGCCCCCTGGTATT CCGGGGGCGATGCCCTGGTGGAGCTTCAACCTTT GGTAGTGAGTGATACTCTTGAGGTTACTGAAATTGCTGGCTTCA TGAGGTTTTTCAAAAGAGAGGTTCTTGCGTGTGAGGTAA TGCAAGTACGGCTTGAAGCTTACCAACTGCGCTAATCTTTTAA TACTGAGCGTATTGAAACGTTATCGATAAGAAGAGAGCGTCAAGGC ACAATGTTCTAAAGT
KA 2.NM S6 (K45)	<i>Saccharomyces cerevisiae</i>	99%	ITS	GGATCATTAAGAAATTAAATAATTGAAAATGGATTTTTGTTTGG CAAGAGCATGAGAGCTTTACTGGCAAGAAGACAAGAGATGGAGAGT CCAGCCGGGCGTGCCTTAAGTGCCTGGCTTGTCTAGGTTGTAAGTT CTTCTCTGATTCCAACCGTGGAGAGATTCTGTGCTTTGTTATAGGA CAATTAAACCGTTCAATAACACACACTGTGGAGGTTTCATATCTTGC AACTTTTCTTGGCATGGTGGAGCTTCAACACCGTGGAGGTT CAAACAAATTATTATTCAAAATTGTCAAAACAAGAATTG TAACGGAAATTAAATTTAAACCTTCAACACGGATTGTTGG TTCTCGCATCGATGAAGAACGCAGCGAAATGCGATACGTAATGTGAATT GCAGAATTCCGTGAATCATCGAATCTTGAGCGACATTGCGCCCCCTG GTATTCCGGGGCATGCTGTGAGCGTCAATTCTTCAACCTT TGTTGGTAGTGAGTGATACTCTTGAGGTTACTGAAATTGCTGGCT TTTCATTGGAGTTTTTCAAAAGAGAGGTTCTTGCGTGTGAGG TATAATGCAAGTACGGCTTTAGGTTTACCAACTGCGCTAATCTT TTTATACGAGCGTATTGAAACGTTATCGATAAGAAGAGAGCGTCAAGGC GCGAACATGTT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 2.NM S4 (K46)	<i>Saccharomyces cerevisiae</i>	100%	ITS	CCTCGGAAAGGATCATTAAGAAATTATAATAATTGAAAATGGATT TTGTGTTGGCAAGAGCATGAGAGCTTACTGGCAAGAACAGAG ATGGAGAGTCCAGCGGGCCTGCGTTAAGTGCAGGCTTGCTAGGCT TGTAAGTTCTTCTGCTATTCAAACGGTGAAGAGATTCTGCTTT GTTATAGGACAATTAAAACCCTTCATAACACACACTGTGGAGTTTC ATATTTGCAACTTTCTTGGCATTGAGCAATCGGAACTGGGGCCAGAG TAACAAACACAAACAATTATACTATTCAATTAAATTTGCAAAAC AAGAATTCTGTAACTGAAATTAAATATAAAACTTCAACAC GGATCTCTGGTCTCGCATCGATGAAGAACGCAAGCAGCAG TAATGTGAATGCAAGATTCCGTGAATCATCGAATCTTGAACGCACAT TGCAGCCCTGGTATTCCAGGGGAGCTGCTGTTGAGCGTCAATTCT CTCAACATCTGTTGGTAGTGAATCTTTGGAGTTAACCTG ATTGCTGCCCTTTCATTGGATGTTTCAAAAGAGAGGTTCTG CGTGTGAGGTATAATGCAAGTACGGCTGTTAGGTTACCAACTG CGGCTAATCTTTTATACTGAGCGTATTGGAACGTTATCGATAAGAA GAGAGCGTCTAGCGAACATGTTCTAAA
KA 4.NM S4 (K47)	<i>Saccharomyces cerevisiae</i>	100%	ITS	AAATTAATAATTGAAAATGGATTGTTGGCAAGAGCATGA GAGCTTACTGGCAAGAACAGACAAGAGATGGAGAGTCCAGCGGGC TGCCTTAAGTGCAGGCTTGCTAGGCTGTAAGTTCTTCTGCTAT TCCAAACGGTGAAGAGATTCTGCTTGTATAGGACAATTAAAC GTTCAATTAACACACTGAGGTTCTGCTATCTTGCACACTTTCTT TGGGCATTGCAAGCAATCGGGCCAGAGGTAACAAACACAA TATCTATTCAATTAAATTGTCAAAACAAGAATTCTGTAACGGAA ATTTAAATATTAAACACTTCAACACGGATCTTGCATCGC GATGAAGAACGAGCGAAATCGATACGTAATGTGAATTGCAAGAAC CGTGAATCATGAATTGCAACGCACATTGCCCTTGGTATTCCAG GGGGCATGCTGTTGAGCGTCAATTCTCTCAAACATTCTGTTGGTA GTGAGTGATACTTTGGAGTTACTGAAATTGCTGGCTTTCATGG ATGTTTCTTCAAAAGAGGTTCTGCGTGTGAGGTATAATGCA AGTACGGCTTTAGGTTACCAACTCGGGCTAATCTTTTATACT GAGCGTATTGGAACGTTATCGATAAGAAGAGAGCGTCAAGCGAACAA TGTTCTTAAAG
KA 4.NM S8 (K48)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGGCAAAGCTCAAATTGAAATCTGGTACCTCGTGCCCAGTTGAA TTTGGAGAGGGCAACTTGGGCGCTCTGCTATGTTCTTGGAAC AGGACGTATAGGGTGAAGAATCCCGTGGCAGGGAGTGC TTGTAAGTCCTCGAAGAGTCAGTTGGATGCACT TGGGTGTTAAATTCCATCTAAAGCTAAATTGGCGAGAGACCGATAGC GAACAAGTACAGTGTGAAAGATGAAAAGAACCTTGGAAAAGAGAGTG AAAAGTACGTGAATTGAAAGGGAGGGCATTTGATCAGACATG GTGTTTGTGCCCTCTGCTCTGGGGATGGGAATCTGCATTCT GGGCAGCATCAGTTGGTGCAGGATAATCCATAGGAATGTAGCT GCCTCGTAAGTATTATAGCTGTGGGAATACTGCCAGCTGGACTGAG GACTGCGACGTAAGTCAAGGATGCTGCCATAATGGTATATGCCCG TCTTGA
	<i>Saccharomyces cf. cerevisiae/paradoxus</i>	100%	ITS	GGACTCGGAAAGGATCATTAAGAAATTATAATAATTGAAAATGGATT TTTGTGTTGGCAAGAGCATGAGAGCTTACTGGCAAGAACAGAG AGATGGAGAGTCCAGCGGGCCTGCGTTAAGTGCAGGCTTGCT CTTGTAAAGTTCTTCTGCTATTCAAACGGTGAAGAGATTCTGCTT TTGTTATAGGACAATTAAAACGGTTCAATAACACACACTGTGGAGTT TCATATCTTCAACTTTCTTGGCATTGAGCAATCGGGCCAGA GGTAACAAACAAACAAATTATCTTAAATTGTCAAAA CAAGAATTCTGTAACTGGAAATTAAATATAAAACTTCAACAA CGGATCTTGGTCTCGCATCGATGAAGAACGCAAGCAG GTAATGTGAATTGCAAGAATTCCGTGAATCATCGAATCTTGAACGCACA TGCAGCCCTGGTATTCCAGGGGGCATGCTGTTGAGCGTCAATTCT TCTCAAACATCTGTTGGTAGTGAATCTTTGGAGTTAACCTG AATTGCTGCCCTTTCATTGGATGTTTCAAAAGAGAGGTTCT GCGTGTGTTGGTATAATGCAAGTACGGCTGTTAGGTTACCAACT CGGCTAATCTTTTATACTGAGCGTATTGGAACGTTATCGATAAGA GAGAGCGTCTAGCGAACATGTTCTAAAGTGTGACCTCAAAT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 4.NM S2 (K49)	<i>Saccharomyces cerevisiae</i>	99%	ITS	GGAAGGATCATTAAAGAAATTATAAATTGAAAATGGATTTTTGT TTGGCAAGAGCATGAGAGCTTACTGGCAAGAAGACAAGAGATGG AGAGTCAGCGGGCTCGCTTAAGTGCAGGGTCTGCTAGGCTTGTA AGTTCTTCTTGTATTCAAACGGTAGAGAGATTCTGTGCTTGT TAGGACAATTAAACCGTTCAATACAACACACTGTGGAGTTTCATAT CTTGCAACTTTCTTGGCATTGAGCAATCGGGCCAGAGGTTAA CAAACACAAACAAATTATCTATTCAAATTTGTCAAAAACAAAGA ATTTCTGAACGGAAATTAAAATATTAAAATTTCAACAACGGAT TCCTGGTCTCGCATGAGAACGCAGCGAAATGCGATACTGAA GTGAATTGAGAATCCGTGAATCATCGAATCTTGAACGCACATGCG CCCCTGGTATTCCAGGGGATGCCATTGGAGCTTACGCT AACATTCTGGTAGTGAGTAGTACTTTGGAGTTACTGAAATTG CTGGCTTTCATTGGATTTTTCAAAGAGAGGTTCTCGCTG CTTGAGGTATAATGCAAGTACGGCTTTAGGTTACCAACTGCGC TAATCTTTTAACTGAGCGTATTGAAACGTTATCGATAAGAAGAGA GCGCTAGGCGAACATGTTCAAAGT
KB 4.NM S10 (K50)	<i>Saccharomyces cerevisiae</i>	100%	ITS	TTAAAGAAATTATAAATTGAAAATGGATTTTTGTTGGCAAGAG CATGAGAGCTTTACTGGCAAGAAGACAAGAGATGGAGAGTCCAGCC GGGCTGCGCTTAAGTGCAGGGTCTGCTAGGCTGTAAGTTCTTCTT GCTATTCAAACGGTAGAGAGATTCTGTGCTTGTATAGGACAATT AAACGGTTCAATCAAACACACTGTGGAGTTCTATCTTGCAACTTT TCCTGGCATTGAGCAATCGGGCCAGAGGTAACAAACACAAAC AATTATCTATTCAAATTGTCAAAACAAGAATTCTGTAAC GGAAATTAAAATATTAAAATTTCAACAAACGGATCTTGGTCTC GCATCGATGAAGAACGCAGGAATGCGATACTGAAATTGCA AATTCCGTGAATCATCGAATCTTGAACGCACATTGGCCCCCTGGTATT CCAGGGGCGATGCCATTGGAGCGTCAACATTCTGTT GGTAGTGAGTAGTACTCTTGGAGTTACTGAAATTGCTGGCTTCA TGCAAGTACGGCTTTAGGTTACCAACTGCGCTAATCTT TACTGAGCGTATTGAAACGTTATCGATAAGAAGAGAGCGCTAGGCGA ACAATGTTCTAAA
KA 2.NM NS2 (K51)	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAAGGTACATTACATT TCCCTGTAAGTAAACGAATAATCCATAAATACATCACAGCGAGA ACAGCGTCTCAAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTTGAGTATGCCAACACAAAAATAA AAAATTATCTTGGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTCAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAATTATAATAAAAATAAATTGTT TGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAC
KA 2.NM NS4 (K52)	<i>Hanseniaspora uvarum</i>	100%	ITS	CCTTGCTAAAGGTACGTTACCATTCCTGTAAGTAAAAGAATAA ATCCATAAATACATCACAGCGAGAACAGCGTCTCAAAGAAGCTAAGT GTGAATTAAAAAGACTGAAACAGTCTCAATTCAAGCTAACCTGA GTATGCCAACACAAAAATAAATTATCTTGTGAGAAGGAAATG ACGCTAACACAGGCATGCCCTGAGAACGCTCAAAGGGCGAATGCG TCAAAATTCAATGATTACGAGTATCTGCAATTCACTTATCG AATTGCTACGTTCTCATCGATGCGAGAACCAAGAGATCCGTGTTGA AAGTTAAATTATTTAAATTCCGTAGGAATTGGTTAGTTAA AAAATTATAAAAATAAATTGTTGTTGTTGCTTGAACCT TTCGATTCAAAGCAGAAAGAATTAAATTAAAGTAAAAAC
KA 2.NM NS8 (K54)	<i>Hanseniaspora uvarum</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAAGGTACGTTACATT TCCCTGTAAGTAAACGAATAATCCATAAATACATCACAGCGAGA ACAGCGTCTCAAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTTGAGTATGCCAACACAAAAATAA TAAATTATCTTGGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTCAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAATTATAATAAAAATAAATTGTT TGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAC

Table A.1. (*continued*)

Name	BLAST Result	Similarity	Region	Sequence
KB 2.NM NS9 (K60)	<i>Hanseniaspora opuntiae</i>	100%	ITS	ACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCA TTCCCTGTAAGTAAAAGCAATAAAATCATAAATACATCACAGCGAGA CAGCGCTCCAAGAACGACTAAGTGTGAATTAAAAAGACTGAACAG TCTCCAATTCAAGCTAACCCGTAGATTCGCCACAACCAAAAAATAAA AAATTATCTTGAGAAGGAATGACGCTCAAACAGGCATGCCCTGAG AATGCTCAAGGGCGCAATGTGCGTTCAAAATTCAATGATTACGAGTA TCTGCAATTTCACATTACTCGCAATTGCTACGTTCTCATCGATGCG AGAACCAAGAGATCCGTGTTGAAAGTTAAATTATTTAAAATTTC GTTAGGAATTGGTTAGTTAAAAAATTATAATAAAAATTGTT GTGTTGTTGTTTGCTGACCTTCGATTCAAGCAGAAAGAATTAA ATTAAAGTAAAAAAC
KA 4.NM NS1 (K61)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	GTCAACTTGTGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCA ATTCCCTGTAAGTAAAAGCAATAAAATCATAAATACATCACAGCGA GAACAGCGCTCCAAGAACGACTAAGTGTGAATTAAAAAGATGAAA CAATCTCAATTCAAGCTAACCCGTAGATTCGCCACAACCAAAAAAT AAAAATTATCTTGAGAAGGAATGACGCTCAAACAGGCATGCCCTGAG AAGATGCTCAAGGGCGCAATGTGCGTTCAAAATTCAATGATTACGAG ATGATCTGCAATTCACTTACTCGCAATTGCTACGTTCTCATCGA TGCGAGAACCAAGAGATCCGTGTTGAAAGTTAAATTATTTAAAAT TCCGTTAGGAATTGGTTAGTTAAAAAATTATAATAAAAATTGTTGTTGTTGTTGTTGTTGCTGACCTTCGATTCAAGCAGAAAGAATTAAATTAAAGTAAAAAATTCCAACTGTGTAACGTTGACTGAGATT CAAGCAAGGATTACTCTACTGCGACACTTAAAGAACGCGCAATTAA GCGCACATCTCAATAAAGATAACACATTATTGTTAAAGATCTAAAAC TA GACTCGAGCAACAATGATAATTCAATCTAATGATCCTCCGCAGGT
KA 4.NM NS3 (K62)	<i>Hanseniaspora opuntiae</i>	100%	D1/D2 Domain	CGGCGAGTGAAGCGGTAAGCTCAAATTGAAATCTGGTACTTCAGT GCGCGAGTGTAAATTGAGAATTGCTTGTGATTAGTCCTGTCTATG TTCCCTGGAACAGCAGCTCATAGGGTGAGAATCCCGTTGGCGAGG ATACCTTTCTGTAAGACTTTCGAAAGAGTCGAGTTGTTGGAATG CAGCTCAAAGTGGGTGTAATTCCATTAAGCTAAATTGGCGAGA GACCGATAGCGAACAGTACAGTGATGAGAAGATGAAAAGAACATTGAA AAAGAGAGTGAAGGACTCGTAATTGTTGAAAGGAGAAGGGCATTG ATCGACACATGGTTTGGCATGCACTCGCCTCTGGCTGGCGCTCTCA TCAAAATTCACTGGGCAACATCAATTGCGAGTAGGATAATGCT TTAAGAATGTAGTACCTCGGTAGTGTATGCTTATTGAAATGCT AGCTGGGATTGAGGACTGCGCTTCGCAAGGATGTTGCGATAATGGTTA AATGCCGCCGCTTGA
	<i>Hanseniaspora opuntiae</i>	100%	ITS	TTGAGGTCAACTTGTGAATATTAAAAGCAACCCTTGCCTAAGGTACA TTACCATTCCTGTAAGTAAAAGCAATAAAATCATAAATACATCAC AGCGAGAACAGCGCTCCAAGAACGACTAAGTGTGAATTAAAAAGAC TGAAACAGTCTCAAATTCAAGCTAACCCGTAGATTCGCCACAACCAA AAAAATTATCTTGAGAAGGAATGACGCTCAAACAGGCAT GCCCTGAGAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGAT TCACGAGTATGCTCAATTCACTTACTCGCAATTGCTACGTTCTTC ATCGATGCGAGAACCAAGAGATCCGGTTGAAAGTTAAATTATTTT AAAATTCCGTTAGGAATTGGTTAGTTAAAAAATTATAATAAAAATTGTT AAAATTGTTGTTGTTGTTGCTGACCTTCGATTCAAAAGCAGAAGAATTAA AGAATTAAATTAAAGTAAAAAA
KA 4.NM NS5 (K63)	<i>Hanseniaspora opuntiae</i>	99%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCA TTCCCTGTAAGTAAAAGCAATAAAATCATAAATACATCACAGCGAGA ACAGCGCTCCAAGAACGACTAAGTGTGAATTAAAAAGACTGAACACA GTCTCCAATTCAAGCTAACCCGTAGATTCGCCACAACCAAAAAATAAA AAAATTATCTTGAGAAGGAATGACGCTCAAACAGGCATGCCCTGAGA GAATGCTCAAGGGCGCAATGTGCGTTCAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTCGCAATTGCTACGTTCTCATCGATGCGAGA GAGAACCAAGAGATCCGGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAAAATTATAATAAAAATTAAATTGTT TTGTTGTTGTTTGGCTGACCTTCGATTCAAGCAGAAAGAATTAA ATTAAAAGTAAAAAAACTCCAATGTGTAACGTTGACTGAGATTCAAGCAGA GCAAGACTACTCTACTGGGACACTCTAATAAAAGCAGCGCAATTACCGC ACATCTTCATTAAAGATAACACATTATTGTTAAAGATCTAAACAAAGAAC TCGAGCAACAATGATAATTCAATCTAATGATCCTCCGCAGGT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KA 4.NM NS7 (K64)	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACATT TCCCTTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGA ACAGCGTCTCCAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTGAGTATGCCAACACCAAAAAATAA AAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCGTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAAAAATTGT TTGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAA
KA 4.NM NS9 (K65)	<i>Hanseniaspora opuntiae</i>	99%	D1/D2 Domain	ACGGCGAGTGAAGCGTAAAAGCTCAAATTGAAATCTGGTACTTCAG TGCCCAGTTGTAATTGAGAATTGCTTGTATTAGGCCTTGTCTAT GTTCCTTGAACAGGACGTCATAGAGGGTGAGAATCCCGTTGGCAG GATACCTTTCTGTAAAGACTTTTCAAGAGTCGAGTTGGAAAT GCAGCTCAAAGTGGGTGTTAAATTCCACTAAAGCTAAATATTGGCAG AGACCGATAGCGAACAAAGTACAGTGAGGAAAGATGAAAAGAACATTG AAAAGAGAGTAAAAAGTACGTGAAATTGTTGAAAGGGAGGGCATT GATCAGACATGGTGTGTTTGATGCACTCGCCTCTGTTGGCCTGGC TCTCAAAAATTTCACTGGGCCAACATCAATTGCGTAGGATAAAC ATTAAGAATGTAGTACCTCGGTAGTTAGCTTATTGAAACTG TAGCTGGGATTGAGGACTGCGCTTGGCAAGGATGTTGGCATAATGGTT AAATGCCGCCGCTTGAACACCGAC
	<i>Hanseniaspora opuntiae</i>	99%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACATT TCCCTTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGA ACAGCGTCTCCAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTGAGTATGCCAACACCAAAAAATAA AAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCGTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAAAAATTGT TTGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAAACTCCAATGTGTTAACAGTGTGAGATTCAAG CAAGACTACTCTACTGCGACACTCTAATGAAGCAGCGCAATTAGC ACATCTCAAATAAGATACACATTATTGTTAAAGATCTAACAGAAACT CGAGCAACAATGATAATTCAATCTAATGATCCTCCGAGGTTCCCT
KB 4.NM NS2 (K66)	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACATT TCCCTTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGA ACAGCGTCTCCAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTGAGTATGCCAACACCAAAAAATAA AAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCGTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAAAAATTGT TTGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAA
KB 4.NM NS4 (K67)	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACATT TCCCTTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGA ACAGCGTCTCCAAGAAGCTAAGTGTGAATTAAAAAGACTGAAACA GTCTCCAATTCAAGCTAACCTGAGTATGCCAACACCAAAAAATAA AAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGA GAATGCTCAAGGGCGCAATGTGCGTTAAAATTCAATGATTACGAGT ATCTGCAATTCACTTACTATCGCAATTGCGTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTTGAAAGTTAAATTATTTAAAATTTC CGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAAAAATTGT TTGTGTTGTTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATT AATTAAAGTAAAAAC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KB 4.NM NS6 (K68)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCAATTCCCTTGTAAAGTAAAAGCAATAAATTCTAAATACATCACAGCGAGAACAGCGTCTCCAAGAGAAGCTAAGTGTGAAATTAAAAAGATTGAAACAATCTTCAATTCAAGCTAACCCCTGAGTATCGCCCACAAACCAAAAAATAAAAAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGAGATGCTCAAGGGCGCAATGTGCGTCAAAAATTCAATGATTACAGACTCTGCAATTACAGCTTCAAGGGTTAGTTAAAAAAATTATAATAAAAATAAAATTGTCTGTGTTGTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATTAATTAAAGTAAAACCTCAATGCGACACTTAAAAAGCAGCGCAATTAAAGCAGCATCTICAATAAGATAACACATTATGAAAGATCTAAACTAGAACTCGAGCAACAATGATAATTCAATCTAATGATCCTCCGCAGGTTCA
KB 4.NM NS8 (K69)	<i>Hanseniaspora guilliermondii</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCAATTCCCTTGTAAAGTAAAAGCAATAAATTCTAAATACATCACAGCGAGAACAGCGTCTCCAAGAGAAGCTAAGTGTGAAATTAAAAAGATTGAAACAATCTTCAATTCAAGCTAACCCCTGAGTATCGCCCACAAACCAAAAAATAAAAAAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGAGATGCTCAAGGGCGCAATGTGCGTCAAAAATTCAATGATTACAGACTCTGCAATTACAGCTTCAAGGGTTAGTTAAAAAAATTATAATAAAAATAAAATTGTCTGTGTTGTTGCCTGAACCTTCGATTCAAAGCAGAAAGAATTAATTAAAGTAAAACCTCAATGCGACACTTAAAAAGCAGCGCAATTAAAGCAGCATCTICAATAAGATAACACATTATGAAAGATCTAAACTAGAACTCGAGCAACAATGATAATTCAATCTAATGATCCTCCGCAGGTTCA
KB 4.NM NS10 (K70)	<i>Hanseniaspora uvarum</i>	98%	D1/D2 Domain	TACGGCGAGTGAGCGGTAAGCTCAAATTGAAATCTGGTACTTTAGCTGGCCCGAGTTGAATTGAGAATTGTTGATTAGGTCTTGTCTATGTTCTTGGAAACAGGACGTCATAGAGGGTGAGAAATCCGTTGGCAGGATACCTTTCTCTGTAAGACTTTCTGAAGAGTCGAGTTGTTGGAAATCAGCTCAAAGCTAATATTGGCAGAGCCGATAGCGAACAGTACAGTGATGAAAGATGAAAGAAACTTGAAGAGAGTGAAAAGTAAGTGAAATTGTAAGGGAAAGGGCATTGATCAAACACTGGTGTGTTTGATGCACTCCCTCTCGGGGCTTGGGCCTCTCAAATT
	<i>Hanseniaspora uvarum</i>	98%	ITS	GAATATTAAAAGCAACCCTTGCCTAAGGTACATTACCAATTCCCTTGTAAAGTAAAAGCAATAAATTCTAAATACATCACAGCGAGAACAGCGTCTCCAAGAGCTAAGTGTGAAATTAAAAAGATTGAAACAACTCTCAATTCTGTAACCCCTGAGTATCGCCCACAAACCAAAAAATAAAATTATCTTTGAGAAGGAAATGACGCTAACACAGGCATGCCCTGAGAATGCTCAAGGGCGCAATTGCGTCAAAAATTCAATGATTCAAGCTACAGTGATGTTGAGAATTCTGCAATTCTGCAACTTCTGCAATTCTGCTACGTTCTCATGATGCGAGAACCAAGAGATCCGTTGTTGAAAGTTAAATTATTTAAAATTCCGTTAGGAATTGGTTAGTTAAAAAAATTATAATAAAAATAAAATTGTTGTTGTTGTTGCTTGAACCTTCGATTCAAAGCAGAAAGAATTAAATTAAAGTAAAACCTCAATGTTGAAACAGTGACTGAGATTCAAGCAAGATTCTACTGCGACACTTAAAGAAGCAGCGCAATTAGCGCACATTCTCAATAAGATAACACATTATGAAAGATCTAAACTAGAAACTCGAGCAACAATGATAATTCAATCTAATGATCCTCCGCAGGTTCA
KA 32.NM S2 (K71)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGCGAGTGAGCGGAAAAGCTCAAATTGAAATCTGGTACCTTCGGTCCCCGAGTTGAATTGGAGAGGGCAACTTGGGGCCGTTCTTGTCTATGTTCTTGGAAACAGGACGTCATAGAGGGTGAGAAATCCCGTGTGGCGAGGAGCTGTTCTTGTAAAGTGCCTCGAAGAGTCGAGTTGTTGGGAATGCGACTCTAAGTGGGTGTTAAATTCCATCTAAAGCTAATATTGGCGAGAGACCGATAAGCGAACAAAGTACAGTGATGAAAGATGAAAGAAACTTGTGAAAGAGTGAAGAAAGTACGTGAAATTGTTGAAAGGGAAAGGGCATTTGATCAGACATGGTGTGTTGCCCCCTGCTCCTGTGGGTAGGGGAATTCTGCATTCACTGGGCCAGCATCAGTTGGTGGCAGGATAAATCCATAGGAATGTAGCTGCTCGGTAAAGTATTATAGCTGTGGGAATACTGCCAGCTGGGACTGAGGACTGCGACGTAAGTCAAGGATGCTGGCATATGGTTATATGCCGCCGTT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
KA 4.NM S2 (K72)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	GGCAAAGCTCAAATTGAAATCTGGTACCTTCGGTCCCCGAGTTGAA TTGGAGAGGGCAACTTGGGCCGTCCCTTGCTATGTTCTTGGAAC AGGACGTCATAGAGGGTGAAGAATCCCGTGTGGCGAGGAGTGCAGTCT TTGTAAGTGCCTCGAAGAGTCAGTTGTTGGATGCAAGCTAAG TGGTGGTAATTCATCTAAAGCTAAATTGGCAGAGACCGATAGC GAACAAGTACAGTGAAGATGAAAAGAAGACTTTGAAAAGAGAGTG AAAAGTACGTGAAGAGGGCATTTGATCAGACATG GTGTTTGCCCTCTGCCTTGTGGTAGGGGAATCTGCATTCACT GGCCAGCATCAGTTGGCAGGATAATCCATAGGAATGTAGCTT GCCTCGGTAAAGTATTAGCCTGTGGGAATACTGCCAGCTGGACTGAG GACTGCGACGTAAGTCAGGATGCTGCCATAATGGTTATATGCCGCCG TCTTG
EA 0.W NS1 (E1)	<i>Saccharomyces cerevisiae</i>	96%	ITS	AACATTGTCGCCAGACGCTCTTCTTATCGATAACGTTCCAATACGC TCAGTATAAAAAAAGATTACCCGCACTGGTAAACCTAAACGACCG TACTTGCATTATACCTAACCGCAACCGCAAAAAACCTCTTTGGAAAAAA AACATCCAATGAAAAGGCCACCAATTCAAGTTAACCTCAAAGTATC ACTCACTACCAACAAAATGTTGAAAGGAATGACGCTAAACAGG CATGGCCCTGGAATACCAAGGGCGCAATGTGCGTTCAAAATTCGAT GATTACCGGAATTCTGCATTACATTACGTATCGCATTTCGCTGCGTC TTCATCGATCGAAAACCAAAAAATCCGGTTGAAAGTTTTAATT TTAAAATTCCAGTTACAAAATTCTGTTGAAAGAAAATTAAATGAA TAGATAAAAATTGTTGTTGTTACCTCTGGGCCCCGATTGCTCGAATG CCCAAAAAAAAGTTGCAAAATATGAAAACCTCACAGTGTGTTGATT GAAACGGTTTAATTGCTATAACAAAACCACAAAATCTCTCACCGT TIGGAATACCAAAAAAAACTACACCCCTACCAAAACGCCACTT AACCGCAGGGCCGGCTGGACTCTCATCTCTGTTCTGCCCAGTAAA ACCTCTCATGCTCTGCCAAAACAAAAAATCCATTTCAAAATTATT AATTCTTAATGATCCTCCGCAGGT
EA 0.W NS2 (E2)	<i>Saccharomyces cerevisiae</i>	96%	ITS	ACATTGTCGCCAGACGCTCTTCTTATCGATAACGTTCCAATACGCT CAGTATAAAAAAATTACCCGCACTGGTAAACCTAAACGACCGT ACTTGCATTATACCTAACCGCAACCGCAAAAAACCTCTTTGGAAAAAA ACATCCAATGAAAAGGCCACCAATTCAAGTTAACCTCAAAGTATCA CTCACTACCAACAAAATGTTGAAAAGGAATGACCCCTAAACAGGC ATGCCCTGGAATACCAAGGGCGCAATGTGCGTTCAAAATTCGATG ATTACCGGAATTCTGCATTACATTACGTATCGCATTTCGCTGCGTTCT TCATCGATCGAAAACCAAAAAATCCGGTTGAAAGTTTTAATT TAAAATTCCAGTTACAAAATTCTGTTGAAAGAAAATTAAATGAA AAATAAAATTGTTGTTGTTACCTCTGGGCCCCGATTGCTCGAATG CCCAAAAAAAAGTTGCAAAATATGAAAACCTCACAGTGTGTTGATT GAAACGGTTTAATTGCTATAACAAAACCACAAAATCTCACCGT TIGGAATACCAAAAAAAACTACACCCCTACCAAAACGCCACTT AACCGCAGGGCCGGCTGGACTCTCATCTCTGTTCTGCCCAGTAAA ACCTCTCATGCTCTGCCAAAACAAAAAATCCATTTCAAAATTATT AATTCTTAATGATCCTCCGCAGT
EA 0.W NS4 (E4)	<i>Saccharomyces cerevisiae</i>	97%	ITS	AACTTAAGAACATTGTCGCCAGACGCTCTTCTTATCGATAACGTT CCAATACGCTCACTGGTAAACCTAAACGACCGCAACCGCAACCTCTT AAACGACCGTACTTCGATTACCTAACCGCAAGCACGCAACCGCAACCTCTT TGGAAAAAAACATCCAATGAAAAGGCCACCAATTCAAGTTAACCT AAAAAGTATCACTACCAACAGAATGTTGAAAAGGAATGACG CTCAAACAGGCATGCCCTGGAATACCAAGGGCGCAATGTGCGTC AAAATTCGATGATTACCGGAATTCTGCATTACATTACGTATCGCAT TTCGCTCGTTCTCATCGATGCGAAAACCAAAAAATCCGGTTGAA GTTTTAATTAAAATTCCAGTTACGAAAATTCTGTTGAA AAATTAAATGAATAGATAAAATTGTTGTTGTTACCTCTGGGCCCCG ATTGCTCGAATGCCAAAAAAAGTTGCAAAATATGAAAACCTCAC AGTGTGTTGATTGAAACCGTTTAATTGCTTAAACAAAACCAAAA AACTCTCACCGTTGGAATACCAAAAAAAACTACACCCCTACAC AACCGCCCACTAACCGCAGGCCGGCTGGACTCTCATCTCTGTTTC TTGCCAGTAAAACCTCTCATGCTCTGCCAAAACAAAAAATCCATT TCAAAATTAAAATTCTTAATGATCCTCCGCAGGT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EA 0.W NS5 (E5)	<i>Saccharomyces cerevisiae</i>	96%	ITS	AACTTTAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTT CCAATACGCTCAGTATAAAAAAAAATTACCCGCAGTTGGTAAAACCTA AAACGACCGTACTTGCATTATACCTCAACCACGCAAAAAAAAACCTCTCTT TGGAAAAAAAACATCCAATGAAAAGGCCACCAATTCAAGTTAACCTCC AAAAAGTATCACTACCAACAAAATGTTGAAAAGGAAATGACG CTCAAACAGGCATGCCCTGGAATACCAAGGGGCGCAATGTGCCTTC AAAATTGCAATGATTACCGGAATTCTGCAATTACATTACGTTACCGAT TTCGCTGCTCTTCATCGATGCGAAAACCCAGGCCCCGCTGGACTCTCCATCTTGTGTTTC ATTGCTGAATGCCCAAAAAAAAAGTGCACAAATGAAACTCCAC AGTGTGTTGATTGAAACGGTTTAATGTCCTATAACAAAACCACAAA AATCTCTACCGTTGAATACCAAAAAAAACTTACAACCCCTACAC AACCAGCCACTTAACCCAGGCCGGCTGGACTCTCCATCTTGTGTTTC TTGCCAGTAAACCTCATGCTCTGCAAACAAAAAAATCCATT TCAAAATTATTAATTCTTAATGATCCTCCGCAGGTT
EB 0.W NS1 (E6)	<i>Saccharomyces cerevisiae</i>	99%	ITS	AACTTTAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTT CCAATACGCTCAGTATAAAAAAGATTAGCCGCAGTTGGTAAAACCTA AAACGACCGTACTTGCATTATACCTCAAGCACGCAGAGAAACCTCTT TGGAAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACCTCC AAAGAGTATCACTACCAACACAGAATGTTGAGAAGGAAATGACG CTCAAACAGGCATGCCCTGGAATACCAAGGGGCGCAATGTGCCTTC AAAGATTGATGATTACCGGAATTCTGCAATTACATTACGTTACCGAT TTCGCTGCTCTTCATCGATGCGAGAACCAAGAGATCCGTTGAAAG GTTTTAATATTTAAAATTCCAGTTACGAAAATTCTGTTTGTGACAA AAATTAAATGAAATGATAAAAATTGTTGTTGTTACCTCTGGGGCCCG ATTGCTGAATGCCAAAAGAAAAAGTGCACAGGAAATGACCA AGTGTGTTGATTGAAACGGTTTAATGTCCTATAACAAAAGCAGA AATCTCTACCGTTGAATAGCAAGAAAGAAACTTACAAGCCTAAC AGACCGCGCACTTAACCGCAGGCCGGCTGGACTCTCCATCTTGTCT TTGCCAGTAAACCTCATGCTCTGCAAACAAAAAAATCCATT TTCAAAATTATTAATTCTTAATGATCCTCCGCAGGTT
EB 0.W NS2 (E7)	<i>Saccharomyces cerevisiae</i>	95%	ITS	ACCTTAAAGAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTT CAATACGCTCAGTATAAAAAAAAATTACCCGCAGTTGGTAAAACCTAA AACGACCGTACTTGCATTATACCTCAACCACCCAAAAAAACCTCTTT CGAAAAAAAACATCCAATGAAAAGGCCACCAATTCAAGTTAACCTCC AAAAGTATCACTACCAACACAAATGTTGAAAAGGAAATGACCC TCAACACGGCATGCCCTGGAATACCAAGGGGCCAATGTGCCTCA AAAATTGATGATTACCGAATTCTGCAATTACATTACGTTACCGATT TCGCTGCTCTTCATCGATGCGAAAACCAAAAAATCCGTTGAAAG TTTTAATATTTAAAATTCCAGTTACAAAATTCTGTTTGTGACAAA AATTAAATGAAATAAAAATTGTTGGGTTGTTACCTCTGGGCCCG ATTGCTGAATGCCAAAAGAAAAAGTGCACAAATGAAACTCCAC AGTGTGTTGATTGAAACGGTTTAATGTCCTATAACAAAACCACAAA AATCTCTACCGTTGAATACCAACAAAAAAACTTACAACCCCTACAC AACCAGCCACTTAACCGCAGGCCGGCTGGACTCTCCATCTTGTGTTTC TTGCCAGTAAACCTCATGCTCTGCAAACAAAAAAATCCATT CAAAATTATTAATTCTTAATGATCCTCCGCAGTTACTAC
EB 0.W NS3 (E8)	<i>Saccharomyces cerevisiae</i>	99%	ITS	AAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCAAATCG CTCAGTATAAAAAAGATTAGCCGCAGTTGGTAAAACCTAAAAGACC GTACTTGCATTATACCTCAAGCACGCAGAGAAACCTCTTGGAAAAAA AAACATCCAATGAAAAGGCCACCAATTCAAGTTAACCTCAAAGAGTA TCACTCACTACCAACACAGAATGTTGAGAAGGAAATGACGCTCAAACA GGCATGCCCTGGAATACCAAGGGGCCAATGTGCCTCAAAGATT GATGATTACCGAATTCTGCAATTACATTACGTTACGCTTC GTTCTTCATCGATGCGAGAACCAAGAGATCCGTTGAAAGTTTAA TATTTAAAATTCCAGTTACGAAAATTCTGTTGACAAAAATTAA TGAATAGATAAAAATTGTTGTTGTTACCTCTGGGCCGATTGCTCG ATGCCCAAAGAAAAAGTGCACAGGAAATGAAACTCCACAGTGTGTT GTATTGAAACGGTTTAATTGTCCTATAACAAAAGCAGAAATCTCTC ACCGTTGGAAATAGCAAGAAAGAAACTTACAAGCCTAACAGACCGC CACTTAACCGCAGGCCGGCTGGACTCTCCATCTTGTCTTGTGTT GTAAAAGCTCATGCTCTGCAAACAAAAAAATCCATTTCAAAAT TATTAATTCTTAATGATCCTCCGCAGGTTACCTACGGAAAGGATC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EB 0.W NS4 (E9)	<i>Saccharomyces cerevisiae</i>	96%	ITS	AACTTTAAGAACATTGTCGCCTAGACGCTCTCTTATCGATAACGTTCCAATACGCTCAGTATAAAAAAAAATTACCGCAGTTGGTAAAACCTAAACGACCGTACTGCATATACTCTCAACCACGCAAAAAAAAACCTCTTTGAAAAAAACATCCAATGAAAAGGCCACCAATTCAAGTTAACTCCAAAAAGTATCACTCACTACCAACAAAATGTTGAAAAGGAAATGACGCTCAAACAGGCATGCCCTGGAATACCAAGGGGCCAATGCGTTCAAAAATTGGATTCACCGGAATTCTGCACATTACATTAACGTATCGCATTTCGCTCGTTCTCATGCATGCGAAAACCAAAAAATTCTGTTGAAAGTTTTAATATTTAAAATTTCAGTTACAAAATTCTGTTGACAAAATTAAATGATAAATAAAAATTGTTGGTTGACTCTGGGCCCGATTGCTCAAATGCCAAAAAAAGTGCACAAAATATGAAAACCTCACAGTGTGTTGATTGAAACGGTTTAAATTGCTCTATAACAAAACCCAAAATCTCTACCGTTGGAATACCAAAAAAAACTACAACCTACAAACCGCCACTTAACCCAGGCCGCTGGACTCTCCATCTCTGCTCTTGCCCAGTAAACCTCTCATGCTCTGCAAACAAAATCCATTCTCAAATTATTAATTCTTAAATGATCCTCCGCAGGTT
EA 1.W NS1 (E11)	<i>Hanseniaspora uvarum</i>	100%	ITS	AACTTGATGAATATTAAGCAACCCCTTGCTAAGGTACGTTACCAATTCCCTTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGAACAGCGTCTCCAAAGAAGACTAAGTGTGAAATTAAAAAAAGACTGAAACAGTCCAATTTCAGCTAACCCCTGAGTATGCCACACCAAAAAATAATAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGAGAATGCTCAAGGGCAGTTCACGGTCAATTCTACATTACTATCGCAATTGCTACGTTCTCATCGATGCAGAAACCAAGAGATCCGTTGAAAGTTTAAATTATTTAAAATTTCAGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAATTGTGTTGTTGTTGCTGAAACCTTGCAGTCAAAGCAGAAAGAATTAAATTAAAGAAAACCTCAATGTTGAGTGTGAGCAGGTTGACTGAGATTCAGCAAGACTACTTCACTCGCAGACTCTAAAGAAGCAGCGCAATTAGCAGCATCTTCAAAAGATAACACATTATTGTTAAAGATCTAAACAAGAAACTCGAGCAACAATGATAATTCAATCTGATCCTCCGCAGGTT
EA 1.W NS2 (E12)	<i>Hanseniaspora uvarum</i>	100%	ITS	AACTTGATGAATATTAAGCAACCCCTTGCTAAGGTACGTTACCAATTCCCTGAAAGTAAAAGCAATAATCCTAAATACATCACAGCGAGAACAGCGTCTCCAAAGAAGACTAAGTGTGAAATTAAAAAAAGACTGAAACAGTCCAATTTCAGCTAACCCCTGAGTATGCCACACCAAAAAATAATAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGAGAATGCTCAAGGGCAGTTCACGGTCAATTCTACATTACTATCGCAATTGCTACGTTCTCATCGATGCAGAAACCAAGAGATCCGTTGAAAGTTTAAATTATTTAAAATTTCAGTTAGGAATTGGTTAGTTAAAATTATAATAAAAATAATTGTGTTGTTGTTGCTGAAACCTTGCAGTCAAAGCAGAAAGAATTAAATTAAAGTAAAACCTCAAAGTGTGAGTGTGAGGTTGACTGAGATTCAGCAAGACTACTTCACTCGCAGACTCTAAAGAAGCAGCGCAATTAGCAGCATCTTCAAAAGATAACATCTTCAAAAGATACTTAAACAGAAACTCGAGCAACAATGATAATTCAATCTGATCCTCCGCAGGTT
EA 1.W NS3 (E13)	<i>Hanseniaspora uvarum</i>	100%	ITS	ACGCCCTTGCTAAGGTACGTTACCATTCCTTGAAAGTAAAAGAAATAATCCAAATACATCACAGCGAGAACAGCGTCTCCAAAGAAGCTAAGTGTGAAATTAAAAAAAGACTGAAACAGTCTCCAATTTCAGCTAACCCAGTGTATGCCACACCAAAAAATAATAATTATCTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGAGAATGCTCAAGGGCGCAATGTGCGTCAAATTCAATGATTCACTCGAGTATCTGCAATTTCACATTACTTAAGTCAATTGCTACGTTCTCATGCATGCGAGAACAGAAGAGATCCGTTGAGTAAAGTTTAAATTTCAGTTGAGAATTGGTTAGTTAAAAAAATTATAATAAAAATAATTGTGTTGTTGTTGCTTGAACCTTGCAGTCAAAGCAGAAAGAATTAAATTAAAGTAAAACCTCAAATGTTGAGTGTGAGGTTGACTGAGATTCAGCAAGCAAGACTACTTCACTCGCAGACTCTAAAGAAGCAGCGCAATTAGCAGCACATCTCAATAAAAGATAACATTATTGTTAAAGATCTAAACAGAAACTCGAGCAACAATGATAATTCAATCTGATCCTCCGCAGGTT

Table A.1. (*continued*)

Name	BLAST Result	Similarity	Region	Sequence
EA 1.W NS4 (E14)	<i>Hanseniaspora uvarum</i>	100%	ITS	CCTTGCCTAAGGTACGTTACCACTTCCCTGTAAGTAAAGGAAATTAATCATAATACATCACAGCGAGAACAGCGTCTCAAAGAAGCTTAAGTGTGAAATTAAAAGACTGAAACAGTCTCAATTCAAGTCACCCCTGAGTACGCCAACACCAAAAAATAATAAAATTATCTTGTAGAAGGAAATGACGCTAACACAGGGCATGCCCTGAGAATGCTCAAGGGCGCAATGTGCGTTCAAAAATTCAATGATCACAGTACGCTTACATCGATCGAGAACAGAACAGATCCGTTGTGAAATTCTGCTAACCTAAGTAAATTATTTAAAATTCCGTAGGAATTGGTTAGTTAAAAAATTATAAATAAAAATTGTTGTGTTGTGTTTGCTTGAACCTTTCGATTCAAAGCAGAACAGAATTAAGTAAACACTCAATGATAATTCAACATTAATGATCCTCCGAGGT
EA 1.W NS5 (E15)	<i>Saccharomyces cerevisiae</i>	97%	ITS	GAGGTCAACTTAAAGAACATTGTCGCCAGACGCTCTTCTATCGATAACGTTCAATACGCTAACGTTCCAAACGCTAGTGGTAAACCTAAACGACCGTACTTCGATTACCTCAAGCAGCAGCAAAACCTCTTTGAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACTCCAAAAGTATCACTCAGTAACTCACTACCAACAGAAATGTTGAAAAGGAAATGACGCTAACACGGCATGCCCTGGAATACCAAGGGCGCAATGTGCGTCAAAATTCTGATCGATTCGCTGTTCTCATCGATCGAAAACCAAAATCCGTTGTTGAAAGTTAAATTAAAATTCCGTAGTACGAAAATTCTTGTGTTGAAACGTTGAAACGTTTAATTGCTCTATAACAAAGCACAAAATCTCTACCGTTGGAATACCAAAAAAAACTTACAACCCATTACAGGCCACTAACCGCAGGGCCGGTGGACTCTCCATCTCTGCCCCAGTAAACACTCTCATGCTCTTGCCTAAACAAAAAACTTACATTTCAAAATTATTAATTCTTAAATGATCCTCCGAGGT
EA 1.W NS6 (E16)	<i>Saccharomyces cerevisiae</i>	98%	ITS	CATTGTCGCCAGACGCTCTTCTATCGATAACGTTCAATACGCTAGTAAAAAAAGATTAGCCGAGTGGTAAACCTAAACGACCGTCTTCGATTACCTCAAGCAGCAGCAAAACCTCTTTGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACTCCAAAAGTATCCTCACTACCAACAGAAATGTTGAAAAGGAATGACGCTAACACGGCAAGCCCTGGAATACCAAGGGCGCAATGTGCTCAAAATTCTGATGTTACCGGAATTCTGCAATTACATTACGTTACGTCATTCGCTGCTGTTCTTCATCGATCGGAAAACCAAAATCCGTTGTTGAAGTTTAATTATTTAAAATTCCAGTTACGAAAATTCTGTTGTTGACAAAAATTAAATGAAATAGATAAAATTGTTGGTTGTTACCTCTGGCCCCGATTGCTCGAATGCCAAAAAAAGTTGCAAAGATATGAAAACCTCCACAGTGTGTTGATTGAAACGGTTAAATTGCTCTATAACAAAAGCACAAAATCTCACCGTTGGAATAGCACAAAAAAACTTACACCCCAACAGGCCGCACTTAAACCCAGGCCGGCTGACTCTCCATCTCTGCTCTTGCCTGGACTAAAGCTCTCATGCTCTTGCCTAAACAAAAATCCATTTCAAATTAAATTCTTAAATGATCCTCCGAGGT
EA 1.W NS7 (E17)	<i>Saccharomyces cerevisiae</i>	98%	ITS	AACATTGTCGCCAGACGCTCTTCTATCGATAACGTTCAATACGCTACGATAAAAAAAAGATTACCCGAGTGGTAAACCTAAACGACCGTACATTGCAATTACCTCAAGCAGCAGCAAAACCTCTTTGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACTCCAAAAGTATCACTCACTACCAACAAATGTTGAAAAGGAATGACGCTAACACAGGCATGCCCTGGAATACCAAGGGCGCAATGTGCTTCAAAATTCTGATTCGCGGAATTCTGCAATTACATTACGTTACGTCATTCGCTGCTGCTCTTCATCGATGCAAAACCAAAATCGTTGTTGAAAGTTTTAATAATTAAATTCTCAGTTACAAAATTCTGTTGTTGACAAAATTAAATGAAATAGATAAAATTGTTGGTTGTTACCTCTGGGGCCCGATTGCTCGAATGCCAAAAAAAGTTGCAAACAAATGACGCTAACAGGGTTGTTGAAACCGTTAAATTGCTCTATAACAAAAGCACAAAATCTCACTCGTTGGAATACCAAGAAAGAACCTTACAAGCCTAGCAAGACCGCCACTTAAACCCGAGGCCGGCTGACTCTCCATCTCTGCTCTTGCCTGGACTAAAGCTCTCATGCTCTTGCCTAAACAAAAATCCATTTCAAATTATTAATTCTTAAATGATCCTCCGAGGT

Table A.1. (*continued*)

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EB 1.W NS3 (E23)	<i>Saccharomyces cerevisiae</i>	98%	ITS	ACTTTAAGAACATTGTCGCCTNACGCTCTTCTTATCGATAACGTTCC AATACGCTCAGTATAAAAAAGATTACCGCAGTTGGTAAAACCTAAA ACGACCGTACTTGCATTATACCTCAAGCACGCAAAAAAACCTCTTTG GAAAAAAACATCCAATGAAAGGCCAGCAATTCAAGTTAACCTCAA AGAGTATCACTCACTACCAACAGAATGTTGAAAGGAAATGACGCT CAAACAGGCATGCCCTGGAATACCAAGGGGCAGCAATGTCGTTCAA AGATTGATGATTGACCGGAATTCTGCAATTCACTTACATTACGATTCGATT GCTCGTTCTTCATGCGAGAACAAAAGATCGTTGAAAGTT TTAATATTTAAATTCCAGTTACAAAATTCTGTTTGACAAAAA TTAATGAATAGATAAAATTGTTGTGTTACCTCTGGGCCCCGATT GCTCGAATGCCAAAGGAAAGTGCAGATATGAAAACCTCAAAGCT GTGTTGATTGAAACGGTTTAATTGTCATAACAAAAGCACAGAAAT CTCTCACCGTTGGAATACCAAGAAAAACTTACAAGCTAGCAAGA CCGCGCACTAACCGCAGGCCGCTGACTCTCATCTGTCTTCTT GCCAGTAAACCTCTCATGCTCTGCCAAACAAAAATCCATTTC AAAATTATTAATTCTTAAATGATCCTCCGCAGGT
EB 1.W NS4 (E24)	<i>Saccharomyces cerevisiae</i>	97%	ITS	AACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGC TCAGTATAAAAAAAATTACCGCAGTTGGTAAAACCTAAAACGACCG TACTTGCATTATACCTCAACCACGCAAAAACCTCTTTGGAAAAAA AACATCCAATGAAAGGCCACCAATTCAAGTTAACCTCAAAGTATC ACTCACTACCAACAGAATGTTGAAAGGAAATGACGCTAAACAGG CATGCCCTGGAATACCAAGGGGCCAATGTGCGTCAAAATTGAT GATTACGGAATTCTGCAATTCACTTACGATTCGCAATTGCGTGC TTCATCGATGCGAAAACCAAAAAATCGTTGTTGAAAGTTTAATATT TTAAATTCAGTTACAAAATTCTGTTTGACAAAAATTAAATGAA TAGATAAAATGTTGGGTTGTTACCTCTGGGCCCCGATTGCTCGAATG CCCAAAAAAAAGTTGCAAAATGAAACCTCACAGTGTGTTGATT GAAACGGTTTAATTGTCATAACAAAAGCACAGAAATCTCTCACCGT TTGGAATACCAAAAAAAACTTACAACCCCTACCAAGACGCCGCACTT AACCGCAGGCCGCTGACTCTCATCTGTGTTCTGCCAGTAA ACCTCTATGCTCTGCCAAACAAAAATCCATTTCAAAATTATTAA AATTCTTAAATGATCCTCCGCAGGT
EB 1.W NS6 (E26)	<i>Saccharomyces cerevisiae</i>	96%	ITS	TTGAGGTCAACTTAAAGAACATTGTCGCCTAGACGCTCTTCTTATCG ATAACGTTCCAATACGCTCAGTATAAAAAAAATTACCGCAGTTGTA AAACCTAAACGACCGTACTTGCATTATACCTCAACCACGCAAAAAAA CCTCTTGGAAAAAAACCTCAATGAAAAGGCCACCAATTCAAGT TAACTCCAAAAGTATCCTCACTACCAACAGAATGTTGAAAAGGA ATGACGCTAACACAGGATGCCCTGGAATACCAAGGGGCCAATG TGCCTCAAAATTGATGATTACGGAATTCTGCAATTCACTTACGT ATCGCATTGCGTCTTCATCGATGCGAAAACCAAAAAATCGTT GTTGAAAGTTTAATTTTAAATTCCAGTTACAAAATTCTGTT TTGACAAAAATTAAATGAATAGATAAAATTGTTGGGTTGTTACCTCT GGGCCCCGATTGCTCGAATGCCAAAAAAAGTTGCAAAATATGAA AACTCCACAGTGTGTTATTGAAACGGTTTAATTGTCCTATAACAAA ACCACAAAAATCTCTACCGTTGGAATACCAAAAAAAACTTACAA CCCTACCAAAACGCCCACTAACCGCAGGCCGCTGACTCTCCATC TCTGTTCTGCCAGTAAACCTCTCATGCTCTGCCAAACAAAAAA AATCCATTTCAAAATTATTAATTCTTAAATGATCCTCCGCAGGT
EB 1.W NS7 (E27)	<i>Saccharomyces cerevisiae</i>	97%	ITS	CATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTC AGTATAAAAAAAATTACCGCAGTTGGTAAAACCTAAAACGACCGTA CTTGCATTATACCTCAACCACGCAAAAAAACCTCTTTGGAAAAAA CATCCAATGAAAGGCCAGCAATTCAAGTTAACCTCAAAGTATCAC TCACTACCAACAGAATGTTGAAAGGAAATGACGCTAAACAGGCA TGCCCTGGAATACCAAGGGGCCAATGTGCGTCAAAATTGATG TTCACGGAATTCTGCAATTACGATTCGATTCGCTCGTGC CATCGATGCCAAACAAAAATTCTGTTGAAAGTTTAATATT AAAATTCCAGTTACGAAAATTCTGTTGACAAAAATTAAATGAAT AAATAAAATTGTTGTTGTTACCTCTGGGCCCCGATTGCTCGAATGC CCAAAAAAAGTTGCAAAATGAAACACTCACAGTGTGTTGATT GAAACGGTTTAATTGTCCTATAACAAAAGCACAAAAATCTCACCCT TTGGAATGCAAAAAAAACTTACACCGCTAGCAAGACGCCGCACTT AACCGCAGGCCGCTGACTCTCCATCTGTCTTGTCCCAGTAA AAGCTCTATGCTCTGCCAAACAAAAATCCATTTCAAAATTATTAA AATTCTTAAATGATCCTCCGCAGGTTCACCTACGGGAAGGATC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EB 1.W NS9 (E29)	<i>Saccharomyces cerevisiae</i>	99%	ITS	AACTTTAAGAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTTCCAGTATAAAAAGATTAGCCGCAGTTGGTAAAACCTAAACGACCGTACTTGCAATTACCTCAAGCACGCAGAGAAACCTCTCTTGTGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACACTCCAAAGAGTATCACTCACTACCAACAGAACAGATGTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGGAATACCAAGGGGGCGCAATGTCGTTCAAGAGATCCGTTGAAAATTTAAATTAAAATTTCAGTACGAAAATTCTGCAATTTCACATTACGTTACGATTCGCTGCTTCATTGCTGTTGACAAAAATTAAATTGCTGAACAGAACAGATGTTGTTACCTCTGGGGCCCCGATTGCTGAATGAGCTGAATGCCCAGTAAAGCTCATGCTCTTGCACAAACAAAAAAATCCATTTCAGGTTGAAATAGAAGATAAGGTTGTTAACAGTACGTTGTTGACAAAGATATGAAAACCTCCACAGTGTGTTGATTGAAACGGTTTAATTGTCTATAACAAAAGCACAGAAATCTCTCACCGTTGGAATACCAAGGGGGCGCAATGTCGTTCAAGATTGATGATTCACTACCAAGGGGGCGCTGGACTCTCCATCTTGTCTTGCAGTAAAGCTCTCATGCTCTTGCACAAACAAAAAAATCCATTTCAGGTTACCTAAATTCTTAATGATCCTCCGCAGGTTACCTA
EB 1.W NS10 (E30)	<i>Saccharomyces cerevisiae</i>	98%	ITS	ACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTCAGTATAAAAAGATTACCCGCAGTTGGTAAAACCTAAACGACCGTACTGCATTATACCTCAAGCACGCACAAACCTCTTTGGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACCTCAAACAGACTCACTACCAACAGAACAGATGTTGAAAAGGAATGACGCTCAACACAGCATGCCCTGGAATACCAAGGGGGCGCAATGTCGTTCAAGATTGATGACGGAAATTCTGCAATTTCACATTACGTTACGATTCGCTGCTTCATTGCTGTTGAAAGTTTTAAATTGAAATTCTGTTGTTGTTACCTCTGGGGCCCCGATTGCTCGAATGCCCCAAAAGTTGCAAGAGATATGAAAACCTCCACAGTGTGTTGATTGAAACGGTTTAATTGTCTATAACAAAAGCACAGAAATCTCTCACCGTGAATACCAAGGAAGAACCTACCAAGGCCAGTAAAGCTCTCATGCTCTTGCACAAACAAAAAAATCCATTTCAGGTTACCTAAATTCTTAATGATCCTCCGCAGGTTACCTA
EA 2.W NS1 (E31)	<i>Saccharomyces cerevisiae</i>	98%	ITS	AACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTCAGTATAAAAAGATTACCCGCAGTTGGTAAAACCTAAACGACCGTACTTGCAATTACCTCAAGCACGCACAAACCTCTTTGGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACCTCAAACAGACTCACTACCAACAGAACAGCATGCCCTGGAATACCAAGGGGGCGCAATGTCGTTCAAGATTGATGATTCACTACCAAGGGGGCGCTGGACTCTCCATCTTGTCTTGCAGTAAAGCTCTCATGCTCTTGCACAAACAAAAAAATCCATTTCAGGTTACCTAAATTCTTAATGATCCTCCGCAGGTTACCTA
EA 2.W NS2 (E32)	<i>Saccharomyces cerevisiae</i>	98%	ITS	AACTTTAAGAACATTGTCGCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTTCCAGTATAAAAAGATTACCCGCAGTTGGTAAAACCTAAACGACCGTACTTGCAATTACCTCAAGCACGCACAAACCTCTCTTGTGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACACTCCAAAAGTATCACTCACTACCAACAGAACAGATGTTGAAAAGGAATGACGCTCAAACAGGCATGCCCTGGAATACCAAGGGGGCGCAATGTCGTTCAATTACGTTACGATTCGCTGCTTCATTGCTGTTGACAAATTTAAATTAAAATTTCAGTACGAAAATTCTGTTGACAAAAATTAAATGCAATTGCTGAATGCCCACAAAAAAAGTTGCAAGAGATATGAAAACCTCCACAGTGTGTTGATTGAAACGGTTTAATTGTCTATAACAAAAGCACAGAAATCTCTCACCGTTGGAATACCAAGGAAGAACCTACCAAGGCCAGTAAAGCTCTCATGCTCTTGCACAAACAAAAAAATCCATTTCAGGTTACCTAAATTCTTAATGATCCTCCGCAGGTTACCTA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EA 0.W NS6 (E35)	<i>Saccharomyces cerevisiae</i>	91%	ITS	TTGAGGTCAACTTAAGAACATTGTTGCCAGACGCTCTTCTTATCGATAAACGTTCCAATACGCTCAGTATAAAAAAAAATTACCGCATTGGTA AAACCTAAACAAACCGTACTTCGATTACCTCAGGCACGCACGGAA CCTCTTTGAAAAAAACATCCAAGGAAAAGGCCAGCAATTCAAGTTAACCAAAAAGTATCACTCACTACCAACAAAAGGTTGAAAAGGAAAGGACGCTCAA AAAGGACGCTCAAACAGGCAGGCCCGGAATACCAAGGGCGCATG GTGCGTTCAAACATTCAATGATTCACCGGAATTCTGCAATTCAATTACG TATCGCATTCCCTCGCTTCTCATCAATCGCAAAAACCCAAAAATTCC GTTGTTGAAAGTTTTAATATTAAAATTCCATTACAAAATTCTGG TTTTGACAAAAATTAAAGGAATAAAATAAAATTGTTGGGGTTGTTACC TCTGGGCCAATTGCTCAAGGCCAA GAAAACCTCACATTGGTTGAATTGAAACGGTTTAATTGTC AAACCCACCAGAACGCCACTTAACCCCAGGCCGCTGGACTCCCC ATCCCTGGCTCTTGCCTTAAACACCTTCAGGCCTCTGCA CCCCAAAAATCCC
EA 0.W NS7 (E36)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TGAGAGCTTTACTGGCAAGAACAGACAAGAGATGGAGAGTCCAGCCGG GCCTGCGCTTAAGTGCAGGGCTTGCTAGGCTGTAAGTTCTTCTGC TATTCCAAACCGTGAGAGATTCTGTGCTTTGTTAGGACAATTAAA ACCGTTCAATACAACACACTGTGGAGTTTCATATTTGCAACTTTT CTTGGGCTTCAGCAATCGGGGCCAGAGGTAACAAACACAAACAA TTTATATTCAATTAACTTGTCAAACAGAAATTTCGTAACTGG AAATTAAAATTAAAACATTCAACACGGATCTTGGTCTCG ATCGATGAAGAACGCAGCGAAATGCGATACGTAAATGTGAATTGAGAA TCCGTAATCATCGAATCTTGAACGCACATTGCGCCCTGGTATTCC AGGGGCATGGCTTTGAGCGTCAATTCTCAACATTCTGTTGG TAGTGAATGACTCTTGGAGTTAACTGAAATTGCTGGCTTCTTATT GGATGTTTATTCCAAGAGAGGTTCTGCGTGTGAGGTATAAT GCAAGTACGGCTTTAGGTTTACCAACTGCGGCTAATTTTTTAT ACTGAGCGTATTGGAACGTTATCGATAAGAACAGAGCGTCTAGGC CAATGTTAAAGTTGACCTCAAATCAGTAGGAGTACCCGCTGA TTAAGCATATCAAT
EA 0.W NS8 (E37)	<i>Wickerhamomyces anomalus</i>	100%	ITS	CGCTTATTGCGCGGCATAAACCTTACACACATTGCTAGTTTTGAA CTTGCTTGGTGGTGAACCTGGCTACTGCCAACGGCTAAACACA TTTTTTAATGTTAAACCTTAAACCAATAGTCATGAAAATTAA AAAATTAAAATCTTCAAACACTTCAACACGGATCTTGGTCTCG ACGATGAAGAACGCAGCGAAATGCGATACGTATTGTAATTGAGATT TCGTAATCATCGAATCTTGAACGCACATTGACCCCTGGTATTCA GAGGGTATGCCCTGGTGAACGGCTATTCTCTCAACCTCGGGTTGG TATTGAGTGAATCTGCAAGGGTTAACTGAAATTGACTTAGCAA GAGTGAATGACTCTGCAAGGGTTAACTGAAATTGACTTAGCAA ACTCGTTATACAGCTAGGCAGGTTAGAAGTATTAGGCTTCCA ACAACAATAAAACTAAAGTTGACCTCAAATCAGTAGGAGTACCCGCT GAACCTAACGATATCAATA
EA 0.W NS9 (E38)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTGTTGGCAAGAGCATGAGAGCTTTACTGGCAAGA AGACAAGAGATGGAGAGTCCAGCCGGCCTGCCTTAAGTGCAGGGTC TTGCTAGGCTGTAAGTTCTTCTGCTATTCCAAACGGTGAGAGATT CTGTGCTTTGTTAGGACAATTAAAACGGTTCAATACAACACACTG TGGAGTTTCAATCTGCAACTTTCTTGGCATTGAGCAATCG GGCCAGAGGTAACAAACACAAACATTATCTATTCAATTAA GTCAAAAACAAGAATTTCGTAACTGGAAATTAAAATATTAAA TTCAACAAACGGATCTTGGTCTCGCATCGATGAAGAACGCAGCG TGCATACGTAATGTAATTGAGCAATTCTGAGTGAATTGACTCTGGAG AACGCACATTGCGCCCTGGTATTCCAGGGCATGCCTGTTGAGCG TCATTCTCTCAACATCTGTTGAGTGAATTGACTCTGGAG TTAACCTGAAATTGCTGGCTTTTATTGGATGTTTTTCCA GGTTTCTCGCTGTTGAGGTATAATGCAAGTACGGCTGTTAGGTT TACCAACTGCGGCTAATCTTTTATACTGAGCGTATTGAAACGTT GATAAAAAAAAAGCGTCTAGGCAAACAAATGTTAAAGTTGACCTCA AATCAGGTAGGAGTACCCGCTGAACCTAACCATATCATAAA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EA 0.W NS10 (E39)	<i>Saccharomyces cerevisiae</i>	99%	ITS	AACTTTAAGAACATTGTCGCCTAGACGCTCTCTTATCGATAACGTTCCAATACGCTCAGTATAAAAAAGATTAGCCGCAGTTGGTAAAACCTAACAGACCGTACTTGCAATTACCTCAAGCACGCAGAGAAACCTCTTCTGGAATAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTAACTCCAAGAGTATCACTCACTACCAACAGAACAGATGTTGAGAAGGAAATGACGCTCAACACGGCATGCCCTGGAATACCAAGGGGCCAATGCGTTCAAAAGATTCAACTACGTTACAGTATCGCATTCAGATTGATGTCAGGAATTCTGCAATTTCACATTACGATTCGCTGCTCTTCATCGATGCGAGAACCAAAGAGATCCGTTGAAA GTTTTAATATTTAAAATTCCAGTTACGAAAATTCTGTTTGACAAAAATTAATGAAAGATAAAATTGTTGTGTTGACCTGGGGCCCCGATTGCTGAATGCCAAAGAAAAGTTGACATGAAAAGATATGAAAACCTCACAGTTGTTGATGAAACGGTTAATTGCTCTATAACAAAAGCACAGAAATCTCACCGTTGGAATAGCAAGAAAGAAAATCAAGCCTAGCAAGCCGCACACTTAAGCGCAGGCCGCTGACTCTCCATCTTGCTCTTGCCAGTAAAGCTCATGCTCTGCAAACAAAAAAATCCATTCAAATTAAATTCTTAAATGATCCTTCGC
EB 0.W NS6 (E40)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTTGTTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAGACAAGAGATGGAGAGTCCAGCCGGCCTGCCTTAAGTGCAGCGTC TTGCTAGGCTGTAAGTTCTTCTGCTATTCAAACCGGTGAGAGATTCTGTGTTTGTATAGGACAATTAAACCGTTCAATACAACACTG TGGAGTTTCATATCTTGCACCTTTCTTGGCATTCGAATCGGCGGCCAGAGTAACAAACACAACAAACATTTATCTATTCAATTAAATTTGTCAAAACAGAATTTCGTACTGGAATTAAATTTAAATATTAAAAAAACTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATGGATACGTAATGTGAATTGCAAGAACATCCGTGAATCATCGAATCTTG AACGCACATTGCGCCCTGGTATTCCAGGGGCATGCCGTGTTGAGCGTCATTCTCTCAAACATTCTGTTGGTAGTGAGTGATACTCTTGGAGTTAACCTGAAATTGCTGCCCTTCATTGGATGTTTTTCAAAGAGAGGTTCTCGGTGCTTGAGGTATAATGCAAGTACGGTGTGTTAGGTTTACCAACTGCCGCTAATCTTTTATACTGAGCGTATTGAAACGTTATCGATAAAAGCGTCTAGGCAAACAAATGTTCAAAGTTGACCTAAATCAGGTAGGAGTACCCGCTGAACCTAACAGCATATCAAAGCGAAGGA
EB 0.W NS7 (E41)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTTGTTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAGACAAGAGATGGAGAGTCCAGCCGGCCTGCCTTAAGTGCAGCGTC TTGCTAGGCTGTAAGTTCTTCTGCTATTCAAACCGGTGAGAGATTCTGTGTTTGTATAGGACAATTAAACCGTTCAATACAACACTG TGGAGTTTCATATCTTGCACCTTTCTTGGCATTCGAATCGGCGGCCAGAGTAACAAACACAACAAATTATCTATTCAATTAAATTTGTCAAAACAGAATTTCGTACTGGAATTAAATTTAAATATTAAAAAAACTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATGGATACGTAATGTGAATTGCAAGAACATCCGTGAATCATCGAATCTTG AACGCACATTGCGCCCTGGTATTCCAGGGGCATGCCGTGTTGAGCGTCATTCTCTCAAACATTCTGTTGGTAGTGAGTGATACTCTTGGAGTTAACCTGAAATTGCTGCCCTTCATTGGATGTTTTTCAAAGAGAGGTTCTCGGTGCTTGAGGTATAATGCAAGTACGGTGTGTTAGGTTTACCAACTGCCGCTAATCTTTTATACTGAGCGTATTGAAACGTTATCGATAAAAGCGTCTAGGCAAACAAATGTTCAAAGTTGACCTAAATCAGGTAGGAGTACCCGCTGAACCTAACAGCATATCAA
EB 0.W NS8 (E42)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTTGTTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAGACAAGAGATGGAGAGTCCAGCCGGCCTGCCTTAAGTGCAGCGTC TTGCTAGGCTGTAAGTTCTTCTGCTATTCAAACCGGTGAGAGATTCTGTGTTTGTATAGGACAATTAAACCGTTCAATACAACACTG TGGAGTTTCATATCTTGCACCTTTCTTGGCATTCGAATCGGCGGCCAGAGTAACAAACACAACAAATTATCTATTCAATTAAATTTGTCAAAACAGAATTTCGTACTGGAATTAAATTTAAATATTAAAAAAACTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATGGATACGTAATGTGAATTGCAAGAACATCCGTGAATCATCGAATCTTG AACGCACATTGCGCCCTGGTATTCCAGGGGCATGCCGTGTTGAGCGTCATTCTCTCAAACATTCTGTTGGTAGTGAGTGATACTCTTGGAGTTAACCTGAAATTGCTGCCCTTCATTGGATGTTTTTCAAAGAGAGGTTCTCGGTGCTTGAGGTATAATGCAAGTACGGTGTGTTAGGTTTACCAACTGCCGCTAATCTTTTATACTGAGCGTATTGAAACGTTATCGATAAA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
EB 0.W NS9 (E43)	<i>Saccharomyces cerevisiae</i>	98%	ITS	ACTTTAAGAACATTGTTGCCCTAGACGCTCTTCTTATCGATAACGTTCCAATACGCTCAGTATAAAAAAAGATTAGCCGCAGTGTGAAACCTAA AACGACCGTACTTGCATTATACCTCAAGCACGCAAAGAAACCTCTTT CGAAAAAAAACATCCAATGAAAAGGCAGCAATTCAAGTAACTCCA AAGAGTATCACTCACTACCAACAAAGGTTGAAAAGGAATGACGC TCAAAACAGGATGCCCGGGAAATACCAAGGGGCGCAAGGGGCCGTTCA AGAGTCGATGATTCAACGGAAATTCTGCAATTACATACGTTACGCTCATT TCCCTCGCTTCTCATCGATGCGAGAACCAAAAATCCGTTGTGAAAG TTTTAATATTTAAAATTCCAGTTACAAAATCTGTTTGACAAA ATTTAATGAATAAAATTTAAATTGTTGTGTTACCTCTGGGCCCGA TTGCTGAATGCCAAAAAAAGGAAATGCAAAAATATGAAACTCCACAA GTGTGTTGATTGAAACGTTTAATTGCTCTATAACAAAAGCACAAA ATCTCTACCGTTGGAATAGCAAGAAAGAAACTACAAGCTAGCAA GACCGCGCACTTAAGCGCAGGCCGCTGGACTCTCCATCTGTCTT CTTGCCAGTAAAGCTCATGCTCTGCCAAACAAAAAATCCATT TTCAAAATTATTAATTTCAATGATCCTTCC
EB 0.W NS10 (E44)	<i>Saccharomyces cerevisiae</i>	99%	ITS	GCTCTCTTCTTATCGATAACGTTNAATACGCTCAGTATAAAAAAAAT TACCGAAGTGGTAAAACCTAAAACGACCGCTCGCTTATCCCTAA GGCCCCCAAAAAACCTTTTTGGAAAAAAAACATCCAAGGAAAGG CCAGCATTAAAGTTAACTCCAAAAGTTAACTNNTTCCAAACCAAA AGGTTGAAAAGGAAAGGCTTAAACCCAGGCTGCCCGGGAAATCC CAAGGGCGCATGGGCGTTCAAGGATTNCAGGATAACGGAATTG CATTNNACATCCGTATNGAATTNTCTGNNTTTAATNGAGGCAAAA CCCAAAAATCCGGTGGAAAGTTTAATTTAAAATTTCAAGT CCAAAATTTGGTTTGCACAAATTAAAGGAATAATAAATGGT GGGGTTGGTACCTNTGGCCCNATGGTCTNAAGGCCAAAAAAAGG GTGGCAANANTGAAACCNACAGGGGTGGNTTGGAACCGGTTT AATGGNCNNNTNNNANAAGCNAAAATTAACTCGTGGGATTA
EB 3.W S5 (E45)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGGCAAAGCTCAAATTGAAATCTGGTACCTCGGTGCCCGAGTTGAA TTTGGAGAGGGCAACTTGGGCGTCTTGTCTATGTCCTTGAAC AGGACGTATAGAGGTGAGAATCCCGTGTGGCGAGGAGTCGCGTTCT TTGTAAGTGCCTCGAAGAGTCGAGTTGGGAATGCAGCTAAG TGGGTGGTAATTCATCTAAAGCTAAATTTGGCGAGAGACCGATAGC GAACAAAGTACAGTGTGGAAAGATGAAAGAAACTTGAAGAGAGTG AAAAGTACGTGAAATTGTTGAAGGGAGGGCATTGATCAGACATG GTGTTTGTGCCCTGCTCTGTGGTAGGGGAATCTCGCATTTCACT GGGCCAGCATCAGTTGGTGCAGGATAATCCATAGGAATGAGCTT GCCTCGTAAGTATTATAGCTGTGGGAATACTGCCAGCTGGACTGAG GACTGCGACGTAAGTCAGGATGCTGGCATAATGGTTATATGCCCGCCG TCTTGAAA
OA 0.CM NS1 (O1)	<i>Lachancea thermotolerans</i>	99%	D1/D2 Domain	CGCGGAGTGAAGCGGAAAGCTCAAATTGAAATCTGGCACCTCGG TGTCGAGTTGAATTGAAAGAAGCTACTTGGGCTAGTCCTGTCTAT GTTCCCTGGACAGGACGTATGGAGGGTGAGAATCCGTATGGCAG GAGTCTAGTCTATGTAAGTGTCTTCGACGAGTCGAGTTGGAA TGCAGCTCAAGTGGGTGTAATTCATCTAAAGCTAAATTTGGCGA GAGACCGATAAGCAACATACAGTGTGGAAAGATGAAAGAAACTT GAAAAGAGAGTGGAAAGTACGTGAAATTGTTGAAGGGAGGGCATT TGATCAGACATGGTGTGCAACATCAGTTGGCGTAGGATAATCTTGG CGCAGCTCACTGGGCAACATCAGTTGGCGTAGGATAATCTTGG GAATGTGGCTCTCGAAAAGCTTATAGCCCAGGGGAATACTACC AGCCGGGACTGAGGACTGCGACTTTGTCAGGATGTTGCATAATGGT TAAATGCCGCCGTCTT
		99%	ITS	GTCAACTTATGAATAACTGTTGCCAGACTGCTGACTTCGTAAGC CTTAAAACGCCACTAACGCCACTACGAGTTGAAACCTAAACGC AGAGTATCAGCGACTGGACTAAACCTCAGTCAGAACAGCCAGATG GCCAGCATTTCAAGTTAACCCAGAAAGAGTACCACTCACTACCAACC CGAGGGTTGAGAAGGAAATGACGCTCAAACAGGCATGCCCTGGAA TACCAAGGGCGCAATGCGTTCAAAGATTGATGATTCAAGTACGAG AACCAAGAGATCGTTGAAAGTTAAATATTTCTAAATGA AAAATAATTGACAATGTTAAATACAATAATGTTGTGTTGAAAC CTTGGGCCGTAAGGCCAAAGAAGAAGCGTTGAAATAAAATTACTCC ACAGTGTGTGTTAGAGAGACAGTCGTTAACAGAAAGCATCATCGGCC GCGCAATCAAGCGCAGGCCCTGCAACTTCCGGCTGCTCAACAAAAT TCTTAATGATCCTCCGCAGGTTCA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OB 0.CM NS2 (O2)	<i>Lachancea thermotolerans</i>	99%	ITS	ACTTTATGAATACTGTTGCCAGACTGTCTGTACTTCAGTAAAAGCCTT TAAAACGCCTACTAACGCCACTACGAGTTGGTAAAACCTAATACGCAG AGTATCAGCGGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGGC CAGCATTTCAAGTTAACCCAGAAAGAGTACCACTACTACCAAACCCG AGGGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGAATA CCAGAGGGCGCAATGTGCTTAAAGATTGATCGATGATTCAAGATATCTG CAATTCAACAATCTTATCGCATTTCGCTGCGTTCTCATCGATGCGAGAA CCAAGAGATCCGTTGAAAGTTAAATATTTCCTAAATGAAA AATAATTGACAATGTTAAATACAATAATTGTTGTGTTGTAACCCTT GGGCGTAAAGCCAAAGAAGAAGCCTGAAAATAATTACTCCACA GTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCATCATGGCCGCG CAATCAAGCGCAGGCTCTGAACATTCCCGCTGCTAACAAAATTCT TTAATGATCCTCCGCAGGTTCC
OB 0.CM NS3 (O3)	<i>Lachancea thermotolerans</i>	99%	ITS	AACTTTATGAATACTGTTGCCAGACTGTCTGTACTTCAGTAAAAGCCTT TAAAACGCCTACTAACGCCACTACGAGTTGGTAAAACCTAATACGCAG AGTATCAGCGGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGGC CAGCATTTCAAGTTAACCCAGAAAGAGTACCACTACTACCAAACCCG AGGGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGAATA CCAGAGGGCGCAATGTGCTTAAAGATTGATCGATGATTCAAGATATCTG CAATTCAACAATCTTATCGCATTTCGCTGCGTTCTCATCGATGCGAGAA CCAAGAGATCCGTTGAAAGTTAAATATTTCCTAAATGAAA AATAATTGACAATGTTAAATACAATAATTGTTGTGTTGTAACCCTT GGGCGTAAAGCCAAAGAAGAAGCCTGAAAATAATTACTCCACA GTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCATCATGGCCGCG CAATCAAGCGCAGGCTCTGAACATTCCCGCTGCTAACAAAATTCT TTAATGATCCTCCGCAGGTTCC
OB 0.CM NS5 (O4)	<i>Lachancea thermotolerans</i>	99%	ITS	AACTTTATGAATACTGTTGCCAGACTGTCTGTACTTCAGTAAAAGCCTT TAAAACGCCTACTAACGCCACTACGAGTTGGTAAAACCTAATACGCAG AGTATCAGCGGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGGC CAGCATTTCAAGTTAACCCAGAAAGAGTACCACTACTACCAAACCCG AGGGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGAATA CCAGAGGGCGCAATGTGCTTAAAGATTGATCGATGATTCAAGATATCTG CAATTCAACAATCTTATCGCATTTCGCTGCGTTCTCATCGATGCGAGAA CCAAGAGATCCGTTGAAAGTTAAATATTTCCTAAATGAAA AATAATTGACAATGTTAAATACAATAATTGTTGTGTTGTAACCCTT GGGCGTAAAGCCAAAGAAGAAGCCTGAAAATAATTACTCCACA GTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCATCATGGCCGCG CAATCAAGCGCAGGCTCTGAACATTCCCGCTGCTAACAAAATTCT TTAATGATCCTCCGCAGGTTCC
OA 4.CM NS2 (O5)	<i>Hanseniaspora opuntiae</i>	100%	ITS	CCCTTGCCTAAGGTACATTACCATTTCTGTAAAGTAAAAGAATA AATCCATAAATACATCACAGCGAGAACAGCGTCTCCAAGAAGCTAAG TGTGAAATTAAAAAGACTGAAACAGTCTCCAATTCAAGCTAACCTG AGTATGCCCAACACAAAAAAATTATCTTTGAGAAGGAAA TGACGCTCAACAGGCATGCCCTTGAGAATGCTCAAGGGCGCAATGTG CGTTAAAAATCAATGATTGACGATCTGCAATTACATTACTATC GCAATTGCTACGTTCTCATCGATGCGAGAACCAAGAGATCCGTTGT GAAAGTTAAATATTAAATTCGTTAGGAATTGGTTAGTT AAAAATTATAATAAAAATTGTTGTGTTGTTGCTTGAAC CTTCGATTCAAAGCAGAAAGAATTAAATTAAGTAAAAACTCCAA TGTGTAAACGTTGACTGAGATTCAAGCAAGACTACTTCACTGCGAC ACTCTAATGAAGCAGCGCAATTAGCCACATCTCAATAAGATAACAC ATTATTGAAAGATCTAACAAAGACTCGAGCAACAATGATAATTCA ATCTAATGATCCTCCGCAGGTT
OA 4.CM NS3 (O6)	<i>Lachancea thermotolerans</i>	99%	ITS	CTTTATGAATACTGTTGCCAGACTGTCTGTACTTCAGTAAAAGCCTTA AAACGCCTACTAACGCCACTACGAGTTGGTAAAACCTAATACGCAGAG TATCAGCGGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGCCA GCATTTCAAGTTAACCCAGAAAGAGTACCACTACTACCAAACCCGAG GGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGAATACC AGAGGGCGCAATGTGCGTCAAAGATTGATGATTCAAGAATATCTGCA ATTCAACAATCTTATCGCATTTCGCTGCGTTCTCATCGATGCGAGAAC AAGAGATCCGTTGAAAGTTAAATATTTCCTAAATGAAA ATAATTGACAATGTTAAATACAATAATTGTTGTGTTGTAACCCTG GGGCGTAAAGCCAAAGAAGAAGCCTGAAAATAATTACTCCACAG TGTGTGTAGAGAGACAGTCGTTAACAGAAAGCATCATGGCCGCGCA AATCAAGCGCAGGCTCTGAACATTCCCGCTGCTAACAAAATTCT TTAATGATCCTCCGCAGGTT

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OA 4.CM NS4 (O7)	<i>Hanseniaspora opuntiae</i>	100%	D1/D2 Domain	CGGCAGTGAAGCGTAAAGCTCAAATTGAAATCTGGTACTTCAGT CCCCAGTTGTAAATTGAAATTGCTTGGATTAAGTCCTTGCTATG TTCCTTGGAACAGGACGTATAGAGGGTGAGAATCCGTTGGCAGG ATACCTTTCTCTGTAAAGACTTTCAAGAGTCAGTTGGAGAATG CAGCTCAAAGTGGGTGAAATTCCATCTAAAGCTAAATATTGGCAGA GACCAGATAGCAACAAGTACAGTGATGAAAGATGAAAAGAACTTG AAAGAGAGTGAAGAAAGTACGTGAAATTGTTGAAGGGAAAGGGCATTG ATCAGACATGGTGTGTTGCATGCACTGCCCTCGTGGCTTGGCCT CTCAAAAATTCACTGGCCAACATCAATTCTGGCAGTAGGATAAATCA TTAAGAATGTAGTACCTCGTAGTGTATAGTTATTGAAACTGCT AGCTGGGATTGAGGACTGCGCTCGCAAGGATGTTGCATAATGGTA ATGCCGCCCTTGA
	<i>Hanseniaspora opuntiae</i>	100%	ITS	AACTTGATGAATATTAAAAGCAACCCCTTGCTTAAGGTACATTACATT TCCCTGTAAAGTAAAGAACGAAATAATCCATAAATACATCACAGCGAGA ACAGCGTCTCAAAGAACGTAAGTGTGAAATTAAAAAGACTGAAACA GTCTCAAATTCAAGCTAACCTGAGTATGCCAACACAGGCATGCCCTGA GAATGCTCAAGGCCAATGTGCGTCAAAATTCAATGATTACAG ATCTGCAATTCACTTACTATCGCAATTGCTACGTTCTCATCGATGC GAGAACCAAGAGATCCGTTGAAAGTTAAATTATTAAAATTTC CGTTAGGAATTGGTTAGTTAAAAATTATAATAAAAATAATTGT TTGTGTTGTTGCCTGCAACCTTGATTCAAAGCAGAAAAGAATTA AATTAAAGTAAAAAACTCCAATGTTGAAACGTTGACTGAGATTCA CAAGACTACTTACTCGCACACTTAATGAAGCAGCGCAATTAAAGC ACATCTCAAAGATAACATTATTGTTAAAGATCTAAACAAGAACT CGAGCAACAATGATAATTCAATGATCCTCCGAGGT
OB 4.CM NS3 (O8)	<i>Lachancea thermotolerans</i>	99%	ITS	AACTTATGAATACTGTTGCCACTGCTGTACTTCGAAAGCCTT AAAACGCCACTAACGCCACTACGAGTTGTAACCTAATACGAGA GTATCAGCGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGCC AGCATTTCAGTTAACCCAGAAAGAGTACCACTACCTACCAAAACCGA GGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGATAC CAGAGGGCGCAATGTGCGTCAAAGATTGATGATTACGAATATCTGC ATTACAATACTTATCGCATTGCTCGTTCTCATCGATGCGAGAAC CAAGAGATCCGTTGTTGAAAGTTAAATTTTTCTAAATGAAA ATAATTGACAATGTTAAATAATAATTGTTGTTGTAACCCCT GGCGCTAAAGCCAAAGAAGAAGCGTTGAAAATAATTACTCCACA GTGTGTTGAGAGAGACAGTCGTTAACAGAAAGCATCATGCCCG CAATCAAGCGCAGGCTCTGAACCTCCCGCTGCTTAACAAAATTCT TTAATGATGCTCCGAGGTT
OB 4.CM NS4 (O9)	<i>Lachancea thermotolerans</i>	99%	ITS	CTTATGAATACTGTTGCCACTGCTGTACTTCGATAAGCCTT AAACGCCACTAACGCCACTACGAGTTGTAACCTAATACGAGA TATCAGCGACTGGACTAAAACCTCAGTCAGCAACAGCCAGATGCC GCATTTCAGTTAACCCAGAAAGAGTACCACTACCTACCAAAACCGAG GGTTTGAGAAGGAATGACGCTAACACAGGCATGCCCTGGATAC AGAGGGCGCAATGTGCGTCAAAGATTGATGATTACGAATATCTGC ATTACAATACTTATCGCATTGCTCGTTCTCATCGATGCGAGAAC AAGAGATCCGTTGTTGAAAGTTAAATTTTTCTAAATGAAA ATAATTGACAATGTTAAATAATAATTGTTGTTGTTGTAACCCCT GGCGCTAAAGCCAAAGAAGAAGCGTTGAAAATAATTACTCCACAG TGTGTTGAGAGAGACAGTCGTTAACAGAAAGCATCATGCCCG AATCAAGCGCAGGCTCTGAACCTCCCGCTGCTTAACAAAATTCT TAATGATCCTCCGAGGTT
OB 4.CM NS5 (O10)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TTGAGGTCACTTAAAGAACATTGTCGCCAGACGCTCTTCTTATCG ATAACGTTCCAATACGCTCAGTATAAAAAAGATTAGCCGAGTTGTA AAACCTAAACGACCGTACTGCAATTACCTCAAGCAGCAGAGAAA CCTCTTTGAAATAAAACATCCAATGAAAGGCCAGCAATTCAAGT TAACTCCAAGAGTACTCACTACCAAAACAGAAATTGAGAAGGA AATGACGCTAACACAGGCATGCCCTGGATACCAAGGGCGCAATG TGCCTCAAGATTGATTCACGGAAATTGCAATTACATTACGT ATCGCATTGCTCGTTCTCATCGATGCGAGAACAGAGATCGTT GTTGAAAGTTAAATTAAATTCCAGTTACGAAAATTCTGTT TTGACAAAATTAAATGAATGATAAAATTGTTGTTGTTACCTCTG GGCCCGATTGCTGAATGCCAAAGAAAAAGTTGCAAAGATATGAAA ACTCCACAGTGTGTTGATGAAACGTTTAAATTGCTCTATAACAAAAA GCACAGAAATCTCACCGCTTGGATAGCAAGAAAGAAACTACAAG CCTAGCAAGACCGCGCACTTAAAGCGCAGGCCGGCTGGACTCTCATCT CTTGCTCTGCCAGTAAAGCTCTCATGCTCTGCCAAAACAAAAAA AATCCATTCAAAATTAAATTCTTAAATGATCCTCCGCA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OA 0.CM NS2 (O11)	<i>Lachancea thermotolerans</i>	99%	ITS	GGAAGTTCAGAAGCCTGCCTTGATTGCGCGGCCGATGATGCTTCTGT TAACGACTGTCTCTACACACACTGTGGAGTAATTATTTACAAC GCTTCTTCTTGGGCTTACGGCCCAAGGGTACAACACAAACAACTTA TTGTATTTAACATTGCAATTATTTCATTTAGAAAAAAAATTT AAAACTTCAACACAGGATCTCTGATTCTCGCATCGATGAAGAACGCA GCGAAATGCAGATAAGTATGTGAATTTCAGAGATATTCTGTAATCATGAA TCTTGAGCCACATTGCCCTCTGGTATTCAGGGGGCATGCCCTGTT GAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAATGACTCT TTCTGGGTTAACTTGAAGATGCTGCCATCTGCTGTTGACTGAGG TTTAGTCCAGTCGCTGATACTCTGCGTATTAGGTTTACCAACTCGTA GTGGCGTTAGTAGGCCTTAAAGGTTTACTGAAAGTACAG
OA 0.CM NS3 (O12)	<i>Saccharomyces cerevisiae</i>	100%	ITS	TTTTGAAAATGGATTTTTGGCAAGAGCATGAGAGCTTTACTG GGCAAGAACAGAACAGAGATGGAGAGTCCAGCCGGCTGCGCTTAAGTG CGCGGTCTTGTAGGCTTGAAAGTTCTTCTGCTATTCAAACCGTGA GAGATTCTGTGCTTGTATAGGACAATTAAACCGTTCAATACAA CACACTGTGGAGTTTCATATCTTGCAACTTTTCTTGGCATTGAG CAATCGGGGCCAGAGGTAAACACAAACAAACAAATTATCTATTCA AATTGGTCAAAAACAAGAAATTCTGAACTGGAAATTAAACATT AAAACCTTCACAAACGGATCTGGTCTCGCATCGATGAAGAACG AGCGAAATGCAGATACTGAATTGCAAGATTCCGTGAATCATGAA ATCTTGAAACGACATTGCCCTTGTATTCAGGGGGCATGCCCTG TTGAGCGTCATTCTCTCAACATTCTGTTGGTAGTGAATGACTC TTGGAGTTAACTTGAAGATTCTGCTGCTGAGGTATAATGCAAGTACGGCGT TTAGGTTTACCAACTGCGCTAATCTTTTATACTGAGCGTATGGA ACGTTATGATAAGAAGAGAGCGCTAGCGAACATGTTAAAGTT TGACCTAAATCAGGTAGGAGTACCCGCTGAACCTAAAGCATATCAA
OA 0.CM NS4 (O13)	<i>Lachancea thermotolerans</i>	99%	ITS	GCCGGGAAGTTCAGAAGCCTGCCTTGATTGCGCGGCCGATGATGCTT CTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTTAC AACGCTTCTTGGGCTTACGGCCAAGGGTACAACACAAACAA TTATTGTATTAAACATTGCAATTATTTCATTTAGAAAAAAAATA TTAAAACCTTCACAAACCGGATCTGGTCTCGCATCGATGAAGAAC GCAGCGAAATGCAGATAAGTATTGCAATTGAGATATTCTGGAATCATC GAATCTTGAGCGCACATTGCCCTCTGGTATTCAGGGGGCATGCC GTTGAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAAGTGGTA CTCTTCTGGTTAACTTGAAGATGCTGCCATCTGGCTGTTGACTG AGGTTTAGCCAGTCCGCTGATACTCTGCTATTAGGTTTACCAACTC GTAGTGGCGTTAGTAGGCCTTAAAGGTTTACTGAAAGTACAGACA GTCTGGCAAACAGTATTCAAAAGTTGACCTAAATCAGGTAGGATTA CCGCTGAACCTAAAGCATATCAAATA
OA 0.CM NS5 (O14)	<i>Lachancea thermotolerans</i>	99%	ITS	GAGGTCAACTTTATGAATACTGTTGCCAGACTGTCTGACTTCA AAGCCTTAAACGCCACTAACGCCACTACGAGTTGGAAAACCTAA ACCGAGAGTATCAGCGACTGGACTAAACCTCAGTCAGCACAGCCA GATGCCAGCATTTCAAGTTAACCCAGAAAGAGTACCAACTCACCA AACCGAGGGTTGAGAAGGAATGACGCTAACACAGGCATGCC GGAATACCAAGGGCGCAATGTGCGTCAAAGATTGCGATTCACGA ATATCTGCAATTCAAATACTTATCGCATTGCTGCGTCTTCATCGAT GCGAGAACAGAGATCCGTTGTGAAGTTAAATATTTTCTA AAATGAAAAAAATGACAATGTTAAATACAATAATTGTTGT GTAACCCCTGGGCCGTAAGGCCAAAGAAGAAGCGTTGAAATTA TACTCCACAGTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCAT CGGCCGCGCAATCAAGCCAGGCTCTGAACTTCCGGCTGCTCAAC AAAATTCTTAATGATCCTCCGAGGT
OA 0.CM NS6 (O15)	<i>Lachancea thermotolerans</i>	99%	ITS	ATTGAGGTCAACTTTATGAATACTGTTGCCAGACTGTCTGACTTCA GTAAGGCTTAAACGCCACTAACGCCACTACGAGTTGGAAAACC TAATACGCAAGTATCAGCGACTGGACTAAACCTCAGTCAGCAACA GCCAGATGCCAGCATTTCAAGTTAACCCAGAAAGAGTACCAACT ACCAAACCGAGGGTTGAGAAGGAATGACGCTAACACAGGCATGCC CCCTGGAAATACCAAGAGGGCGCAATGTGCGTCAAAGATTGCGATGATTCA CGAATATCTGCAATTCAAATACTTATCGCATTGCTGCGTCTTCATC GATGCGAGAACCAAGAGATCGTTGTGAAGTTAAATATTTT CTAAAATGAAAATAATTGACAATGTTAAATACAATAATTGTTGT TTGTAACCCCTGGGCCGTAAGGCCAAAGAAGAAGCGTTGAAATA AATTACTCCACAGTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCAT CATGCCGCGCAATCAAGCCAGGCTCTGAACTTCCGGCTGCTCAAC AAAATTCTTAATGATCCTCCGGG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OA 0.CM NS7 (O16)	<i>Lachancea thermotolerans</i>	99%	ITS	CCGGGGAAAGTTCAAGAACGCTGCCTGATTGCGGCCGATGATGCTT CTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTTAC AACGCTTCTCTTGGGCTTACGGCCAAGGGTTACAAACACAAACAA TTATTGTATTTAACATTGTCAATTATTTCATTTAGAAAAAAAATA TTAAAACCTTCACAAACGGATCTCTGGTTCTCGCATCGATGAAGAAC GCAGCGAAATCGGATAAGTATTGTGAATTGCAGATATTGCTGAATCATC GAATCTTGAACGCACATTGCGCCCTCTGGTATTCCAGGGGATGCGCT GTTGAGCGTCACTTCCTCAAACCCCTCGGGTTGGTAGTGAGTGGTA CTCTTCTGGGTTAACCTGGAAAATGTCGGCATCTGCGTGTGACTG AGGTTTTAGTCCAGTCCGTTACTCTGCGTATTAGGTTTACCAACTC GTAGTGGCTTAGTAGGCCTTTAAGGCTTTACTGAAAGTACAGACA GTCTGGCAAACAGTATTATAAAGTTGACCTAAATCAGGTAGGATTA CCCGCTGAACCTAACGATATCAAA
OA 0.CM NS8 (O17)	<i>Lachancea thermotolerans</i>	99%	ITS	AACTTTATGAATACTGTTGCCAGACTGTCTGACTTCAGTAAAAGCCT TTAAAACGCCTACTAACGCCACTACGAGTTGGTAAACCTAACAGCAG AGTATCAGCGGACTGGACTAAACCTCAGTCAGCAACAGCAGATGGC CAGCATTTCAGTTAACCTGGAAAAGAGTACCACTCACTACCAACCCG AGGTTTTAGAAGGAAATGACGCTAACAGGGATGCCCCCTGGAAATA CCAGAGGGCGCAATGTGCGTCAAAGATTGCGATGATTCACTGAATATCTG CAATTACAATACTTATCGCATTGCGTGTCTCATCGATGCGAGAA CCAAGAGATCCGTTGGTAAAGTTAAATAATTTTCTAAATGAAAA ATAATTGACAATGTTAAATAATAATTGTTGTGTTGAACCCCTT GGGCGTAAAGCCCAAAGAAGAAGCGTTGAAATAATTACTCCACA GTGTGTGTAGAGAGACAGTCGTTAACAGAAAGCATCATGGCCCGC CAATCAAGCGCAGGCTCTGAACCTCCCGCTGCTAACAAATTCTTAAATGATCCTCCGCA
OA 0.CM NS9 (O18)	<i>Lachancea thermotolerans</i>	99%	ITS	AGCCGGGAAGTTCAAGAACGCTGCCTGATTGCGGCCGATGATGCTT TCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTTCA AACGCTTCTCTTGGGCTTACGGCCAAGGGTTACAAACACAAACA ATTATTGTATTTAACATTGTCAATTATTTCTATTAGAAAAAAAAT ATTAAAACCTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAA CGCAGCGAAATCGGATAAGTATTGTGAATTGCGAGATATTGCTGAATCAT CGAATCTTGAACGCACATTGCGCCCTCTGGTATTCCAGGGGATGCC TGTTGAGCGTCACTTCCTCAAACCCCTGGGTTGGTAGTGAGTGGT ACTCTTCTGGGTTACTGAAAATGTCGGCATCTGGCTGTGACTGAGGTT GAGGTTTAAGTCCAGTCCGCTGATACTCTGCGTATTAGGTTTACCAACT CGTGTGGCTTAGTAGGCCTTTAAGGCTTTACTGAAAGTACAGAC AGTCTGGCAAACAGTATTCAAAAGTTGACCTAAATCAGGTAGGATT ACCCGCTGAACCTAACGATATC
OA 0.CM NS10 (O19)	<i>Lachancea thermotolerans</i>	99%	ITS	ATTGAGGTCAACTTTATGAATACTGTTGCCAGACTGTCTGACTTC GAAAAGCCTTAAACGCCACTAACGCCACTACGAGTTGGTAAACCTA TAATACGCGAGTATCAGCGGACTGGACTAAACCTCAGTCAGCAACA GCCAGATGGCCAGATTTCAGTTAACCCAGAAAAGAGTACCACTCACT ACCAAACCCGAGGGTTAGAAGGAAATGACGCTAACACAGGCATGCC CCCTGGAATACAGAGGGCGCAATGTGCGTCAAAGATTGCGATGATCA CGAATATCTCAATTCAAATACTTATCGCATTGCGTGTCTCATC GATGCGAGAACCAAGAGATCCGTTGTTGAAAGTTAAATATTTTCTT CTAAATGAAAATAATTGACAATGTTAAATAATAATTGTTGTG TTGTAACCCCTGGGCGTAAAGCCCAAAGAAGAAGCGTTGAAATAATTACTCACAGTGTGTAGAGAGACAGTCGTTAACAGAAAGCAT CATGGCCGCGCAATCAAGCGCAGGCTCTGAACCTCCCGCTGCTAAC AAAAATTCTTAATGATCCTTC
OB 0.CM NS1 (O20)	<i>Lachancea thermotolerans</i>	99%	ITS	GATTGAGGTCAACTTTATGAATACTGTTGCCAGACTGTCTGACTTC AGTAAAAGCCTTAAACGCCACTAACGCCACTACGAGTTGGTAAACCTA TAATACGCGAGTATCAGCGGACTGGACTAAACCTCAGTCAGCAAC AGCCAGATGGCCAGCATTCAAGTTAACCCAGAAAAGAGTACCACTCAC TACCAAACCCGAGGGTTGAGAAGGAAATGACGCTAACACAGGCATGC CCCCTGGAATACAGAGGGCGCAATGTGCGTCAAAGATTGCGATGATTC ACGAATATCTCAATTCAAATACTTATCGCATTGCGTGTCTCATC CGATGCGAGAACCAAGAGATCCGTTGTTGAAAGTTAAATATTTTCTT CTAAAATGAAAATAATTGACAATGTTAAATAATAATTGTTGTGTTGTAACCTCCACAGTGTGTAGAGAGACAGTCGTTAACAGAAAGC ATCATGGCCGCGCAATCAAGCGCAGGCTCTGAACCTCCCGCTGCTAAC ACACAAATTCTTAATGATCCTTCAGGTTCACTACGGAAAGGTT GGAACACCCCGGAAGTTGAAAGACCGCCGGAAAGTTCGGACCCGA ACTTGTGCGGACGAGGATGCTTGTATAACGAC

Table A.1. (*continued*)

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OA 4.CM NS7 (O26)	<i>Lachancea thermotolerans</i>	99%	ITS	AAGCCTGCGCTTATTGCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTAAACCGCTTCTCTTGGGCTTACGGCCAAGGGTTACAACACAAACAAATTATTGATTTAAACATTGTCAATTATTTCATTAGAAAAAAATATTAAAACCTCAAACAGGATCTTGGTTCTGCATCGATGAAGAACGCCAGCGAAATGCGTAAGTATTGCAATTGCAAGATATTGTAATCATCGAATCTTGACGCACTTGGCCCTCTGGTATTCCAGGGGATGCCGCTTCTCAACACCCTGGGTTGGTAGTGAAGTGGACTCTTCTGGTAACTGAAAGTACAGACAGTCTGGCAAACTATTATAAGTTGACCTCAAATCAGGTAGGATTACCCGCTGAACCTAA
OA 4.CM NS8 (O27)	<i>Lachancea thermotolerans</i>	99%	ITS	GCCGGGAAGTTCAAGAACGCTGCGCTTATTGCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTAAACCGCTTCTTGGGCTTACGGCCAAGGGTTACAACACAAACAAATTATTGATTTAAACATTGTCAATTATTTCATTAGAAAAAAATATTAAAACATAACAAACTTCACAAACACGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATGCGATAAGTATTGTAATTGCAAGATATTGTAATCATCGAATCTTGAACTTIGAACGCACATTGCCCTCTGGTATTCCAGGGGATGCCGCTTGTGAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAAGTGGACTCTTCTGGGTTAACATGCTGGCCATCTGGCTTGTGACTGAGGTTTAGTCAGTCCGCTGATACTCTGGTATTAGGTTTACCAACTGTAGTGGCTTGTAGGCGTTAAAGGCTTTACTGAAAGTACAGACAGTCTGGAAACAGTATTCAAAAGTTGACCTCAAATCAGGTAGGATTACCCGCTGAACCTAA
OA 4.CM NS9 (O28)	<i>Lachancea thermotolerans</i>	99%	ITS	CAGAAGCCTGCGCTTATTGCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTAAACCGCTTCTTGGGCTTACGGCCAAGGGTTACAACACAAACAAATTATTGATTTAAACATTGTCAATTATTTCATTAGAAAAAAATATTAAAACCTAACACGGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATCGATAAGTATTGTAATTGCAAGATATTGTAATCATCGAATCTTGAAACGACATTGCCCTCTGGTATTCCAGGGGATGCCGCTTGTGAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAAGTGGACTCTTCTGGGTTAACATGCTGGCCATCTGGCTTGTGACTGAGGTTTAGTCAGTCCGCTGATACTCTGGTATTAGGTTTACCAACTGTAGTGGCTTGTAGGCGTTAAAGGCTTTACTGAAAGTACAGACAGTCTGGAAACAGTATTCAAAAGTTGACCTCAAATCAGGTAGGATTACCCGCTGAACCTAA
OA 4.CM NS10 (O29)	<i>Lachancea thermotolerans</i>	99%	ITS	AGCGGGGAAGTTCAAGAACGCTGCGCTTATTGCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTAAACCGCTTCTTGGGCTTACGGCCAAGGGTTACAACACAAACAAATTATTGATTTAAACATTGTCAATTATTTCATTAGAAAAAAATATTAAAACCTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAACCGCAGCGAAATGCGATAAGTATTGTAATTGCAAGATATTGTAATCATCGAATCTTGAAACGCACATTGCCCTCTGGTATTCCAGGGGATGCCGTTTGAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAAGTGGACTCTTCTGGGTTAACATGCTGGCCATCTGGCTTGTGACTGAGGTTTAGTCAGTCCGCTGATACTCTGGTATTAGGTTTACCAACTGGAGTTAGCCGTTAGTAGGCGTTAAAGGCTTTACTGAAAGTACAGACAGTCTGGCAAACAGTATTCAAAAGTTGACCTCAAATCAGGTAGGATTACCCGCTGAACCTAA
OB 4.CM NS1 (O30)	<i>Lachancea thermotolerans</i>	99%	ITS	GCCGGGAAGTTCAAGAACGCTGCGCTTATTGCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACACTGTGGAGTAATTATTAAACCGCTTCTTGGGCTTACGGCCAAGGGTTACAACACAAACAAATTATTGATTTAAACATTGTCAATTATTTCATTAGAAAAAAATATTAAAACCTTCACAAACGGATCTCTGGTCTCGCATCGATGAAGAACGCAGCGAAATGCGATAAGTATTGTAATTGCAAGATATTGTAATCATCGAATCTTGAAACGCACATTGCCCTCTGGTATTCCAGGGGATGCCGTTTGAGCGTCATTCTCTCAACACCCTGGGTTGGTAGTGAAGTGGACTCTTCTGGGTTAACATGCTGGCCATCTGGCTTGTGACTGAGGTTTAGTCAGTCCGCTGATACTCTGGTATTAGGTTTACCAACTGTAGTGGCTTGTAGGCGTTAAAGGCTTTACTGAAAGTACAGACAGTCTGGCAAACAGTATTCAAAAGTTGACCTCAAATCAGGTAGGATTACCCGCTGAACCTAAAGCATATCAATA

Table A.1. (*continued*)

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
OB 4.CM NS10 (O36)	<i>Lachancea thermotolerans</i>	99%	ITS	GCGCGGCCGATGATGCTTCTGTTAACGACTGTCTCTACACACAC TGTGGAGTAATTATTTACAACCGCTTCTTGGCTTACGGCCAA GGGTTACAAACACAACAAACATTATTGTATTAAACATTGTCAATTATTT TCATTTAGAAAAAAAATATTAAAACCTTCAACAACGGATCTTGGT TCTCGCATCGATGAAGAACGCAGCGAAATGCGATAAGTATTGTGAATTG CAGATATTCCGGAATCATCGAATCTTGAAACCGCACATTGCGCCCTTGG TATTCCAGGGGCATGCCCTGTTGAGCGCTATTCCCTCTCAAACCCCTG GTTTGGTAGTGAGTGACTCTTCTGGTTACTTAAAGTGTG CATCTGGCTTGTGACTGAGGTTTATGCCAGTCCGCTGATACTCTGC GTATTAGGTTTACCAACTGTAAGTGGCTTAGTAGGCCTTTAAAGGC TTTACTGAAAGTACAGACAGTCTGGCAACAGTATTCTAAAGTTGA CTCAAATCAGTAGGATTACCCGCTGAACCTAAAGCATAT
OA 4.CM S3 (O37)	<i>Wickerhamomyces anomalus</i>	100%	D1/D2 Domain	CGAGTGAAGCGGCAAAAGCTCAAATTGAAATCTAGCACCTCGGTGTT CGAGTTGTAATTGAAAGATGGTAAACCTGGGTTGGCTTGTCTATGTT CCTTGGAACAGGACGTAGAGGGTGAGAATCCCGTGTGAGATG CCCCTCTATGTAAGGTGCTTGAAGAGTGCAGTTGTTGGGAAATG AGCTCTAAGTGGTGGTAATTCCATCTAAAGCTAAATATTGGCAGAG ACCGATAGCGAACAAAGTACAGTGTGAAAGATGAAAAGAACTTGA AAGAGAGTAAAAAGTACGTGAAATTGTTGAAGGGAAAGGGCATTAGA TCAGACTTGGTGTACGATTCTCTTGTGAGTTGTCAGTCTG ATTCACTGGGCCAGCATGATTGGATGGCAAGATAATGGCAGTGAA TGTGGCTTCACTTGGTGGAGTGTATAGCTTGTGATATTGCCGTC TGGATCGAGGGCTGCGTTGTGACTAGGATGCTGGCGTAATGATCTAA TGCCGCCGCTT
OB 0.CM S6 (O38)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CCTTAGTACCGCGAGTGAAGCGGCAAAAGCTCAAATTGAAATCTGGT ACCTTCCGGTCCCGAGTTGTAATTGGAGAGGGCAACTTGGGCCGTT CCTTGTCTATGTTCTTGGAACAGGACGTAGAGGGTGAGAATCCCG TGTGGCGAGGAGTGCAGTTGTTAAAGTGCCTCGAAGAGTCAGTT GTTTGGGAATGCAGCTAAAGTGGGTGTAATTCCATCTAAAGCTAA TATTGGCGAGAGACCGTAGCGAACAAAGTACAGTGTGAAAGATGAA AAGAACCTTGGAAAGAGAGTGAAGGAAAGTACGTGAAATTGTTGAAGGG AAGGGCATTGATCAGACATGGTTGCTTGTGCCCTGCTTGTGGT AGGGGAATCTCGCATTTCACTGGGCCAGCATCAGTTGGTGGCAGGAT AAATCCATAGGAATGTAGCTTGCCTCGTAAGTATTATAGCCTGTGGG ATACTGCCAGCTGGACTGAGGACTGCGACGTAAGTCAAGGATGCTG CATATGGTTATGCGCCCGTC
OB 4.CM S2 (O40)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	GGCAAAGCTCAAATTGAAATCTGTACCTCGTGCCTGAGTTGAA TTGGAGAGGGCAACTTGGGCCGTTCTGTCTATGTCCTTGAAC AGGACGTAGAGGGTGGAGAATCCCGTGTGGCGAGGAGTGCAGCTCTAAG TGTAAAGTGCCTTGAAGAGTGCAGTTGTTGGGAATGCAAGCTCTAAG TGGGTGGTAATTCCATCTAAAGCTAAATATTGGCGAGAGACCGATAGC GAACAAGTACAGTGTGAAAGATGAAAAGAACTTGAAGAGAGTG AAAAAGTACGTGAAATTGTTGAAGGGAGGGCATTGATCAGACATG GTGTTTGTGCCCTGCTCTGTGGTAGGGGAATCTGCATTCACT GGCCAGCATCAGTTGGTGGCAGGATAATCCAAGGAATGTAGCTT GCCTCGTAAGTATTATAGCTGTGGGAATACTGCCAGCTGGACTGAG GACTGCGACGTAAGTCAAGGATGCTGGCATAATGGTTATGCGCCCG TCT
OB 4.CM S1 (O39)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGGCGAGTGAAGCGGCAAAAGCTCAAATTGAAATCTGTACCTCGGT GCCGAGTTGTAATTGGAGAGGGCAACTTGGGCCGTTCTGTCTA TGTCCTTGGAACAGGACGTAGAGGGTGGAGAATCCCGTGTGGCGA GGAGTGCCTCTTGTAAAGTGCCTCGAAGAGTCAGTTGTTGGGA ATGCAGCTAAGTGGGTGTAATTCCATCTAAAGCTAAATATTGGCG AGAGACCGATAGCGAACAAAGTACAGTGTGAAAGAGTGAAGAAACTT TGAAAAGAGAGTGAAGGAAAGTACGTGAAATTGTTGAAGGGAGGGCAT TTGATCAGACATGGTTGCTTGTGCCCTGCTCTGTGGTAGGGGAAT CTCGCATTCACTGGGCCAGCATCAGTTGGTGGCAGGATAATCCAT AGGAATGTAGCTGCTCGTAAGTATTATAGCCTGTGGGAATACTGCC AGCTGGGACTGAGGACTGCGACGTAAGTCAAGGATGCTGGCATAATGG TTATATGCCGCC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BA 0.CM NS2 (B2)	<i>Saccharomyces cerevisiae</i>	100%	ITS	GTGTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAAGACAAGAGATGGAGAGTCCAGCGGGCTGCGCTTAAGTGCAGGCTCTGCTAGGCTGTAAGTTCTTCTGCTATTCCAACACGGTGAGAGATTCTGTGCTTTGTATAGGACAATTAAACCGTTCAATACAACACACTGAGCTTCAATCGAGCAATCGGGGCCAGAGGTAAACAAACAAACAATTTTATCATTCAATTAAATTTGTCAAACAAACAAAGAATTTCGTAACTGGAATTTTAAATATTAAAACATTTCAACAAACGGATCTCTGGTCTCGCATCGATGAAGAACGCAACGATACGTAATGTGAATTGAGAATTCCCGTGAATCATCGAATCTTGAACGACATTGCGCCCTGATTCCAGGGGATGCTGTGAGCCTATTCCCTCTCAAACATTCTGTTGGTAGTGAGTGAATCTTGGAGTTAACTTGAAAATGCTGCTTCAATGGATTTTTCAAAGAGAGGTTCTGCCTGCTGAGGTATAATGCAAGTACGGCTTCTAGGTTACCAACTGGCTAATGGAACGTTATCGATAAGAAGAGCGT
BA 0.CM NS3 (B3)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTTGTTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAAGAACAGAGATGGAGAGTCCAGGGGCCCTGCGCTTAAGTGCAGGCTGTTAGCTGCTTCTGCTATTCCAACACGGTGAGAGATTCTGTGCTTGTATAGGACAATTAAACACACTGTGGAGTTTCAATATCTTGCACACTTTCTTGGCATTGAGCAATCGGGCCAGAGGTAAACAAACAAACAATTTTATCATTCAATTAAATTTGTCAAACAAACAAATTAAACATTAAACAAAGAATTTCGTAACTGGAATTTTAAATTTAAATTTAAACATTAAACAAAGATTCACAAACGGGATCTTGGTCTCGCATCGATGAAGAACGCAACGCAAATCGGATACGTAATGTGAATTGAGAATTCCCGTGAATCATCGAATCTGAGTACATTGAAACCGCACATTGCGCCCTGGTATTCCAGGGGATGCTGTGAGCGTCATTCTCTCAAACATCTGTTGGTAGTGAGTGAATCTTGGAGTTAACCTGAAATTGCTGGCTTCTGATGGAGTTTTCAAAGAGAGTTCTGCGTGTGAGGTATAATGCAAGTACGGCTTCTAGGTTACCAACTGCGGCTAATCTTTTATAGCAGCGTATTGAAACGTTATC
BA 0.CM NS4 (B4)	<i>Solicoccozyma aeria</i>	99%	D1/D2 Domain	ATTTGAAATCTGCAGCCTCAGGTTGCCAGTGTAACTCTATAGAACGTTTCCGCGCTGGCCCAGTACAAGTCCCTTGAATAGGGCGTCATAGAGGGTGAGAATCCCGCTCTGACACGGGACCCAGTGCCTTGTGATACGTTTCACAGAGCTGAGTTGGGAATGTCAGCTGCAATGGTGGTAAATTCCATCTAAAGCTAAATATTGGCAGAGACCGATAGCGAACAGTAACCGTGAGGGAAAGATGAAAAGCACTTGGAAAGAGAGGTTAACAGTATGTGAAATTGTAAGGGAAACGATTGAGAATGCACTGCTGTCTATGGGACTCAGCCGGTTCTGCGGTGACTCTCCTTGAAGATGGGTCACATCAGTTTGATCGTGGAAAAGGGCGGGAGGAATGAGACTCTCGGGTGAACCTATAGCCTTCCCGTGTACAGGTGGTGGGACTGAGGAACGCAACGATGCTTATGCCGGGTTGCCACGTACATGCTTAGGATGTTGACATAATGGCTTAAACAACCGCTCT
		100%	ITS	GCAAGGGCCCTGCTTAACTCACATCCAAACACCTGTGAACGTGAAGCGCATGACTAGGTTGCCAAGTCATGCTCTGCCCTTTAACAAACAAATTAAATGTAACAAACGTAAGTCTTATTATAACCTAAATAAAACCTTCAACAAACGAGATCTCTGGCTCTCGCATCGATGAAGAACGCAACGAAATGCGATAAGTAATGTGAATTGAGAATTCACTGTAATCATCGAATCTTGAACGCAACCTTGCCTTGGTATTCCGAAGAGCATGCCCTGTTGAGTGTCTGAAATATCAACCTTGGACTGGGTTGTGCTCTAGTCTCGGCTTGGAAATTGGGTGCTTGGCTTACAGGCCGCTCACCTAAATGTATTAGCTGGATCTGCTTGGAGACTGTTGACTTGGCTAATAAGTATTGCTAAGGACATTCTCGGAGTGGCCTGTTCTAGGACGCTTGCCTTCAATACAAGTTCCACCTCGTGGACATGACTTTTATTATCTGCCCTCAAATCAGGTAGGACTACCCGCT
BA 0.CM NS5 (B5)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TTATCGATAACGTTCAATACGCTCAGTATAAAAAAGATTAGCCGAGTGGTAAAACCTAAACGACCGTACTTGCATTACCTCAAGCACGAGAGAAACCTCTTGGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACAGAGTATCACTCACTACCAACAGAAATGTTGAGAAGGAATGACGCTCAAACAGGCATGCCCTGGAAATACCAAGGGCGCAATGGGCGTTCAAAGATTCGATGATTGAGTACGGAAATTCTGCAATTACATTACGTATCGCATTTCGCTCGTCTCATCGATGCGAGAACCAAGAGATCGTTGTTGAAAGTTTAAATATTAAAATTCAGTTACGAAAATTCTGTTTGACAAAATTAAATGAATGATAAAATGTTGGGTTGTACCTCTGGGCCGATTGCTGAATGCCAAAGAAAAGTTGCAAAGATATAAAAGCACAGAAATCTCACCGTTGGAATAGCAAGAAAGAAACTTAACAGCCTAGCAAGACGCCACTTAAGCGCAGG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BA 0.CM NS6 (B6)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ACTTTAAGAACATTGTTGCCCTAGACGCTCTTCTTATCGATAACGTTCAATACGCTCAGTATAAAAAAAGATTAGCCGCAGTTGTAACACTAA AACGACCGTACTTCGATTATACTCTCAAGCACGCAAAGAAACCTCTCTTGAAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTAACTCCA AAGAGTATCACTCACTACCACAGAAATGTTGAGAAGGAATGACGC TCAAAACAGGATGCCCTGGAAATACCAAGGGGCGCAATGCGTTCA AGAGTCGATGATTCAAGGAATTCTGCAATTCAACATTACGTATCGCATT TCGCTCGTCTTCATCGATGCGAGAACCAAAGAGATCCGTTGAAAG TTTTAATATTTAAAATTCCAGTTACGAAAATTCTGTTTGACAAA ATTTAATGAAATAGATAAAATTGTTGTTGTTACCTCTGGGCCCGA TTGCTGAATGCCAAAGAAAAGTGCAGAAAGATATGAAACACTCCACA GTGTTGTTATTGAAACGTTTAAATTGCTCTATAACAAAAGCACGAA ATCTCTACCGTTGGAATAGCAAGAAAGAAACTACAAGCTAGCAA GACCGCGCACTTAAGCGCAGGCCGCTGGACTCTCCATCTTGTCTT CTTGCCAGTAAAGCTCATGCTTGCACAAACAAAAATTCATTTCACAAATTATTAATTTCAATGATCCTTCGC
BA 0.CM NS7 (B7)	<i>Saccharomyces cerevisiae</i>	100%	ITS	AAAATGGATTTTTGTTTGGCAAGAGCATGAGAGCTTTACTGGGCA AGAAGACAAGAGATGGAGAGTCCAGCCGGCTGCGCTTAAGTGC CGCTTGTAGGCTTGTAAAGTTCTTCTGCTATTCAAACGGTAGAGA TTTCTGTGCTTGTATAAGGACAATTAAAACCGTTCAATACAACACAC TGTGGAGTTCTCATCTGCAACTTTCTTGGCATTGAGCAATC GGCCCCAGAGTAACAAACACAATTAACTTCTATTCAATTAAATT TTGTCAAAACAAGAATTCTGTAACTGGAAATTAAAATATTTAAA CTTCAACACGGATCTTGGTCTCGCATCGATGAAGAACGCGAGCGA ATATGCGATACTGAAATGCAATTGAGAATTCCGTGAATCATGAACTT TGAACCGCACATTGCGCCCTTGGTATTCCAGGGGCAATGCCCTGTTGAG CGTCATTCTCTCAAACATTCTGTTGGTAGTGAGTGATACTCTTGG AGTTAACTGAAATTGCTGGCTTTCATGGATGTTTTTCCAAAGA GAGGTTCTGCGTGTGAGGTATAATGCAAGTACGGTCGTTAGG TTTTACCAACTGCGCTAATCTTTTATATGAGCGTATTGGAAAGCTT ATCGATAAGAAGAGACGCTAGGCAACAAATGTTCTAAAGTTGACC TCAAATCAGGTTAGGAGTACCCGCTGAACCTAACGATATCAATAA
BA 0.CM NS8 (B8)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TGTTTGGCAAGAGCATGAGAGCTTTACTGGGCAAGAACAGAG ATGGAGAGTCCAGCCGGCTGCGCTTAAGTGCAGGCTTGTAGGCT TGTAAGTTCTTCTGCTATTCAAACGGTAGAGAGATTCTGTGCTTT GTTATAGGACAATTAAACCGTTCAATACAACACACTGTGGAGTTTC ATATCTTGCAACTTTCTGGCATTGAGCAATCGGGGCCAGAG GTAACAAACAAACAAATTAACTTATCTATTCAATTAAATTGTCAAAAC AAGAATTCTGTAACTGGAAATTAAAATATTTAAAACATTCAACAC GGATCTCTGTTCTCGCATCGATGAAGAACGAGCGAAATGCGATACG TAATGTGAATTGAGAATTCCGTGAATCATCGAATCTTGAACGACAT TGGCCTCTGGTATTCCAGGGGCAATGCCCTGTTGAGCGTCATTCTT CTCAAACATTCTGTTGGTAGTGAGTGATACTCTTGGAGTTAACTGAA ATTGCTGCCCTTTCATTGGATGTTTTTCCAAAGAGAGGGTTCTCG CGTGTGTTGAGGTATAATGCAAGTACGGTCGTTAGGTTTACCAACTG CGGCTAATCTTTTATACTGAGCGTATTGGAACGTTATCGATAAAAA AAAAGCGCTAGGCAACAAATGTTCTAAAGTTGACCTCAAATCAGGT AGGAGTACCC
BB 0.CM NS1 (B9)	<i>Saccharomyces cerevisiae</i>	100%	ITS	GAGAGCTTTACTGGGCAAGAACAGACAAGAGATGGAGAGTCCAGCCGG CCTGCGCTTAAGTGCAGGCTTGTAGGCTTGTAAAGTTCTTCTGCT ATTCAAACCGTGGAGAGATTCTGTGCTTTGTATAAGGACAATTAAA CCGTTCAATACAACACACTGTGGAGGTTCTCATATCTTGCAACTTTCT TTGGCATTGAGCAATCGGGGCCAGAGGTAAACAAACACAAT TTTATCTATTCAAATTGTCAAACAAAGAATTCTGTAACTGGAA ATTAAAATTTAAACAAACTTCACAACAGGATCTTGGTTCTCGCAT CGATGAAGAACGCAAGCGAAATGCGATACGTAATGTAATTGCAAGATT CCGTGAATCATGAAATCTTGAACGCAATTGCGCCCTTGGTATTCCA GGGGGCATGCCCTGTTGAGCGTCATTCTCTCAACATTGTTGGT AGTGAGTGATACTCTTGAGGTTACTGAAATTGCTGGCCTTTCATG GATGTTTTTCCAAAGAGAGGGTTCTGCGTGTGAGGTATAATGC AAAGTACGGTCGTTAGGTTACCAACTGCGCTAATCTTTTATAC TGAGCGTATTGAAACGTTATGATAAGAAGAGAGCGCTAGGCGAACAA AAGCATATCA

Table A.1. (*continued*)

Table A.1. (*continued*)

Name	BLAST Result	Similarity	Region	Sequence
BB 0.CM NS7 (B15)	<i>Metschnikowia pulcherrima</i>	97%	ITS	AACTCTAACCTTAAACCTCAATAACTTATTAAAAAAACTTCAACAACG GATCTCTGGTTCTCGCATCGATGAAAACGCACCGAATTCGATACGT AATATGACTTGCACAACTGAATCATTGAAATCTTGAAACGCACATTGCGC CCCGGGGTATTCCCCAGGGCATGCGTGGGTGAGCGATATTACTCTCAA ACCTCCGGTTGGTCTGCTCGGCCCTAATATCAACCGCGCTAGAATAA GTTTACCCCATTCTTTCTCACCTCGTAAACTACCCGCTGAAC TAACCATATCA
BB 0.CM NS8 (B16)	<i>Saccharomyces cerevisiae</i>	99%	ITS	GGCAAGAGCATGAGAGCTTTACTGGGCAAGAAGACAAGAGATGGAGA GTCCAGCCGGCCTGCCTTAAGTGCCTGGCTTAGGCTTGTAAAGT TTCTTCTTGTCTATTCCAACACGGTGAAGAGATTCTGTGCTTGTATAG GACAATTAACACCCTTCAATACAACACACTGTGGAGTTTCATATCTT TGCAACTTTCTTGGCATTTCGAGCAATCGGGGCCAGAGGTAAACAA ACACAAACAATTTATCTATTCAATTAATTTCGTCAAAACAAAGAATT TCCTGAATGGAAATTAAAATATTAAACACTTCAACACGGATCTCT TGGTCTCGCATCGATGAAGAACGCAGCGAAATGCAGTACGTAAATGTG AATTCGAGAATCCGTGAATCATCGAACATTGGAACGCCATTCGCCC CTTGGTATTCCAGGGGCATGCCTGTTGAGCTCATTCCTTCAAAAC ATTCTGTTGGTAGTGAGTGATACTCTTGGAGTTAATCTGAAATTGCTG GCCTTTCATGGATTTTTCCAAAGAGAGGTTCTGCGTCTGAGGTAA GAGGTATAATGCAAGTACGGCTGTTAGGTTTACCAACTCGGGCTAA TCTTTTTTATACTGAGCGTATTGGAACGTTACGATAAGAAGAGAGC TCTAGGCAGAACATGTTTAAAGTTGACCTCAATCANGTAGGAGTA CCCGCTGA
BA 4.CM NS1 (B17)	<i>Saccharomyces cerevisiae</i>	100%	ITS	AGCTTTACTGGCAAGAAGACAAGAGATGGAGAGTCCAGCCGGCC GCGCTTAAGTGCCTGGCTTGCTAGGCTTGTAAAGTTCTTCTGCTATT CCAAACCGTGAGAGATTCTGTGCTTGTATAGGACAATTTAACCG TTCAATACAACACACTGGAGTTTCATATCTTGCACATTTCCTT GGGCATTCGAGCAATCGGGGCCAGAGGTAAACAAACACAAACATT ATCTATTCAATTAATTTCGTCAAAACAAAGAATTTCGTAATGGAAA TTTAAATATTTAAACACTTCAACAACGGATCTTGGTCTCGCATCG ATGAAAGAACGCAGCGAAATGCGATACGTAATGTGAGAATTCGAGGTTATCC GTGAATCATCGAATCTTGAACGCACATTGCGCCCTGGTATTCCAGG GGGCATGCCTGTTGAGCGTCATTCTCTCAACATTCTGTTGGTAG TGAGTGATACTCTTGGAGTTAATCTGAAATTGCTGGCCTTTCATTGGA TGTTTTTCCAAAGAGAGGTTCTGCGTGTAGGTATAATGCAA GTACGGTCTTTAGGTTTACCAACTCGGGCTAATCTTTTATACTG AGCGTATTGGAACGTTACGATAAGAAGAGAGCCTAGGCAGAACAT GTCTTAAAGTTGACCTCAATCA
BA 4.CM NS5 (B21)	<i>Saccharomyces cerevisiae</i>	99%	ITS	AGCTTTACTGGCAAGAAGACAAGAGATGGAGAGTCCAGCCGGCC GCGCTTAAGTGCCTGGCTTGCTAGGCTTGTAAAGTTCTTCTGCTATT CCAAACCGTGAGAGATTCTGTGCTTGTATAGGACAATTTAACCG TTCAATACAACACACTGGAGTTTCATATCTTGCACATTTCCTT GGGCATTCGAGCAATCGGGGCCAGAGGTAAACAAACACAAACATT ATCTATTCAATTAATTTCGTCAAAACAAAGAATTTCGTAATGGAAA TTTAAATATTTAAACACTTCAACAACGGATCTTGGTCTCGCATCG ATGAAAGAACGCAGCGAAATGCGATACGTAATGTGAGAATTCGAGGTTATCC GTGAATCATCGAATCTTGAACGCACATTGCGCCCTGGTATTCCAGG GGGCATGCCTGTTGAGCGTCATTCTCTCAACATTCTGTTGGTAG TGAGTGATACTCTTGGAGTTAATCTGAAATTGCTGGCCTTTCATTGGA TGTTTTTCCAAAGAGAGGTTCTGCGTGTAGGTATAATGCAA GTACGGTCTTTAGGTTTACCAACTCGGGCTAATCTTTTATACTG AGCGTATTGGAACGTTACGATAAGAANAGAGCCTAGGCAGAACAT GTCTTAAAGTTGACCTCAATCAAGTAGGTTAGGATACCC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BA 4.CM NS6 (B22)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TTAAGAACATTGTCGCCTAGACGCTCTTATCGATAACGTTCA ATACGCTCAGTATAAAAAGATTAGCCGCAGTTGGTAAACCTAAAA CGACCGTACTTGCATTATACCTCAAGCACGCAAAGAAACCTCTTGG AAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTAACTCCAAA GAGTATCACTCACTACCAACAGAATGTTGAGAAGGAATGACGCTC AACAGGCATGCCCTGGAATACCAAGGGCGCAATGTGCGTCAAA GATTGATGATTCAAGGAATTCTGCATTACACATTACGTATCGCATTTCG CTGCGTTCTCATCGATGCCAGAACAGAGATCGTTGAAAGTT TTAATATTTAAAATTCCAGTTACGAAAATTCTGTTTGACAAAAAT TTAATGAATAGAAAAATTGTTGTTGACCTCTGGGCCCCGATTG CTCGAATGCCAAAGAAAAAGTGCACAGATGAAAACCTCCAGTG TGTGTTGATTGAAACGGTTTAAATTGCTTATAACAAAAGCACAAATC TCTCACCGTTGGAATAGCAAGAAGAAACTTACAAGCTAGCAAGAC CGGCCACTTAAGCGCAGGCCGGCTGACTCTCCATCTTGCTCTTG CCCAGTAAAGCTCATGCTCTGCAAAACAAAAAAATCCATTTC AAATTATTAATTCTTAATGATCCTTCAGCAGT
BA 4.CM NS7 (B23)	<i>Saccharomyces cerevisiae</i>	98%	ITS	ACTTTAACATGTCGCCTAGACGCTCTTATCGATAACGTT CAATACGCTCAGTATAAAAAGATTAGCCGCAGTTGGTAAACCTAA AACGACCGTACTGCAATTACCTCAAGCACGCAAACCTCTT GGAAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTAACTCCA AAAAGTATCACTCACTACCAACAAAATGTTGAAAAGGAATGACCC TCAACAGGCATGCCCTGGAATACCAAGGGCGCAAGGTGCGTC AAGATTGATGATTACCGAATTCTGCATTACATTACGTATCGCATT TCCCTCGTCTTCATCGATGCCAGAACAAAAATCCGTTGAAAG TTTTAATATTTAAAATTCCAGTTACAAAATTCTGTTTGACAAA AATTAATGAATAAAATTGTTGTTGTTGACCTCTGGGCCCCGA TGTGCGAATGCCAAAGAAAAAGTGCACAAATATGAAAACCTCCACA GTGTGTTGATTGAAACGGTTTAAATTGCTTATAACAAAAGCACAAA ATCTCTCACCGTTGGAATAGCAAGAAGAAACTTACAAGCTAGCAA GACCGCGACTTAAGCGCAGGCCGGCTGACTCTCCATCTTGCTT CTGCCAGTAAAGCTCATGCTCTGCAAAACAAAAAAATCCATT TCAAAATTATTAATTCTT
BA 4.CM NS8 (B24)	<i>Saccharomyces cerevisiae</i>	98%	ITS	AACTTTAACATGTCGCCTAGACGCTCTTATCGATAACGTT CCAATACGCTCAGTATAAAAAGATTAGCCGCAGTTGGTAAACCTA AACGACCGTACTGCAATTACCTCAAGCACGCAAAGAAACCTCTT TGAAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTAACTCC AAAGAGTATCACTCACTACCAACAGAATGTTGAAAAGGAATGACG CTCAACAGGCATGCCCTGGAATACCAAGGGCGCAAGGGCGTTC AAAGATTGATGATTACCGAATTCTGCATTACATTACGTATCGCAT TTCGCTCGTCTTCATCGATGCCAGAACAAAAATCCGTTGAAAG TTTTAATATTTAAAATTCCAGTTCAAAAATTCTGTTTGACAA AAATTAATGAATAGATAAAATTGTTGGTTGTTACCTCTGGGCCCC GATTGCTGAATGCCAAAGAAAAAGTGCACAGATATGAAAACCTCC CAGGGGGTTGATTGAAACGGTTTAAATTGCTTATAACAAAAGCACAG AAATCTTCACCGTTGGAATAGCAAGAAGAAACTTACAAGCTAGCA AGACCG
BB 4.CM NS1 (B26)	<i>Saccharomyces cerevisiae</i>	100%	ITS	GGATTTTTTGTTGGCAAGAGCATGAGAGCTTTACTGGCAAGAAG ACAAGAGATGGAGACTCAGGCCGGCTCGCCTTAAGTGCAGGCTT GCTAGGCTTGTAAGTTCTTCTGCATTCCAAACCGTTCAATACAACACTGTG GAGTTTCACTTGCACATTCTGGCATCGAGCAATCGGG CCCAGAGGTAAACAAACAAACAATTCTATTCAATTAAATTTTGT AAAAACAAAGAATTCTGTAATTGGAATTAAATTAAATTAAAAACTTT CAACACGGATCTTGGCTCGCATCGATGAAGAACGCAAGCGAAATG CGATACGTAATGTAATTGAGAATTCCGTGAATCATCGAATCTTGAA CGCACATTGCGCCCTGGTATTCCAGGGGCTGCTGTTGAGGTC ATTCCTCTCAAACACATTCTGTTGGTAGTGAGTGATACTCTTGAGGTT AACTGAAATTGCTGGCCTTTCAATTGGATGTTTTTCAAAAGAGGAG TTCTCTGCGTGTGAGGTATAATGCAAGTACGGTGTGTTAGGTTA CCAATGCGGCTAATCTTTTATACTGAGCGTATTGAAAGCTTAC TCAGGTAGGAGTACCGCTGAACATAC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BB 4.CM NS2 (B27)	<i>Saccharomyces cerevisiae</i>	99%	ITS	TGTTTGGCAAGAGCATGAGAGCTTACTGGCAAGAAGACAAGAGA TGGAGAGTCAGCCGGCCTGCGCTTAAGTGCAGCTTGTAGCCT GTAAGTTCTTCTGCTATTCAAACCGTGAGAGATTCTGTGCTTTG TTATAGGACAATAAAACCGTTCAATACAACACACTGTGGAGTTCA TATCTTGCAACTTTCTTGGCATTGAGCAATCGAGCACTGGGCCAGAGGT ACAAAACACAAACAATTATCTATTCAAATTTGTCAAAACAA GAATTTCGTAACTGGAATTAAATTTAAATATTAAAAACTTCAACACGG ATCTCTGGTCTCGCATCGATGAAGAACCGAGCAGCAAATGCAGATACTGA ATGTGAATTGCAAGATTCCGTGAATCATCGAATCTTGAACGCACATTG CGCCCCTTGGTATTCCAGGGGGCCTGCTTTGAGCGTCACTTCTTCT CAAACATTCTGTTGGTAGTGAGTGACTCTTGGAGTTACTGAAA TTGCTGGCTTCTCATGGATTTTATTCAAAGAGAGGTTCTGC GTGCTGAGGTATAATGCAAGTACGGTGTCTTAGGTTTACCAACTGC GGCTAATCTTTTATACTGAGCGTATGGAACGTTATCGATAAGAAG AGAGCGCTAGGCGAACATGTTCTAAAGTTGACCTCAAATCAGGT GGAGTACCCGCTGAACCTAACATATCATAA
BB 4.CM NS3 (B28)	<i>Saccharomyces cerevisiae</i>	100%	ITS	TGGATTTTTGTTTGGCAAGAGCATGAGAGCTTACTGGCAAGAA GACAAGAGATGGAGAGTCAGCCGGCCTGCGCTTAAGTGCAGCT TGCTAGGTTGTAAGTTCTTCTGCTATTCAAACCGTGAGAGATTTC TGTGCTTTGTTAGGACAATTAAACCGTTCAATACAACACACTGT GGAGTTTCATATCTTGAACCTTTGGCATTGAGCAATCGAGCAATCGG GCCCAGAGGTAACAAACACAACAAATTATCTATTCAAATTTG TCAAAAACAAGAATTCTGAACCTGGAATTAAATTTAAATATTAAAAACTT TCAACAACGGATCTTGTGCTCGCATCGATGAAGAACCGAGCGAAT CGGATACGTAATGTGAATTGAGAATTCGGTGAATCATCGAATCTTGA ACGCACATTGCGCCCTGGTATTCCAGGGGGCATGCTGTTGAGCGT CATTCTCTCAAACATTCTGTTGGTAGTGAGTGACTCTTGGAGT TAACTGAAATTGCTGGCTTTCATGGATGTTTTTCAAAGAGAG GTTTCTGCGTGTGAGGTATAATGCAAGTACGGTGTCTTAGGTTT ACCAACTGCGCTAATCTTTTATACTGAGCGTATGGAACGTTATCG ATAAGAAGAGAGCGCTAGGCGAACATGTTCTAAAGTTGACCTCAA ATCAGGTAGGAGTACCCGCTGAACCTAACATATCATA
BB 4.CM NS4 (B29)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATTTTTGTTTGGCAAGAGCATGAGAGCTTACTGGCAAGAAGAC AAGAGATGGAGAGTCAGCCGGCCTGCGCTTAAGTGCAGCTTGT AGGCTTGTGTAAGTTCTTCTGCTATTCAAACCGTGAGAGATTCTGTG CTTTGTTATAGGACAATTAAACCGTTCAATACAACACACTGTGGAG TTTCATATCTTGAACCTTTCTTGGCATTGAGCAATCGGCC AGAGGTAACAAACACAACATTATCTATTCAAATTTGCAA AAACAAGAATTCTGAACCTGGAATTAAATTTAAATATTAAAAACTTCAA CAACGGATCTTGTGCTCGCATCGATGAAGAACCGAGCGAAATCGA TACGTAATGTGAATTGAGAATTCGGTGAATCATCGAATCTTGAACGC ACATTGCCCCCTGGTATTCCAGGGGGCATGCTGTTGAGCGTCAATT CCTCTCAAACATTCTGTTGGTAGTGAGTGACTCTTGGAGTTACT TGAAATTGCTGGCTTTCATGGATGTTTTTCAAAGAGAGGTTTC TCTGCGTGTGAGGTATAATGCAAGTACGGTGTCTTAGGTTTACCA ACTGCGGCTAATCTTTTATACTGACCGTATTGGAACGTTATCGATAA AAAAAGCGCTAGGCAACAAATGTTCTAAAGTTGACCTCAAATCA GGTAGGAGTACCCGCTGAACCTAACATATCATA
BB 4.CM NS5 (B30)	<i>Metschnikowia pulcherrima</i>	96%	ITS	TAACTTTATTAACAAACGGATCTGGGTCGCATCG ATGAAAACCCACCGAATGCGATACTTAATATGACTTCAAATGAA TCATTGAATCTTGAACGACATTGCGCCGGGTATTCCCAGGGCA TGCCTGGGTGAGCGATTTACTCTCAAACCTCGGGTTGGCTGCTTC GGCTAATATCAACGGCCCTAGAATAATTACCCCATCTTCC CACCCCTCGAAAACCTACCGCTGAACCTAACATATCATA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BB 4.CM NS6 (B31)	<i>Saccharomyces cerevisiae</i>	99%	ITS	ATGGATTTTTGTGCAAGAGCATGAGAGCTTACTGGCAAGAAGACAAGAGATGGAGAGACTCCAGCGGCCGCTTAAGTGCGCGCTTGCTAGGCTTGAAGTTCTGCTATTCCAAACGGTGAGAGATTCTGTGCTTGTATAGGACAATTAAAACCGTTCAATACAAACACACTGTGGAGTTTATCTTGTCAACTTTCTTGGCATTGAGCAATCGGGCCAGCCCAGAGGTAACAAACACAAACATTTCGTAACTGGAATTTTAAATAATTAAAAAACCTTCACAAACAGCGAAGAACCGAGCGAAATCGATACGTAATTGCAATCTGGAATCTGAACTTTGAAACCGACATTGCGCCCTTGGTATTCCAAACATCTGTGGTAGTGAGTGATACTTTGGAGTTAACTGAAATTGCTGGCTTGTAGGTATAATGCAAGTACGGCTGTTTANGTTTACCAACTGCGGCTAATCTTTTTATACTGAGCGT
BB 4.CM NS7 (B32)	<i>Saccharomyces cerevisiae</i>	99%	ITS	GGCAAGAGCATGAGAGCTTACTGGCAAGAAGACAAGAGATGGAGAGTCAGCGCCGCGCTTAAGTGCGCGCTTGAAGTTCTGCTAGGCTTGAAGTTCTGCTTTGTATAGGACAATTAAAACCGTTCAATACAAACACACTGTGGAGTTTATCTTGTCAACTTTCTTGGCATTGAGCAATCGGGCCAGAGGTAACAAACACAAACATTTCGTAATTCTTGTCAAATTGAGGAAATTAAATAATTAAATAACACAGGATCTCTGTTCTGCATCGATGAAGAACGCGAGGAAATCGATACGTAATGTGAAATTGAGAATTCCGTGAATCATGAATCTTGAACGCACTTGCAGCCCTTGGTATTCCAGGGGCAATGCCATTGAGCGTCATTCTCTCAAACATTCTGTTAGTGAGTGATACTTTGGAGGTTAACITGAAATTGCTGGCTTATGGATGTTTCAAAGAGAGGTTCTGCTGTTGAGGTATAATGCAAGTACGGCTTGTGGTAGTTTACCAACTGCGCTAACTTTTTTACTGACCGTATTGAAACGTTATCGATAANAAAA
BB 4.CM NS8 (B33)	<i>Metschnikowia aff. fructicola</i>	100%	D1/D2 Domain	GGCAAAAGCTCAAATTGAAATCCCCGGATTGAAATTGAAAGAGATTGGCTCCGGCCGGCAGGGGTTAAGTCCACTGGAAAGTGCGCCACAGAGGGTGACAGCCCCGTGAACCCCTCAAGCCCTCATCCCAGATCTCAAAGTCGAGTTGGGAATGCACTGGGTGTAATTCTCAAAGCTAAATACCGGGAGAGACCGATAGCGAACAAAGTACAGTGTGAAAGATGAAAGCACTTGGAAAGAGAGTGAAAGAATGCACTGGGTGAAATTTGCTCAAAGGAAAGGGCTGCAAGCACACTAACCTTGAACGCACTGGCCAGCATCGGGCGGGAAAACAAACACCGGGGAATGTAACCTTCGAGGATTATAACCCGGTCTCAATTCTTGTGTTGCCCGAGGCCTGCAATCTAAGGATGCTGGCGTAATGGTGCAGTCGCCGTCTGAAAC
	<i>Metschnikowia pulcherrima</i>	99%	ITS	ACTCTAAATCTAACCTCTAAACTTATTAAAAAAACTTCAACAAACGATCTTGTGTTCTGCATCGATGAAGAACGCACTGGGATCTGAAACGCACATTGCGCCCGGGGTATCTCCAGGGCATGCGTGGTGAGCGATATTACTCTCAAACCTCCGGTTGGTCTGCTCGGCTTAATATCAACGGCGTAGAATAAAGTTAGCCCCATTCTTCTCACCTCGTAAGACTACCCGCTGAACTTAAGCATATCAATAA
BA 6.NM NS1 (B34)	<i>Saccharomyces cerevisiae</i>	99%	ITS	CTTATCGATAACGTTCCAATCGCTAGTATAAAAAAGATTAGCGCAATTGGTAAACCTAACGACCAGCTACTTCGATTACCTCAAGCACGCAAAAGAACCTCTTTGGAAAAAAACATCCAATGAAAAGGCCAGCAATTCAAGTTAACAGGATGCTCAAACAGGCATGCCCTGGAATACCAAGGGGCACATGTGCTCAAAGATGCTGATGATTACGGAATTCTGCAATTCAAATCGTATCCGCTTCAAGATGCTGTTCTCATCGATGCGAGAACCAAAAAAATCGTTGTTGAAAGTTTAAATTAAAAATTCCAGTTACAAAAATTCTGTTGACAAAAAATTAAATGAAATAAATAAAATTGTTGTTGTTAATTCTGGCCCGATTGCTGCAATGCCAAAGAAAAAGTTGCAAAGATATGAAAACCTCCACAGTGTGTTGATTGAAACGGTTTAATTGCTCTAAACAAAGCACAAAATCTCACCGTTGGAAATAGCAAGAAAAGAAACTTACAAGCCTAGCAAGACGGCGCACTTAAAGCGCAGGGCCGGCTGGACTCTCCATCTTGTCTTGTGCCCAGTAAAAGCTCTATGCTCTTGAAC

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
BA 6.NM NS2 (B35)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	GGCGAGTGAGCGGCAAAAGCTCAAATTGAAATCTGGTACCTTCGGTGC CCGAGTTGTAATTGGAGAGGGCAACTTGGGCCCTTGTCTATG TTCCTTGAACAGGACGTATAGAGGGTGAGAATCCGTGAGGAG AGTGCGGTTCTTGTAAAGTGCTCGAAGAGTCGAGTTGGGAAT GCAGCTAAGTGGTGGTAATTCATCTAAAGCTAAATTTGGCAG AGACCGATACGAACAAAGTACAGTGAGAAGATGAAAAGAACIT AAAAGAGAGTGAAGAAAGTACGTGAAATTGTTGAAAGGGAGGGATT GATCAGACATGGTTGCTGCCCCTGCTCTTGTGGTAGGGGAATC TCGCATTTCACTGGGCCAGCATCAGTTGGTGGCAGGATAATCCATA GGAATGTTAGCTGCCTCGTAAGTATTAGCTGTGGGAATACTGCCA GCTGGGACTGAGGACTGCACGTAAGTCAGGATGCTGGCATAATGGT TATATGCCCGCTTGA
	<i>Saccharomyces cf. cerevisiae/paradoxus</i>	100%	ITS	GTCAACTITAAGAACATTGTCGCCAGCGCTCTTATCGATAAC GTTCCAATACGCTCAGTATAAAAAAGATTAGCCGAGTTGGTAAAACC TAAAACGACCGTACTTGCAATTACCTCAAGCACGAGAGAAACCTCTC TTTGGAAAAAAACATCAAATGAAAAGGCCAGCAATTCAAGTAACT CCAAAGAGTACTACTCACTACCAACAGAATGTTGAGAAGGAATGA CGCTCAAACAGGCATGCCCTCGGAATACCAAGGGCGCAATGCGT TCAAAGATTGATGATTCAAGGAATTCTGCAATTCAAGTATCGC ATTCGCTGCTTCACTCGATGCGAGAACCAAGAGATCCGTTGA AAGTTTTAATTTAAATTTCAAGTACAGGAAAATTCTGTTGAC AAAAATTAAATGAAATGATAAAATTGTTGTTGTTACCTCTGGCC CCGATTGCTGAATGCCAAAGAAAAGTGCAGGATATGAAAATC CACAGTGTGTGTATTGAAACGGTTAATTGCTCTAACAAAAGCAC AGAAATCTCACCGTTGGATAGCAAGAAAGAAACTTACAAGCCTA GCAAGACCGCGACTTAAGCGCAGGCCGGCTGGACTCTCCATCTTG TCTCTGCCAGTAAAGCTCTATGCTTGTGCAAAACAAAAAAATC CATTTCAAAATTATTAATTTAATGATCCTCCCGAGG
BB 4.CM S10 (B36)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	GTGAAGCGGCAAAAGCTCAAATTGAAATCTGGTACCTTCGGGCCGA GTTGTAATTGGAGAGGGCAACTTGGGCCCTTGTCTATGTTCT TGGAACAGGACGTATAGAGGGTGAGAATCCGTGAGGAGTGC GGTTCTTGTAAAGTGCTCGAAGAGTCGAGTTGGGAATGAGC TCTAAGTGGTGTAAATTCACTAAAGCTAAATATTGGCGAGAGACC GATAGCGAACAGTACAGTGATGAAAGATGAAAAGAACITTTGAAAG AGAGTAAAAGTACGTGAAATTGTTGAAAGGGAGGGCATTTGATCA GACATGGTTTGTGCCCTGCTCTGTGGTAGGGGAATCTCGCAT TTCACTGGCCAGCATCAGTTGGTGGCAGGATAATCCATAGGAATG TAGCTGCTCGTAAGTATTAGCCTGTGGGAATACTGCCAGCTGG ACTGAGGACTGCGACGTAAGTCAGGATGCTGGCATAATGGTTATATGC CGCCCGTCTTGA
DA 0.CM NS2 (D2)	<i>Metschnikowia aff. pulcherrima</i>	99%	D1/D2 Domain	CGGCGAGTGAGCGGCAAAAGCTCAAATTGAAATCCCCGGGAATTG TAATTGAAAGAGATTGGTCCGGCCGGCAGGGGTTAAGTCCACTGGAA AGTGGCGCACAGAGGGTGACAGCCCCGTGAACCCCTCAACGCCCTC ATCCCAATCTCAAGAGTCGAGTTGGGAATGCAGCTCTAAAGTGG CTGGTAATTCATCTAAAGCTAAATACCGCGAGAGACCGATAGCGA ACAAGTACAGTGAAAGTGAAGAACACTTGAAGAGAGTGAAAG AAAGTACGTGAAATTGTTGAAAGGGAGGGCTTGAAGCAGACACTTA ACTGGGCCAGCATGGGGCGGGAGACAAAACCACCGGGGAATGTA CTTTGAGGATTATAACCCCGCCCTACTCCCATACCAACCCGAGGC CTGCAATCTAAAGGATGCTGGCGTAATGGTTGAAGTCGCCCCTTGAA ACACGG
	<i>Metschnikowia chrysoperlae</i>	100%	ITS	TAAACACTTACNTGAATTAAAACACANATTAAAAAATTAAAAA CCGGGTATCTTGGTTCTCATATNNGAAAAAAACACAGAAATGCGATA CCCNATATGACTTGGCACAGAAANATTNAATTTNACACACATT TGCCCCCGGGGTTCCCCGGGTGCGCGGGGAGCGATATTACTCT CAAAACCCCGGTTGGNCCGTGTTNGGCTAAAATCAACNGGGCTCTA AAAANTTANCCCCCTTTTCTCTCCCCCTAAACACCCCCCTG TACTTTAACATATAAAAACGGGGGG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
DA 0.CM NS3 (D3)	<i>Metschnikowia pulcherrima</i>	99%	D1/D2 Domain	GGCAAAAGCTCAAATTGAAATCCCCGGATTGAAAGAGATTTGCGCCACAG TTGGGTCCGCCGGCGGGGTTAAGTCCACTGGAAAGTGCGCCACAG AGGGTGACAGCCCCGTGAACCCCTCAACGCCCTCATCCAAATCTCCA AGAGTCGAGTTGTTGGGAATGCAGCTTAAGTGGGTGTAATTCCAT CTAAAGCTAAATACCGGGAGAGACCGATAGCGAACAGTACAGTGT GGAAAGATGAAAGCACTTGTAAAAGAGAGTGAAGAAAGTACGTGAAAT TGTGAAAGGGAAGGCTTGCAGCAGACACTTAACGGCCAGCATC GGGGCGCGGGAAAGCAAAACACCAGGGGAATGTACCTTCGAGGATTA TAACCCGGTCTTACTCCATGCCGCCCCGAGGCGTCAATCTAAGGA TGCTGGCGTAATGGTTGCAAGTCGCCGTCTGA
	<i>Metschnikowia sinensis</i>	94%	ITS	CTTTTAGGAAAAACCGAACCTTTTTATAAACACAATTAAAAAC TTTAACAACGGATCTTGGTCTCGCATCGATGAAAACGCAAG TTGCAGATCGTAATATGACTTGCAGACGCAGTGAATCTTGAAAC GCACATTGCGCCGGGGTATTCCCAGGGGATGCGTGGGGAGCGAT ATTACTCTAACCTCCGGTTGGGCTGCTTGGCTTAATATCAACG GGCTNTAATAAGTTAGCCCCATTCTTTCTCACCTCGTAAGACT ACCCGCTGAACCTAACATATCAATAAACG
DA 0.CM NS4 (D4)	<i>Metschnikowia pulcherrima</i>	99%	ITS	GGTGAGGAAAAGGAATGGGCTAAAACCTATTCTAGCGCCGTTGATA TTAGGCGAAGCAGGACCAACCAGGAGGTTGAGAGTAAATATCGCT CACCCACGCATGCCCTGGGAATACCCGGGGCGCAATGTGCGTCAA AGATTCAATGATTACGCTGCAAGTCATATTACGTATCGCAATTGCT GCGTCTTCATCGATGCGAGAACCAAGAGATCCGTTGAAAGTTTT T
DB 0.CM NS2 (D6)	<i>Metschnikowia chrysoperlae</i>	100%	ITS	GGGGCTAAAACCTATTCTAGCGCCGTTGATATTAGGCCGAAGCAGGACC AAACCGGAGGTTTGAGAGTAAATATCGCTCACCCACGCATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAGATTCAATGATTACGTC GCAAGTCATATTACGTATCGCAATTGCGTTCTCATCGATGCGAG AACCAAGAGATCCGTTGAAAGTTTTAATTGTTATTGAAAG AGAGTAAAAGTACGTGAAATTGTAAGGGAGGGCTGCAAGCA GACACTTAACCTGGCCAGCATCGGGCGGGAAACAAAACACCG GGAATGTACCTTCGAGGATTATAACCCGGCTTATTCCCTGCTGCC CCGAGGCCGTAATCTAAGGATGCTGGCTAATGGTTGCAAGTCGCCCG TCTTGAAACACGGAC
DB 0.CM NS4 (D8)	<i>Metschnikowia pulcherrima</i>	99%	D1/D2 Domain	CTCAGTAACGGCGAGTGAAGCGGCAAAGCTCAAATTGAAATCCCC GGGAATTGTAATTGAAAGAGATTGGTCCGCCGGCGAGGGGTTAAGTC CACTGGAAAGGTGGCAGACAGGGGTGACAGCCCCGTGAACCCCTCA ACGCCCTCATCCAGATCTCAAAGATCGAGTTGGGAATGCAGCT CTAAGTGGTGGTAATTCCATCTAAAGCTAAATACCGGGAGAGACC GATAGCGAACAGTACGTGATGAAAGATGAAAAGCACTTGTAAAAG AGAGTAAAAGTACGTGAAATTGTAAGGGAGGGCTGCAAGCA GACACTTAACCTGGCCAGCATCGGGCGGGAAACAAAACACCG GGAATGTACCTTCGAGGATTATAACCCGGCTTATTCCCTGCTGCC CCGAGGCCGTAATCTAAGGATGCTGGCTAATGGTTGCAAGTCGCCCG TCTTGAAACACGGAC
	<i>Metschnikowia chrysoperlae</i>	95%	ITS	GGGGCTAAAACCTATTCTAGCGCCGTTGATATTAGGCCGAAGCAGGACC AAACCGGAGGTTTGAGAGTAAATATCGCTCACCCACGCATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAGATTCAATGATTACGTC GCAAGTCATATTACGTATCGCAATTGCGTTCTCATCGATGCGAG AACCAAGAGATCCGTTGAAAGTTTTAATTGTTATTGAAAG AAAATGATAAGTGTATTGCTAAAAGTGTGTAAGTGTATTTAGAGA TCCCTCCGCAAGGCTCACATAGAGAAAAGGAGAATTAGTAAAAAATTTT CGCCTTTGGGGAAAAGTGAATTTTATTATCCCAAACAAATCAATT AACATAACCAACCGATAGGTTGTTCTGCATCAATAAACAGCAACGA AGTGGAGATACTTAGTATGACTTACAGACGTGAATCATCAATTGAA GCGATTGCGCCCTATGTATTCCCGGCATGCGTAGTGAGCGATATT CTCTCAACGCTCTGGTCTGCTCCGCTAATATCACCTCGCTGAAAT ATTATCTAGCCCTTCTCTCCCTAGTATAACTATCACTTAACCTCA

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
DA 4.CM NS2 (D10)	<i>Stamerella bacillaris</i>	100%	D1/D2 Domain	GGCGAGTGAACAGGAAGAGCTCAGATTGAAAGGCACTTTGTGCCG TTGTATTCTGAAGTTAGGGTCTGAGAAACGATGCTTAAGTCCTCTGGA AAGGAGCGCCATGGAGGGTGATAGCCCCGTCTAGCATTGACCTCATATA GGATCTAACATGGAGTCAGTTGTTGGGAATGCAGCTCAAATGGGTG GTATGCTCCATCTAAAGCTAAATATCTCGAGAGACCGATAGTAAACAA GTACTGTGAGGGAAAGATGAAAAGAACTTGTAAAAGAGAGTGAAGAAA GTACGTGAAATTGTTGAAATGGAAGGGTAGGCCGCTAACCATGTAGAG CCGTGTTGGGGGAAGATAAATGCTGTAGAATGAGTCCTCGAGTA TTAGATGCAGTTCATATTCCCACCCAGCGCAGGATCTCAGGTTCT ACTAAATGGTGTCTACCACCGTCTGTA
		100%	ITS	GAGTTAATTAACGTGTCACTCCATGATTTAACTCTAAAGGAGCA AGACCTCATAACAAAATGGTGAGAGGAAGATTATCACTCAACAAG CATGCTATTAGACAAACTAATAGCGCAATATGCGTCAAAAATTCAATG ACTCACGTCGCAATTGCGATTACCTATCGCGCTTGTGCGTTCTCAT CGATACGAGAACAGAGATCGGTGAAAGTTAAATTTCATTG TTTCAGAACAAATAAAAGTTAAAGATTGGGCCCTTCGACACCC AAGCAATTGCAAAGACCGAAGTCTAACGTTGACAGTGGTTGGCAA AAAGCCTCAGAAATGATCCTCCGAGGT
DA 4.CM NS3 (D11)	<i>Metschnikowia pulcherrima</i>	99%	ITS	TGGGGCTAAAACCTATTCTAGGCCGTTGATATTAGGCCGAAGCAGGAC CAAACCGGAGGTTGAGAGTAATATCGCTCACCCACCGATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAAGATTCAATGATTACGTCT GCAAGTCATATTACGTATCGCAATTGCGTGTGCGTTCTCATCGATGCGAG AACCAAGAGATCGGTGAAAGTTAAATTGTTATTGACGGTTA AGATTAGAGTTGTGCTAAAGGGTGTAAAACAAATTAAATGATCC TICCGCAGGTT
DA 4.CM NS4 (D12)	<i>Metschnikowia pulcherrima</i>	96%	ITS	TGGGGCTAAAACCTATTCTAGGCCGTTGATATTAGGCCGAAGCAGGAC CAAACCGGAGGTTGAGAGTAATATCGCTCACCCACCGATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAAGATTCAATGATTACGTCT GCAAGTCATATTACGTATCGCAATTGCGTGTGCGTTCTCATCGATGCGAG AACCAAGAGATCGGTGAAAGTTAAATTGTTATTGAGGGTT AAGATTAGAGTTGTGCTAAAGGGTGTAAAACAAATTAAATGATCC CTTCCGAGGTTAACCTACGGAA
DB 4.CM NS1 (D13)	<i>Metschnikowia pulcherrima</i>	99%	ITS	TGGGGCTAAAACCTATTCTAGGCCGTTGATATTAGGCCGAAGCAGGAC CAAACCGGAGGTTGAGAGTAATATCGCTCACCCACCGATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAAGATTCAATGATTACGTCT GCAAGTCATATTACGTATCGCAATTGCGTGTGCGTTCTCATCGATGCGAG AACCAAGAGATCGGTGAAAGTTAAATTGTTATTGAGGGTT AAGATTAGAGTTGTGCTAAAGGGTGTAAAACAAATTAAATGATCC CTTCCGAGGTTAACCTACGGAA
DB 4.CM NS2 (D14)	<i>Metschnikowia pulcherrima</i>	99%	ITS	GGGGCTAAAACCTATTCTAGGCCGTTGATATTAGGCCGAAGCAGGAC AAACCGGAGGTTGAGAGTAATATCGCTCACCCACCGATGCCCTGGG GAATACCCGGGGCGCAATGTGCGTTCAAAGATTCAATGATTACGTCT GCAAGTCATATTACGTATCGCAATTGCGTGTGCGTTCTCATCGATGCGAG AACCAAGAGATCGGTGAAAGTTAAATTGTTATTGAGGGTT AAGATTAGAGTTGTGCTAAAGGGTGTAAAACAAATTAAATGATCC CTTCCGAGGTTAACCTACGGAA
DB 4.CM NS3 (D15)	<i>Metschnikowia fructicola</i>	99%	D1/D2 Domain	GTACGGCGAGTGAAGCGCAAAAGCTCAAATTGAAATCCCCGGGAA TTGTAATTGAAGAGATTGGTCCGGCGCAGGGTTAAGTCACGT GAAAGTGGCGCCACAGAGGTGACAGCCCGTGAAACCCCTCAACGCC CTCATCCCAGATCTCAAGAGTCAGTTGGGAATGCAGCTCAAG TGGGTGTTAAATTCCATCTAAAGCTAAATACCGCGAGAGACCGATAG CGAACAGTACAGTGTGAAAGATGAAAAGCACTTGTAAAAGAGAGT GAAAAGTACGTGAAATTGTTGAAAGGGCTTGTGCAAGCAAC TTAAGTGGCCAGCATGGGGCGCGGAAACAAAACACCGGGAAAT GTACCTTCGAGGATTATAACCCGGTCTCAATTCTGTTGCCAG CCCTGCAATTAGGATGCTGGCTAATGGTTGCAAGTCGCCGCTTG AACAAACGGAC
	<i>Metschnikowia aff. chrysoperiae</i>	97%	ITS	ATGGGGCTAAAACCTATTCTAGGCCGTTGATATTAGGCCGAAGCAGGA CCAAACCGAGGAGGTTGAGAGTAATATCGCTCACCCACCGATGCCCTG GGGAATACCCGGGGCGCAATGTGCGTTCAAAGATTCAATGATTACGT CTGCAAGTCATATTACGTATCGCAATTGCGTGTGCGTTCTCATCGATGCGA AACCAAGAGATCGGTGAAAGTTAAATTGGGTTATTGAAAAAA TAAATGATAAGTGTGTTCCCTAAAGTG

Table A.1. (continued)

Name	BLAST Result	Similarity	Region	Sequence
DA 4.NM S1 (D16)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGCGAGTGAAGCGGAAAGCTCAAATTGAAATCTGGTACCTTCGGT GCCGAGTTGAATTGGAGAGGGCAACTTGGGGCGTCCCTGTCTA TGTTCCCTTGAACAGGACGTATAGAGGGTGAGAATCCGTGCGA GGAGTGCCTTCTTGAAAGTCCTCGAAGAGTCGAGTTGGGA ATGCAGCTCTAAGTGGGTGTAATTCCATCTAAAGCTAAATATTGGC AGAGACCGATAAGCGAACAGTACAGTGATGGAAAGATGAAAAGACTT TGAAAAGAGAGTAAAAAGTACGTGAAATTGTTGAAAGGGAAAGGGCAT TTGATCAGACATGTTGCCCCCTGCTCCTTGTGGTAGGGAA CTCGCATTTCAGTGGGCCAGCATCAGTTGGTGGCAGGATAATCCAT AGGAATGTAGCTGCCTCGTAAGTATTATAGCTGTGGGAATACTGCC AGCTGGGACTGAGGACTGCGACGTAAGTCAAGGATGCTGGCATAATGG TTATATGCCCG
DA 4.CM S3 (D17)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	GGCGAGTGAAGCGGAAAGCTCAAATTGAAATCTGGTACCTTCGGT GCCGAGTTGAATTGGAGAGGGCAACTTGGGGCGTCCCTGTCTA TGTTCCCTTGAACAGGACGTATAGAGGGTGAGAATCCGTGCGA GGAGTGCCTTCTTGAAAGTCCTCGAAGAGTCGAGTTGGGA ATGCAGCTCTAAGTGGGTGTAATTCCATCTAAAGCTAAATATTGGC AGAGACCGATAAGCGAACAGTACAGTGATGGAAAGATGAAAAGACTT TGAAAAGAGAGTAAAAAGTACGTGAAATTGTTGAAAGGGAAAGGGCAT TTGATCAGACATGTTGCCCCCTGCTCCTTGTGGTAGGGAA CTCGCATTTCAGTGGGCCAGCATCAGTTGGTGGCAGGATAATCCAT AGGAATGTAGCTGCCTCGTAAGTATTATAGCTGTGGGAATACTGCC AGCTGGGACTGAGGACTGCGACGTAAGTCAAGGATGCTGGCATAATGG TTATATGCCCG
DA 4.CM S1 (D18)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CGGCAAAAGCTCAAATTGAAATCTGGTACCTTCGGTGCCCCGAGTTGAA TTGGAGAGGGCAACTTGGGGCGTCCCTGTCTATGTTCTTGGAAAC AGGACGTATAGAGGGTGAAGATCCGTGCGAGGAGTGCCT TTGTAAGTGCCTCGAAGAGTCGAGTTGGGAATGCA GGCTCAGTAAAGCTAAATATTGGCAGAGACCGATAGC GAACAAAGTACAGTGATGGAAAGATGAAAAGACTTGA AAAAGACTGTAAGTGAATTGTTGAAAGGGCAT TTGATCAGACATGTTGCCCCCTGCTCCTTGTGGTAGGGAA CTCGCAGCATCAGTTGGTGGCAGGATAATCCATAGGAATGTAGCTT GCCCTCGTAAGTATTATAGCTGTGGGAATACTGCCAGCTGGGACTGAG GACTGCGACGTAAGTCAAGGATGCTGGCATAATGGTTATATGCCCG TCTTGAAACC
DA 4.CM S8 (D19)	<i>Saccharomyces cerevisiae</i>	100%	D1/D2 Domain	CTCAAATTGAAATCTGGTACCTTCGGTGCCCCGAGTTGTAATTGGAGA GGGCAACTTGGGGCGTCCCTGTCTATGTTCTTGGAAACAGGACGTC ATAGAGGGTGAAGAATCCGTGCGAGGAGTGCCT GCCCTCGAAGAGTCGAGTTGGGAATGCA GCTCTAAGTGGGTGA AATTCCATCTAAAGCTAAATATTGGCAGAGACCGATAGC GAACAAAGTACAGTGAAAGATGAAAAGACTTGA ACAGTGATGGAAAGATGAAAAGACTTGA ACGTTGAATTGTTGAAAGGGCATTGATCAGACATGTT TGCCCTCTGCTCCTTGTGGTAGGGGAATTCG CATCAGTTGGTGGCAGGATAATCCATAGGAATGTAGCTTG TAAGTATTATAGCTGTGGGAATACTGCCAGCTGGGACTGAG ACGTAAGTCAAGGATGCTGGCATAATGGTTATATGCCCG TCTTGAAACC
Commercial <i>Saccharomyces cerevisiae</i>		100%	D1/D2 Domain	GGCGAGTGAAGCGGAAAGCTCAAATTGAAATCTGGTACCTTCGGT GCCGAGTTGAATTGGAGAGGGCAACTTGGGGCGTCCCTGTCTA TGTTCCCTTGAACAGGACGTATAGAGGGTGAGAATCCGTGCGA GGAGTGCCTTCTTGAAAGTCCTCGAAGAGTCGAGTTGGGA ATGCAGCTCTAAGTGGGTGTAATTCCATCTAAAGCTAAATATTGGC AGAGACCGATAAGCGAACAGTACAGTGATGGAAAGATGAAAAGACTT TGAAAAGAGAGTAAAAAGTACGTGAAATTGTTGAAAGGGAAAGGGCAT TTGATCAGACATGTTGCCCCCTGCTCCTTGTGGTAGGGAA CTCGCATTTCAGTGGGCCAGCATCAGTTGGTGGCAGGATAATCCAT AGGAATGTAGCTGCCTCGTAAGTATTATAGCTGTGGGAATACTGCC AGCTGGGACTGAGGACTGCGACGTAAGTCAAGGATGCTGGCATAATGG TTATATGCCCG
<i>Metschnikowia pulcherrima</i> (DSM:70336)		100%	D1/D2 Domain	GGCGAGTGAAGCGGAAAGCTCAAATTGAAATCCCCGGGAATTG AATTGAAAGAGATTGGGTCCGGCGGGGGTTAAGTCCACTGGAA AGTGGCGCCACAGAGGGTACAGGCCGTGAACCCCTCAACGCCCTC ATCCCAGATCTCAAGAGTCGAGTTGGGAATGCA GTGTTAAATTCCATCTAAAGCTAAATACCGGGCAGAGACCGATAGCGA ACAAGTACAGTGATGGAAAGATGAAAAGACTTGA AAAGTACGTGAAATTGTTGAAAGGGAAAGGGCTGCAAGCAGACACTTA ACTGGGCCAGCATGGGGCGGGGGAGCAAAACCCGGGGAAATGTA CCTTCGAGGATTATAACCCGGCTTACTCCCATACCAACCCGAGGC CTGCAATCTAAGGATGCTGGCGTAATGGTGCAAGTCGCCGTCTGAA C

B. Phylogenetic Trees According to UPGMA Method

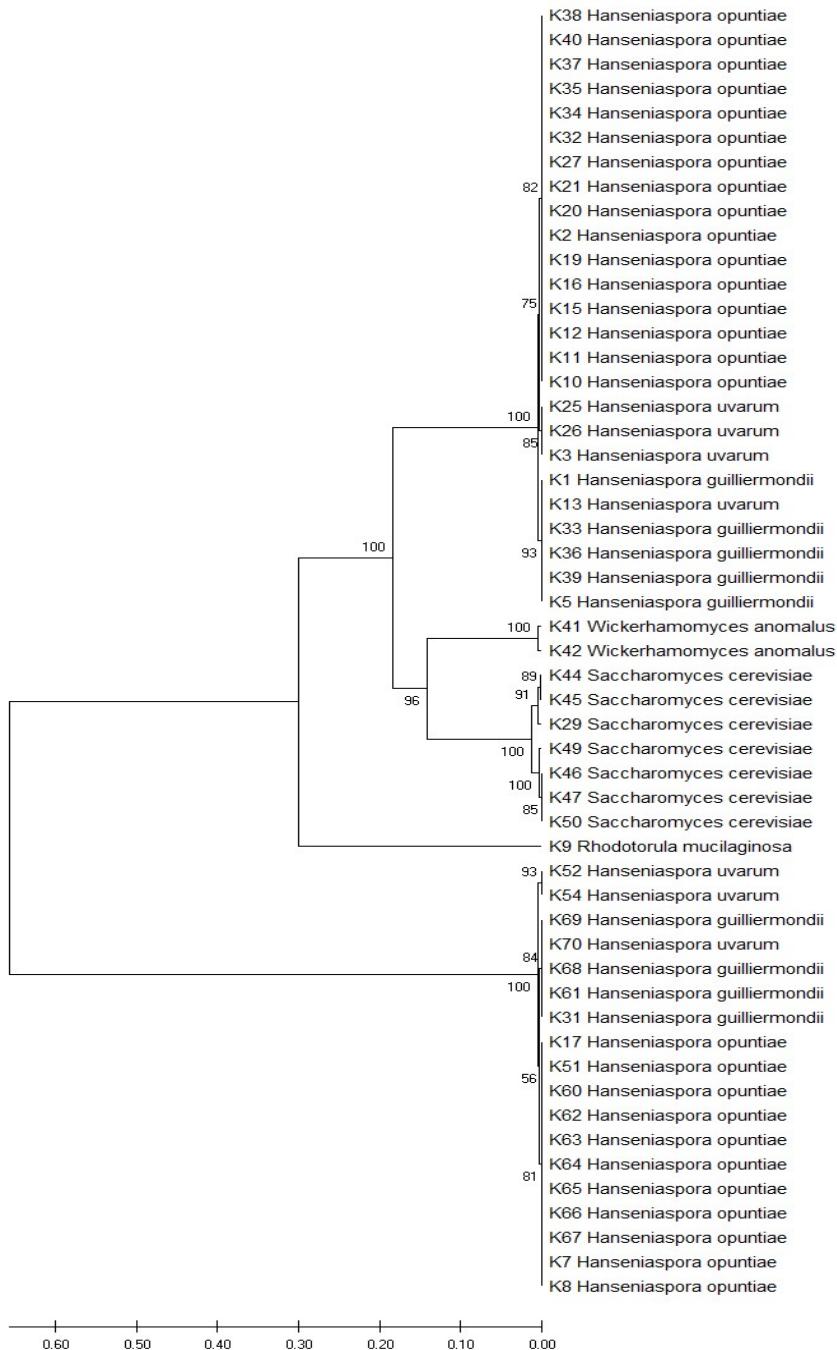


Figure B.1. Phylogenetic Tree with UPGMA Method of Non-Saccharomyces and Saccharomyces in Kalecik Karası Must and Wine with respect to ITS Region

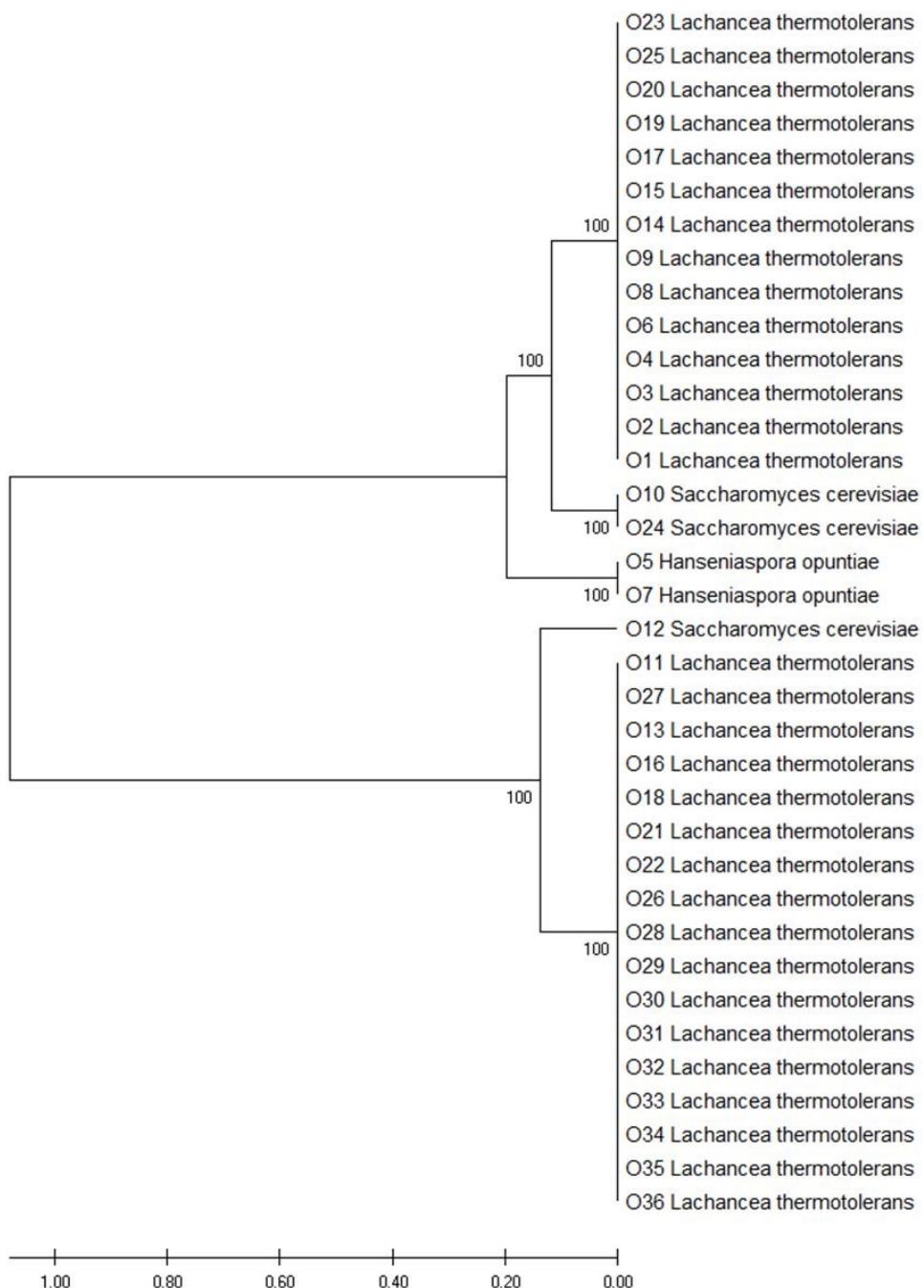


Figure B.2. Phylogenetic Tree with UPGMA Method of Non-*Saccharomyces* and *Saccharomyces* in Öküzgözü Must and Wine with respect to ITS Region

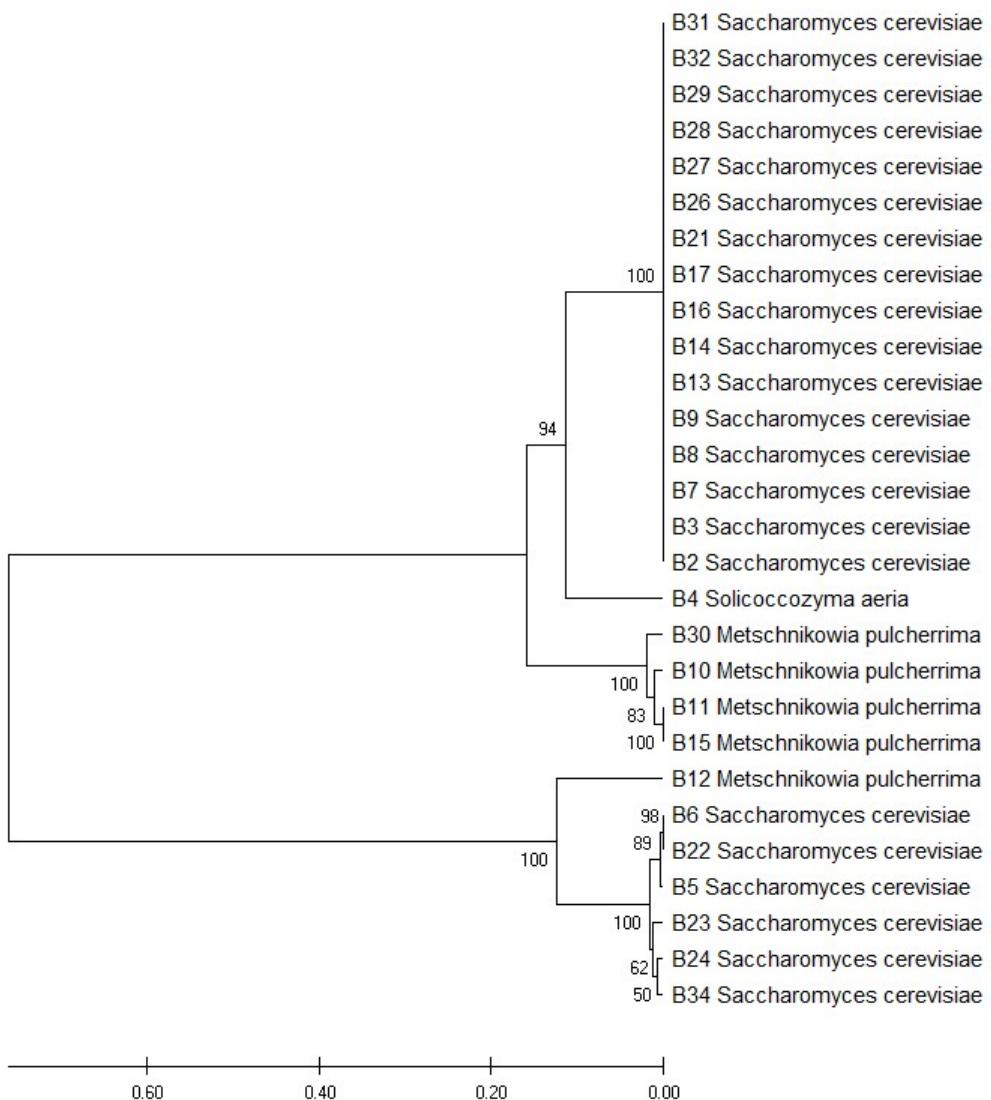


Figure B.3. Phylogenetic Tree with UPGMA Method of Non-Saccharomyces and Saccharomyces in Boğazkere Must and Wine with respect to ITS Region

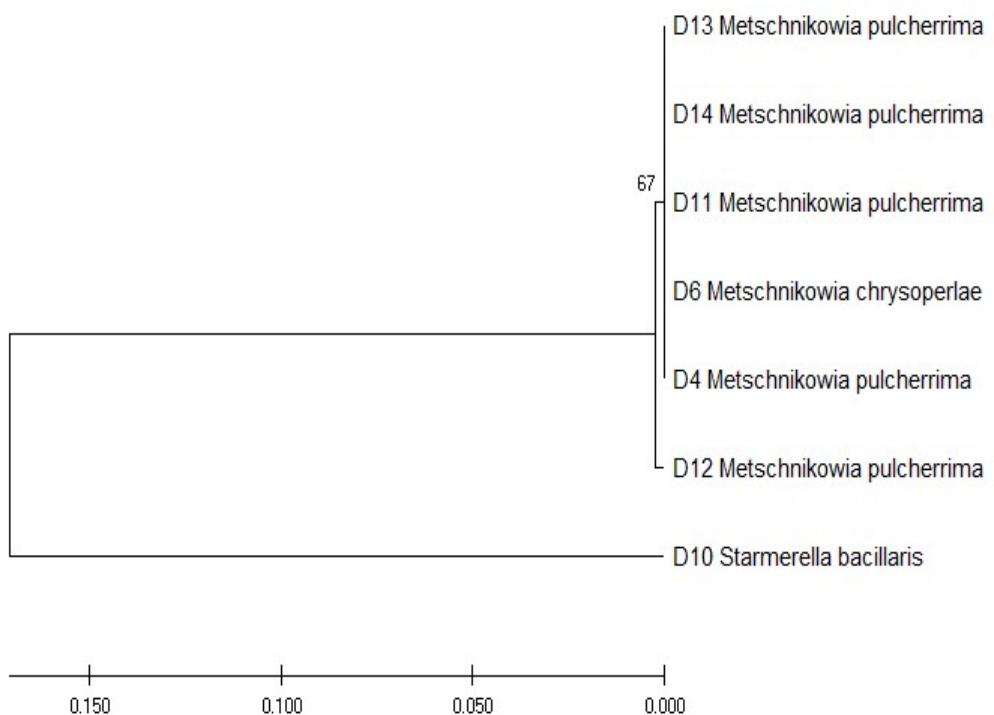
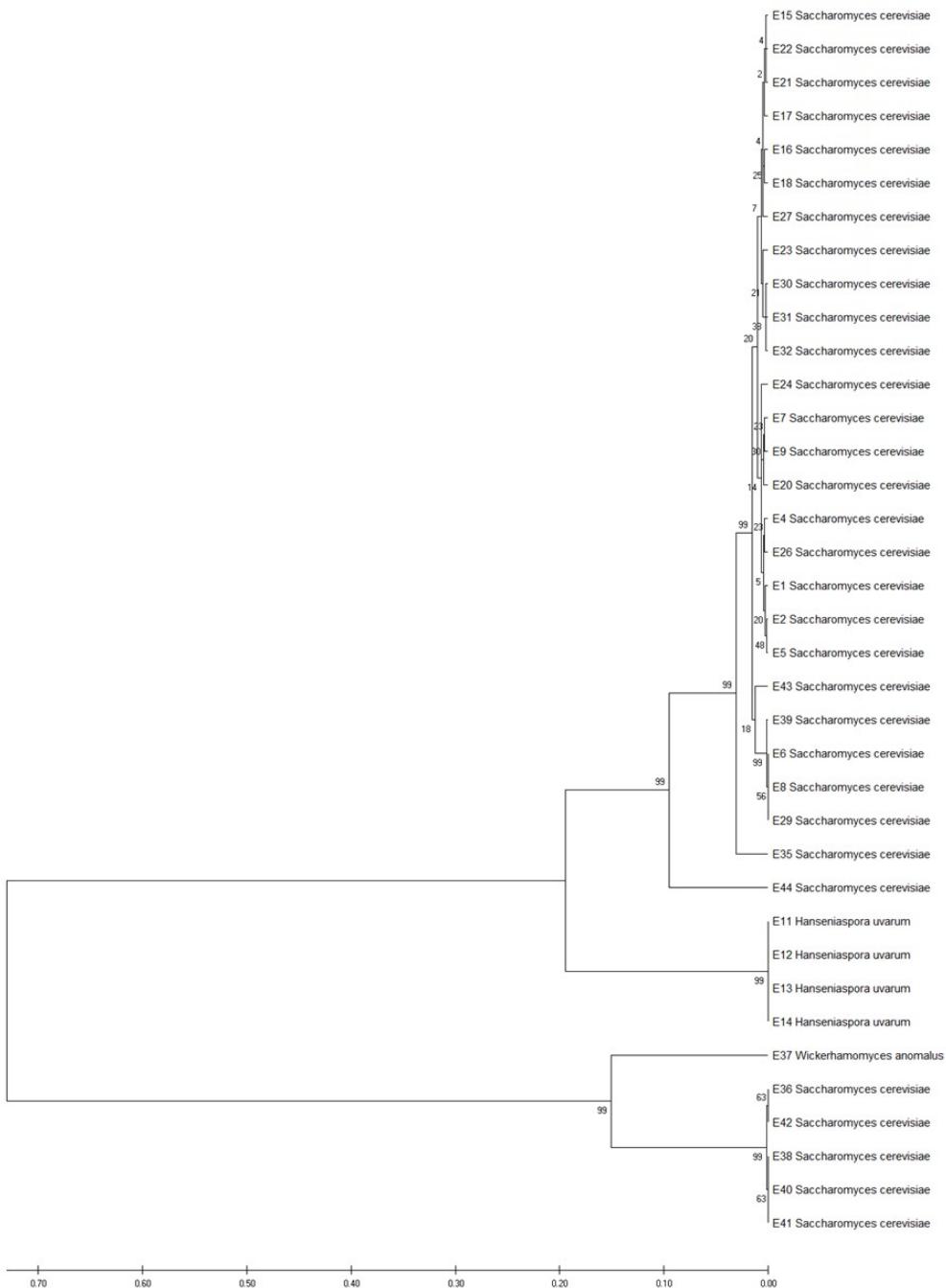


Figure B.4. Phylogenetic Tree with UPGMA Method of Non-*Saccharomyces* and *Saccharomyces* in Dimrit Must and Wine with respect to ITS Region



*Figure B.5. Phylogenetic Tree with UPGMA Method of Non-*Saccharomyces* and *Saccharomyces* in Emir Must and Wine with respect to ITS Region*

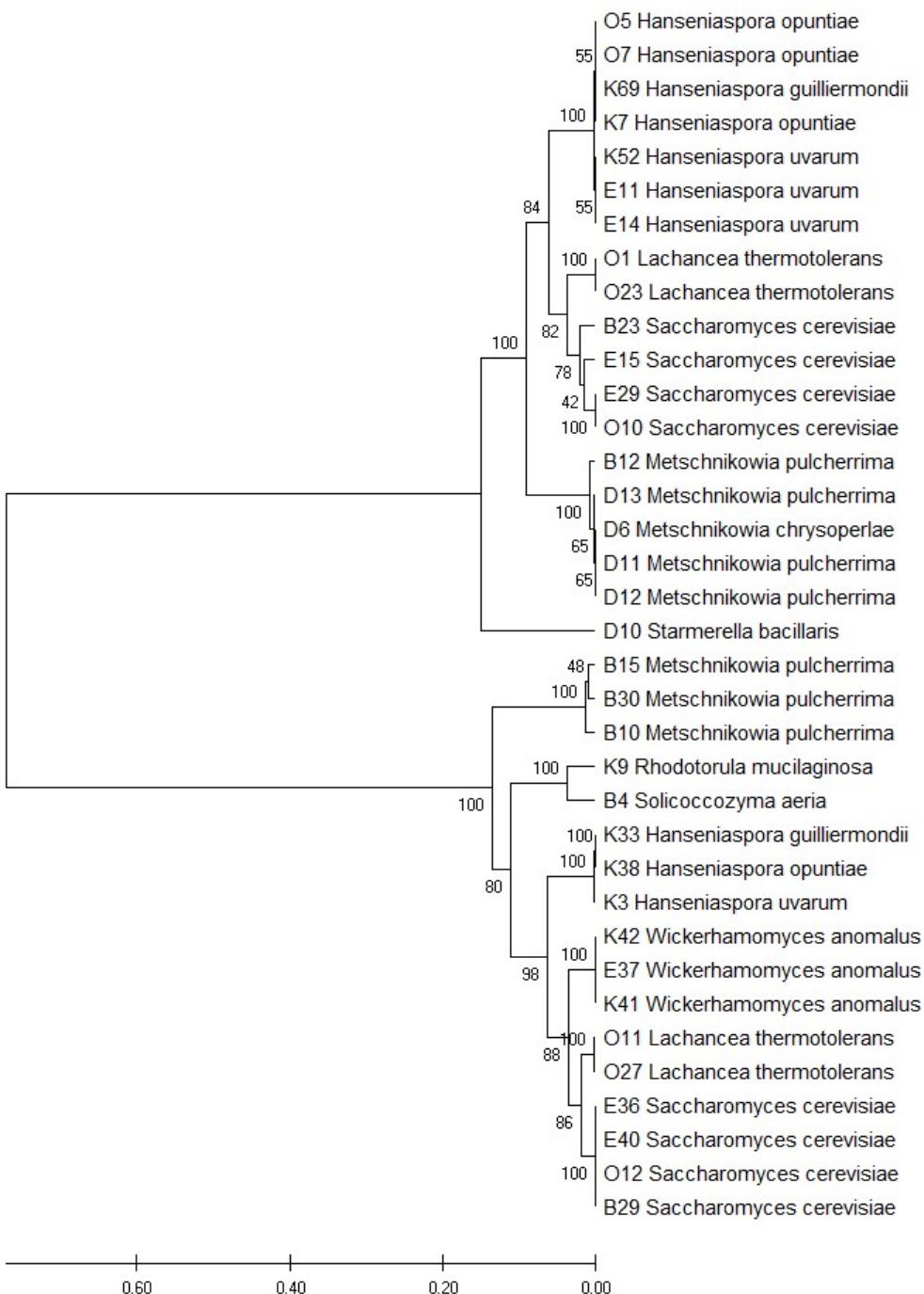


Figure B.6. Phylogenetic Tree with UPGMA Method of Selected Non-Saccharomyces and Saccharomyces in Five Different Grapes' Musts and Wines with respect to ITS Region

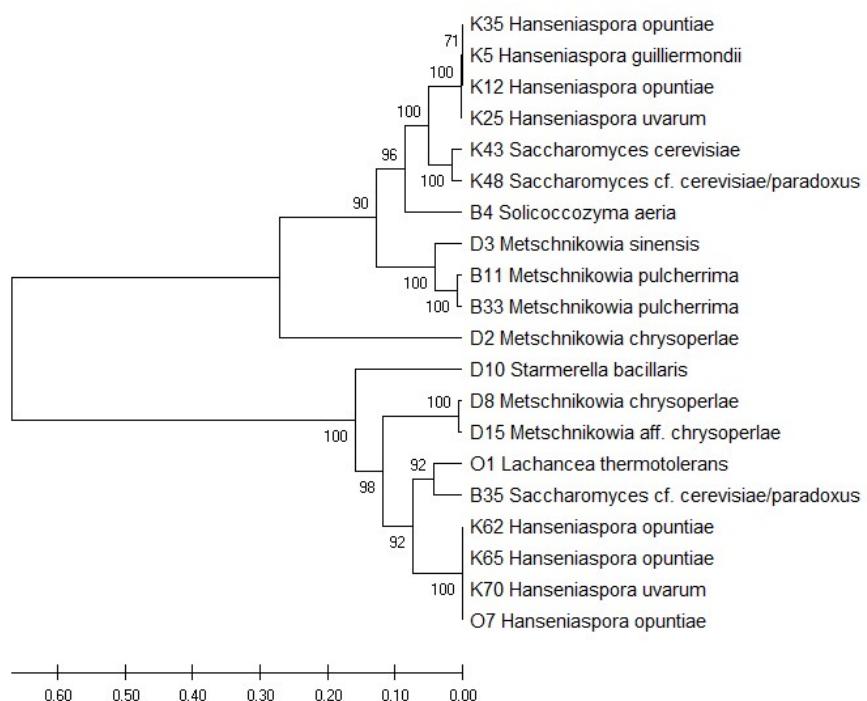


Figure B.7. Phylogenetic Tree with UPGMA Method Some Yeasts with Respect to ITS Region

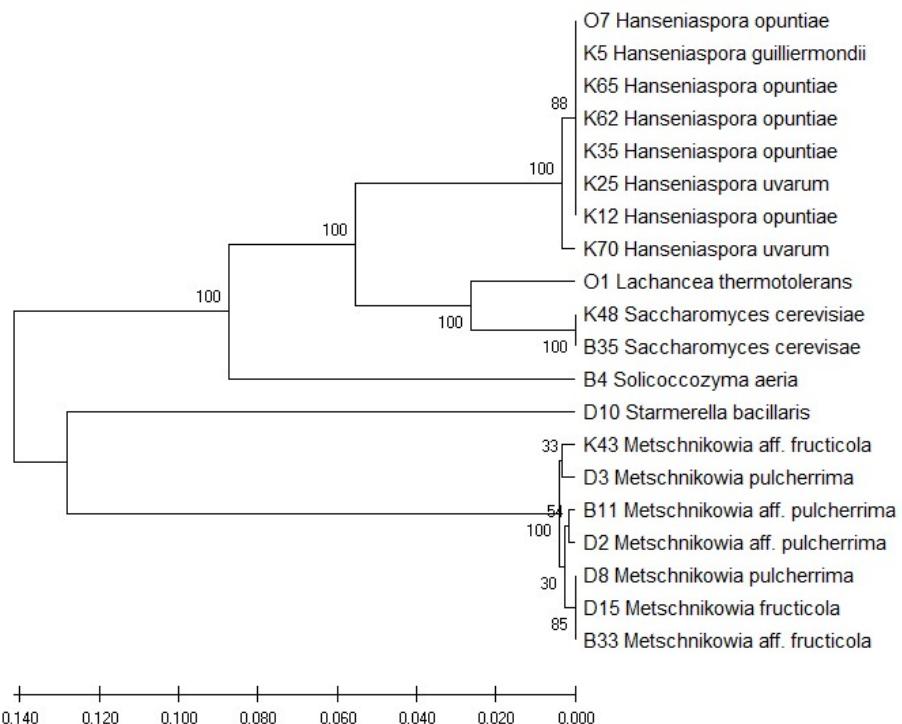


Figure B.8. Phylogenetic Tree with UPGMA Method Some Yeasts with Respect to D1/D2 Domain

C. Phenotypic Properties of All Isolated *Saccharomyces* Yeasts

Table C.1. The Results of Carbohydrate Fermentation Test and Phenotypic Properties of *Saccharomyces* Strains from Isolated in Must and Wine of Kalecik Karası

Name	Alcohol Tolerance (%)			SO ₂ Tolerance (mg/L)		Temperature Tolerance (°C)			Carbohydrate Fermentation Test						<i>H₂S</i> Product	Identification				
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose			
	ITS	D1/D2																		
KA 0CM S2	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	ng	nd	nd
KA 0CM S4	+	-	-	+	+	+	+	+	+	-	w	w	-	w	w	w	w	5	nd	nd
KA 4CM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd
KA 4CM S3	+	w	w	+	+	+	+	+	+	-	-	-	-	w	-	w	-	ng	nd	nd
KA 2NM S1	+	+	+	+	+	+	+	+	+	-	-	-	-	-	+	w	-	4	nd	nd
KA 2NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd
KA 2NM S3	+	+	+	+	+	+	+	+	+	-	+	+	+	-	+	-	w	3	nd	nd
KA 2NM S4	w	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	w	4	nd	nd
KA 2NM S5	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd
KA 2NM S6	+	+	+															4	+	nd
KA 2NM S7	+	+	+	+	+	+	+	+	+	w	+	+	+	+	+	-	-	4	nd	nd
KA 2NM S8	+	W	w	+	+	+	+	+	+	-	-	+	+	+	+	-	-	4	nd	nd
KA 2NM S9	+	w	w	+	+	+	+	+	w	-	-	-	-	+	+	-	-	3	nd	nd
KA 2NM S10	+	+	+	+	+	+	+	+	+	-	+	+	-	+	+	-	-	4	nd	nd
KA 4NM S1	+	-	-	+	+	+	+											2	nd	nd
KA 4NM S2	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	+
KA 4NM S3	+	-	-	+	+	+	+											1	nd	nd
KA 4NM S4	+	-	-	+	+	+	+											1	+	nd
KA 4NM S5	+	-	-	+	+	+	+											3	nd	nd
KA 4NM S6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd
KA 4NM S7	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd
KA 4NM S8	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd
KA 4NM S9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd
KA 4NMS10	+	+	+	+	+	+	+	+	+	-	+	w	+	+	+	-	-	4	nd	nd
KA 18NM S1	-	-	-	w	w	-	-	w	+	-	+	-	-	-	+	-	-	3	nd	nd
KA 18NM S2	-	-	-	+	+	+	+	+	+	-	-	-	-	+	+	-	-	2	nd	nd
KA 18NM S3	-	-	-	+	+	+	+	-	+	-	-	-	-	+	+	-	-	4	nd	nd
KA 18NM S4	-	-	-	+	+	+	+	+	+	-	w	w	w	-	+	-	-	3	nd	nd
KA 18NM S5	w	w	w	+	+	+	+	+	+	-	-	w	-	+	w	-	-	3	nd	nd
KA 32NM S1	w	w	w	+	+	+	+	+	+	-	+	+	+	-	+	-	-	4	nd	nd
KA 32NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	1	nd	+
KA 32NM S3	+	+	w	+	+	+	+	+	+	-	+	+	+	-	+	-	-	4	nd	nd
KA 32NM S4	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	w	4	nd	nd	
KA 32NM S5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd
KB 4CM S5	+	+	+															3	+	nd
KB 2NM S1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd
KB 2NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd
KB 2NM S3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	4	nd	nd	

Table C.1 (continued)

Name	Alcohol Tolerance (%)						Temperature Tolerance (°C)							pH Tolerance			Carbohydrate Fermentation Test			H ₂ S Product		Identification	
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2				
KB 2NM S4	+	-	-	+	+	+	+	+	+	w	+	+	+	+	+	-	-	1	+	nd			
KB 2NM S5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd			
KB 2NM S6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd			
KB 2NM S7	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd			
KB 2NM S8	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd			
KB 2NM S9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd			
KB 2NM S10	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd			
KB 2NM NS8	-	-	-	-	-	-	-	-	-	w	-	-	-	+	+	-	-	1	+	nd			
KB 4NM S1	+	-	-	+	+	+	+	+	+									3	nd	nd			
KB 4NM S2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	nd			
KB 4NM S3	+	+	-	+	+	+	+	+	+									3	nd	nd			
KB 4NM S4	+	-	-	+	+	+	+	+	+									2	nd	nd			
KB 4NM S5	+	-	-	+	+	+	+	+	+									3	nd	nd			
KB 4NM S6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd			
KB 4NM S7	+	+	+	+	+	+	+	+	+	w	w	+	+	+	+	w	-	3	nd	nd			
KB 4NM S8	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	+	3	nd	nd			
KB 4NM S9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd			
KB 4NM S10	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd			
KB 18NM S2																		3	nd	nd			
KB 18NM S1	-	-	-	w	w	w	+	-	+	-	-	-	-	-	-	+	-	2	nd	nd			
KB 18NM S3	+	+	+	+	+	+	+	+	+	-	w	w	-	-	-	+	-	2	nd	nd			
KB 18NM S4	-	-	-	+	w	+	+	+	+	-	w	-	-	-	-	+	-	2	nd	nd			
KB 18NM S5	+	w	w	+	+	+	+	+	+	-	w	+	w	-	w	-	-	3	nd	nd			
KB 32NM S1	+	+	+	+	+	+	+	+	+	w	+	+	+	+	+	w	-	4	nd	nd			
KB 32NM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	-	+	w	-	3	nd	nd			
KB 32NM S3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd			
KB 32NM S4	+	+	+	+	+	+	+	+	+	-	+	+	w	+	+	-	-	4	nd	nd			
KB 32NM S5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	4	nd	nd				
K13 (C)*	+	+	-	+	+	+	+	+	+	-				+	+	-	-	4	+	nd			
K15(C)*	+	+	+	+	+	+	+	+	+	-				+	+	-	-	4	+	nd			
K16(C)*	+	+	-	+	+	+	+	+	+	-				-	+	-	-	4	+	nd			

KA: A Parallel of Wine Made from Kalecik Karası Grape , KB: B Parallel of Wine Made from Kalecik Karası Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 2NM: 2. Day of Normal Maceration, 4NM: 4. Day of Normal Maceration, 18NM: 18. Day of Normal Maceration (Middle of the fermentation), 32NM: 32. Day of Normal Maceration (End of the fermentation); S: *Saccharomyces*; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; C*: Isolated by Çağrı Çavdaroglu; nd: Not determined.

Table C.2. The Results of Carbohydrate Fermentation Test and Phenotypic Properties of *Saccharomyces* Strains from Isolated in Must and Wine of Öküzgözü

Name	Alcohol Tolerance (%)			SO ₂ Tolerance (mg/L)			Temperature Tolerance (°C)			pH Tolerance			Carbohydrate Fermentation Test				H ₂ S Product	Identification		
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2	
OA 0CM S1	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd
OA 0CM S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OA 0CM S3																		4	nd	nd
OA 0CM S4	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OA 0CM S5	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	5	nd	nd
OA 0CM S7	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	5	nd	nd
OA 0CM S8	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OA 0CM S9	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd
OA 0CM S10	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd
OA 0CM NS3	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd
OA 4CM S1	+	w	w	+	+	+	+											4	nd	nd
OA 4CM S2	+	w	-	+	+	+	+											4	nd	nd
OA 4CM S4	w	-	-	+	+	+	+											4	nd	nd
OA 4CM S5	+	-	-	+	+	+	+											4	nd	nd
OA 4CM S6																		4	nd	nd
OA 4CM S7																		4	nd	nd
OA 4CM S8	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OA 4CM S9																		4	nd	nd
OA 4CM NS5	+	+	+															3	+	nd
OB 0CM S1	w	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	+	4	nd	nd
OB 0CM S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OB 0CM S3	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OB 0CM S5	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd
OB 0CM S6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	+
OB 0CM S7																		4	nd	nd
OB 0CM S8	+	+	+	+	+	+	+	+	+	-	-	-	-	-	-	-	-	4	nd	nd
OB 0CM S9	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OB 0CM S10	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd
OB 4CM S1*	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	+
OB 4CM S2	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	+
OB 4CM S3	-	-	-	+	+	+	+											3	nd	nd
OB 4CM S4	+	-	-	+	+	+	+											4	nd	nd
OB 4CM S5	+	w	w	+	+	+	+											4	nd	nd
OB 4CM S6																		4	nd	nd
OB 4CM S7																		4	nd	nd
OB 4CM S9																		4	nd	nd
OB 4CM S10																		4	nd	nd
OB 4CM NS5	+	+	+															4	+	nd

OA: A Parallel of Wine Made from Öküzgözü Grape , OB: B Parallel of Wine Made from Öküzgözü Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration; S: *Saccharomyces*; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black,; nd: not determined; *: the strain using for wine-making.

Table C.3. The Results of Carbohydrate Fermentation Test and Phenotypic Properties of *Saccharomyces* Strains from Isolated in Must and Wine of Boğazkere

Name	Alcohol				SO ₂ Tolerance				Temperature			pH			Carbohydrate Fermentation Test				H ₂ S Product		Identification	
	Tolerance (%)				(mg/L)				Toleransi (°C)			Tolerance										
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2			
BA 0CM S3	-	-	-	+	+	+	+	+	+	-	+	+	+	w	w	-	w	ng	nd	nd	nd	
BA 0CM S4	+	+	+	+	+	+	+	+	+	-	+	+	+	+	w	-	-	4	nd	nd	nd	
BA 0CM S5	+	+	w	+	+	+	+	+	+	-	+	+	-	+	+	-	-	3	nd	nd	nd	
BA 0CM S7	+	w	-	+	+	+	+	+	+	-	+	+	+	+	w	-	-	3	nd	nd	nd	
BA 0CM S8	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	4	nd	nd	nd	
BA 0CM S9																			3	nd	nd	nd
BA 0CM S10	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd	nd	
BA 0CM NS2	+	+	+															4	+	nd	nd	nd
BA 0CM NS3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	nd	nd	nd
BA 0CM NS5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	nd	nd	nd
BA 0CM NS6	+	+	+															4	+	nd	nd	nd
BA 0CM NS7	+	+	+															4	+	nd	nd	nd
BA 0CM NS8	+	+	+															4	+	nd	nd	nd
BA 4CM S2	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	nd	nd
BA 4CM S5	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	3	nd	nd	nd	nd
BA 4CM S6	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	+	w	4	nd	nd	nd	nd
BA 4CM S7	+	w	w	+	+	+	+	+	+	-	+	+	+	+	-	-	-	4	nd	nd	nd	nd
BA 4CM S8																		3	nd	nd	nd	nd
BA 4CM S9	+	w	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	4	nd	nd	nd	nd
BA 4CM S10	+	w	-	+	+	+	+	+	+	-	+	+	+	w	w	-	-	3	nd	nd	nd	nd
BA 4CM NS1	+	+	+															4	+	nd	nd	nd
BA 4CM NS5	+	+	+															4	+	nd	nd	nd
BA 4CM NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	nd	nd
BA 4CM NS7	+	-	+															4	+	nd	nd	nd
BA 4CM NS8	+	+	+															4	+	nd	nd	nd
BA 6NM S1	-	-	-	+	+	+	+										ng	nd	nd	nd	nd	
BA 6NM S2	-	-	-	+	+	+	+										3	nd	nd	nd	nd	
BA 6NM S3	+	-	-	+	+	+	+										3	nd	nd	nd	nd	
BA 6NM S4	-	-	-	+	+	+	+										ng	nd	nd	nd	nd	
BA 6NM S5	-	-	-	+	+	+	+										3	nd	nd	nd	nd	
BA 6NM S6	+	w	w	+	+	+	+										3	nd	nd	nd	nd	
BA 6NM S7	-	-	-	+	+	+	+										ng	nd	nd	nd	nd	
BA 6NM S8	+	w	w	+	+	+	+										3	nd	nd	nd	nd	
BA 6NM NS1	+	+	+															3	+	nd	nd	nd
BA 6NM NS2	+	-	-															3	+	+	+	nd
BB 0CM S1	+	+	w	+	+	+	+	-	w	-	-	-	-	+	-	-	-	4	nd	nd	nd	nd
BB 0CM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd	nd	nd
BB 0CM S3																		4	nd	nd	nd	nd
BB 0CM S4	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd	nd	nd
BB 0CM S8	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd	nd	nd
BB 0CM S9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	+	+	4	nd	nd	nd	nd
BB 0CM S10																		5	nd	nd	nd	nd
BB 0CM NS1	+	+	+															4	+	nd	nd	nd
BB 0CM NS5	+	+	+															4	+	nd	nd	nd

Table C.3 (continued)

Name	Alcohol Tolerance (%)						SO ₂ Tolerance (mg/L)			Temperature Tolerance (°C)			pH Tolerance			Carbohydrate Fermentation Test			H ₂ S Product	Identification	
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2		
BB 0CM NS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	
BB 0CM NS8	+	+	+															4	+	nd	
BB 4CM S1	+	-	-	+	+	+	+	+	+	-	+	+	+	+	w	-	-	ng	nd	nd	
BB 4CM S2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd	
BB 4CM S4	+	+	w	+	+	+	+	+	+	w	+	+	+	+	w	-	-	3	nd	nd	
BB 4CM S6	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	4	nd	nd	
BB 4CM S9	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	w	4	nd	nd		
BB 4CM S10	+	+	w	+	+	+	+	+	+	-	+	+	+	+	w	-	-	2	nd	+	
BB 4CM NS1	+	+	+															3	+	nd	
BB 4CM NS2	+	+	+															4	+	nd	
BB 4CM NS3	+	-	-															4	+	nd	
BB 4CM NS4	+	+	+															4	+	nd	
BB 4CM NS6	+	-	-															4	+	nd	
BB 4CM NS7																		3	+	nd	
BB 6NM S1	+	w	w	+	+	+	+											3	nd	nd	
BB 6NM S2	+	w	w	+	+	+	+											3	nd	nd	

BA: A Parallel of Wine Made from Boğazkere Grape , BB: B Parallel of Wine Made from Boğazkere Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 6NM: 6. Day of Normal Maceration; S: *Saccharomyces*; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; nd: not determined.

Table C.4. The Results of Carbohydrate Fermentation Test and Phenotypic Properties of *Saccharomyces* Strains from Isolated in Must and Wine of Dimrit

Name	Alcohol Tolerance (%)						SO ₂ Tolerance (mg/L)			Temperature Tolerance (°C)			pH Tolerance			Carbohydrate Fermentation Test				<i>H₂S</i> Product	Identification
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose				
																		ITS	D1/D2		
DA 0CM S1																		3	nd	nd	
DA 0CM S2	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd	
DA 0CM S3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
DA 0CM S4	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	2	nd	nd	
DA 0CM S5	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	nd	
DA 4CM S1	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	nd	+	
DA 4CM S2	+	+	w	+	+	+	+	+	w	-	w	w	-	-	+	w	-	2	nd	nd	
DA 4CM S3	+	w	w	+	+	+	+	+	+	-	w	w	-	+	+	w	w	2	nd	+	
DA 4CM S4	+	+	+	+	+	+	+	+	+	-	+	+	+	-	+	w	w	2	nd	nd	
DA 4CM S5	+	w	w	+	+	+	+	+	+	-	+	+	+	-	-	w	w	3	nd	nd	
DA 4CM S6	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	2	nd	nd	
DA 4CM S7	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	nd	nd	
DA 4CM S8	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	+	
DA 4CM S9	w	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	nd	
DA 4CMS10	w	-	-	+	+	+	+	w	+	-	+	+	+	+	+	-	-	ng	nd	nd	
DA 4NM S1	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	nd	+	
DA 4NM S2	+	+	w	+	+	+	+	+										3	nd	nd	
DA 4NM S3	+	-	-	+	+	+	+	+										3	nd	nd	
DA 4NM S4	+	w	-	+	+	+	+	+										3	nd	nd	
DB 0CM S1	w	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	3	nd	nd	
DB 0CM S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	3	nd	nd	
DB 0CM S3	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	+	4	nd	nd	
DB 0CM S4	+	+	+	+	+	+	+	+	+	w	+	+	+	-	+	-	-	2	nd	nd	
DB 0CM S5	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
DB 4CM S1	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
DB 4CM S2	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
DB 4CM S3	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	nd	
DB 4CM S4	+	w	w	+	+	+	+	+	+	w	+	+	+	+	+	w	w	3	nd	nd	
DB 4CM S5	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	w	4	nd	nd	
DB 4CM S6	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	w	3	nd	nd	
DB 4CM S7	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	2	nd	nd	
DB 4CM S8	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	w	w	4	nd	nd	
DB 4CM S9	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
DB 4CMS10	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
DB 4NM S1	+	w	w	+	+	+	+	+										3	nd	nd	
DB 4NM S2	+	w	w	+	+	+	+	+										3	nd	nd	
DB 4NM S3	+	w	w	+	+	+	+	+										4	nd	nd	
DB 4NM S4	-	-	-	+	w	w	w											ng	nd	nd	
DB 4NM S5	+	+	w	+	+	+	+	+										3	nd	nd	
DB 4NMS6	+	w	w	+	+	+	+	+										4	nd	nd	

DA: A Parallel of Wine Made from Dimrit Grape , DB: B Parallel of Wine Made from Dimrit Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 4NM: 4. Day of Normal Maceration; S: *Saccharomyces*; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black;; ng: not grown, nd: not determined.

Table C.5. The Results of Carbohydrate Fermentation Test and Phenotypic Properties of *Saccharomyces* Strains from Isolated in Must and Wine of Emir

Name	Alcohol						Temperature				pH			Carbohydrate Fermentation Test				<i>H₂S</i> Product		Identification	
	Tolerance (%)	SO ₂ Tolerance (mg/L)					Tolerance (°C)	Tolerance			Sucrose	Maltose	Mannitol	Lactose	ITS	DI/D2					
	10	13	15	50	100	150	200	28	37	45	6	4	3								
EA 0W S2	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	w	3	nd	nd		
EA 0W S5	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	w	3	nd	nd		
EA 0W S9	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	w	ng	nd	nd		
EA 0W NS1	+	+	+														3	+	nd		
EA 0W NS2	+	+	-														4	+	nd		
EA 0W NS4	+	+	-														3	+	nd		
EA 0W NS5	+	+	-	+	+	+	+	+	+	-	+	+	+	+	-	-	2	+	nd		
EA 0W NS6	+	+	+														3	+	nd		
EA 0W NS7	+	-	+														3	+	nd		
EA 0W NS9	+	+	+	+	+	+	+	+	+	-	+	+	+	+	-	-	2	+	nd		
EA 0W NS10	+	+	-														3	+	nd		
EA 1W S1	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	+	4	nd	nd		
EA 1W S2	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	w	3	nd	nd		
EA 1W S3	w	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 1W S4	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 1W S5	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	+	3	nd	nd		
EA 1W S6	+	-	-	+	+	+	+	+	+								4	nd	nd		
EA 1W S7	+	-	-	+	+	+	+	+	+								4	nd	nd		
EA 1W S8	+	-	-	+	+	+	+	+	+								4	nd	nd		
EA 1W S9	+	-	-	+	+	+	+	+	+								4	nd	nd		
EA 1W S10	+	w	-	+	+	+	+	+	+								4	nd	nd		
EA 1W NS5	+	+	-	+	+	+	+	+	+	-	+	+	+	+	-	-	1	+	nd		
EA 1WNS6	+	+	+	+	+	+	+	+	+	-	+	+	+	+	-	-	1	+	nd		
EA 1W NS7	+	+	+														4	+	nd		
EA 1W NS8	+	+	+														3	+	nd		
EA 1W S10	+	+	+	+	+	+	+	+	+	w	+	+	+	+	-	-	2	+	nd		
EA 2W S1	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 2W S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	-	4	nd	nd		
EA 2W S3	w	w	-	+	+	+	+	+	+	-	+	+	+	+	w	w	3	nd	nd		
EA 2W S4	+	-	-	+	+	+	+	+	+	-	+	+	+	+	w	4	nd	nd			
EA 2W S5	+	w	w	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 2W NS1	+	+	+														3	+	nd		
EA 2W NS2	+	+	-														3	+	nd		
EA 3W S1	w	w	w	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 3W S2	+	-	-	+	+	+	+	+	+	-	+	+	+	+	w	2	nd	nd			
EA 3W S3	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	2	nd	nd		
EA 3W S4	+	-	-	+	+	+	+	+	+	-	+	+	+	+	w	w	1	nd	nd		
EA 3W S5	+	-	-	+	+	+	+	+	+	-	+	+	+	+	-	-	2	nd	nd		
EA 4W S1	+	-	-	+	+	+	+	+	+	-	+	+	+	+	w	w	3	nd	nd		
EA 4W S2	+	w	w	+	+	+	+	+	+	-	+	+	+	+	w	-	3	nd	nd		
EA 4W S3	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 4W S4	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	-	3	nd	nd		
EA 4W S5	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	-	1	nd	nd		
EB 0W S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	-	-	2	nd	nd		

Table C.5 (continued)

Name	Alcohol Tolerance(%)						SO ₂ Tolerance (mg/L)			Temperature Tolerance (°C)			pH Tolerance			Carbohydrate Fermentation Test				H ₂ S Product	Identification
	10	13	15	50	100	150	200	28	37	45	6	4	3	Sucrose	Maltose	Mannitol	Lactose	ITS	D1/D2		
EB 0W S3	+	w	w	+	+	+	+	+	+	-	+	+	+	-	+	-	-	3	nd	nd	
EB 0W S5	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	4	nd	nd	
EB 0W S7	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	3	nd	nd	
EB 0W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	
EB 0W NS2	+	+	+															3	+	nd	
EB 0W NS3	+	+	-															3	+	nd	
EB 0W NS4	+	+	+															3	+	nd	
EB 0W NS6	+	+	+															3	+	nd	
EB 0W NS7	+	+	+															3	+	nd	
EB 0W NS8	+	+	-															3	+	nd	
EB 0W NS9	+	+	+															3	+	nd	
EB 0W NS10	+	+	+															4	+	nd	
EB 1W S1	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
EB 1W S2	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
EB 1W S3	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	nd	
EB 1W S4	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	w	3	nd	nd	
EB 1W S5	+	+	+	+	+	+	+	+	+	-	+	+	+	+	-	-	-	2	nd	nd	
EB 1W S6	+	w	-	+	+	+	+	+										4	nd	nd	
EB 1W S7	+	w	-	+	+	+	+	+										4	nd	nd	
EB 1W S8	+	w	-	+	+	+	+	+										4	nd	nd	
EB 1W S9	-	-	-	+	+	+	+	+										ng	nd	nd	
EB 1W S10	+	+	-	+	+	+	+	+										4	nd	nd	
EB 1W NS1	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	nd	
EB 1W NS2	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	
EB 1W NS3	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	
EB 1W NS4	+	+	+															3	+	nd	
EB 1W NS6	+	+	+															3	+	nd	
EB 1W NS7	+	+	+	+	+	+	+	+	+	-	+	+	+	+	+	-	-	1	+	nd	
EB 1W NS9	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	+	nd	
EB 1W NS10	+	+	+															3	+	nd	
EB 2W S1	+	+	w	+	+	+	+	+	+	-	+	+	+	+	+	w	3	nd	nd		
EB 2W S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	4	nd	nd		
EB 2W S3	+	w	-	+	+	+	+	+	+	-	+	+	+	-	-	w	w	3	nd	nd	
EB 2W S4	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	w	3	nd	nd	
EB 2W S5	+	+	w	+	+	+	+	+	+	-	+	+	+	-	-	w	w	2	nd	nd	
EB 3W S1	+	-	-	+	+	+	+	+	+	-	+	+	+	-	+	-	-	2	nd	nd	
EB 3W S2	+	+	-	+	+	+	+	+	+	-	+	+	+	+	+	w	-	3	nd	nd	
EB 3W S3	+	-	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
EB 3W S4	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	w	3	nd	nd		
EB 3W S5	+	+	w	+	+	+	+	+	+	-	w	+	w	+	+	-	-	1	nd	+	
EB 4W S1	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	2	nd	nd	
EB 4W S2	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
EB 4W S3	+	w	W	+	+	+	+	+	+	-	+	+	+	+	+	-	-	3	nd	nd	
EB 4W S4	+	w	-	+	+	+	+	+	+	-	+	+	+	+	+	-	+	3	nd	nd	
EB 4W S5	+	w	w	+	+	+	+	+	+	-	+	+	+	+	+	w	3	nd	nd		

EA: A Parallel of Wine Made from Emir Grape , EB: B Parallel of Wine Made from Emir Grape; 0W: 0.Week, 1W: 1.Week, 2W:2. Week, 3W: 3. Week, 4W: 4. Week;
S: *Saccharomyces* ; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black; nd: not determined.

D. Phenotypic Properties of All Isolated Non-Saccharomyces Yeasts

Table D.1. Phenotypic Properties of All Isolated Non-Saccharomyces Yeasts

Non-Saccharomyces	Name	Alcohol Tolerance (%)				SO ₂ Tolerance (mg/L)			H ₂ S Production		Identification		
		10	13	15	50	100	150	200			ITS	D1/D2	
												Domain	
<i>Hanseniaspora guilliermondii</i>	K1	KA 0.CM NS1	-	-	-	+	+	+	+	+	3	+	nd
	K5	KA 0.CM NS5	-	-	-	+	+	+	+	+	4	+	+
	K31	KA 4.NM NS2				+	+	+	-		4	+	nd
	K33	KA 4.NM NS6									4	+	nd
	K36	KB 4.NM NS1									4	+	nd
	K39	KB 4.NM NS7									4	+	nd
	K61	KA 4.NM NS1									4	+	nd
	K68	KB 4.NM NS6	w	-	-	+	+	+	+	+	4	+	nd
	K69	KB 4.NM NS8	-	-	-	+	+	+	+	+	5	+	nd
	K70	KA 0.CM NS2	-	-	-	+	+	+	+	+	3	+	nd
<i>Hanseniaspora opuntiae</i>	K7	KB 0.CM NS2									4	+	nd
	K8	KB 0.CM NS3	-	-	-	+	+	+	+	+	4	+	nd
	K10	KB 0.CM NS5	vw	vw	vw	+	+	+	+	+	3	+	nd
	K11	KA 4.CM NS1									4	+	nd
	K12	KA 4.CM NS2	-	-	-	+	+	+	+	+	3	+	+
	K15	KA 4.CM NS5	w	-	-	+	+	+	+	+	ng	+	nd
	K16	KB 4.CM NS1									ng	+	nd
	K17	KB 4.CM NS2	-	-	-	+	+	+	+	+	3	+	nd
	K19	KB 4.CM NS4	-	-	-	+	+	+	+	+	3	+	nd
	K20	KB 4.CM NS5	+	-	-	+	+	+	+	+	4	+	nd
<i>Rhodotorula mucilaginosa</i>	K21	KA 2.NM NS1	+	-	-	+	+	+	+	-	2	+	nd
	K27	KB 2.NM NS4									4	+	nd
	K32	KA 4.NM NS4	-	-	-	+	+	+	+	+	5	+	nd
	K34	KA 4.NM NS8	w			+	+	+	+	+	5	+	nd
	K35	KA 4.NM NS10									5	+	+
	K37	KB 4.NM NS3									4	+	nd
	K38	KB 4.NM NS5									4	+	nd
	K40	KB 4.NM NS9	+	-	-	+	+	+	+	+	4	+	nd
	K51	KA 2.NM NS2	-	-	-	+	+	+	+	+	4	+	nd
	K60	KB 2.NM NS9									4	+	nd
<i>Hanseniaspora uvarum</i>	K62	KA 4.NM NS3									4	+	+
	K63	KA 4.NM NS5	-	-	-	+	+	+	+	+	2	+	nd
	K64	KA 4.NM NS7	+	-	-	+	+	+	+	-	4	+	nd
	K65	KA 4.NM NS9									ng	+	+
	K66	KB 4.NM NS2	w			+	+	+	+	+	5	+	nd
	K67	KB 4.NM NS4	-	-	-	+	+	+	+	+	3	+	nd
	K9	KB 0.CM NS4	-	-	-	+	+	+	+	+	3	+	nd
	K13	KA 4.CM NS3	+	vw	-	+	+	+	+	+	3	+	nd
	K3	KA 0.CM NS3	-	-	-	+	+	+	+	+	5	+	nd
	K25	KA 2.NM NS9	w			+	+	+	+	+	6	+	+
	K26	KB 2.NM NS2	+	w	-	+	+	+	+	+	5	+	nd
	K52	KA 2.NM NS4	-	-	-	+	+	+	+	+	3	+	nd

Table D.1 (continued)

Non-Saccharomyces	Name		Alcohol				SO ₂				H ₂ S		Identification
			Tolerance (%)				Tolerance (mg/L)				Production		
			10	13	15	50	100	150	200			ITS	D1/D2 Domain
<i>Hanseniaspora uvarum</i>	K54	KA 2.NM NS8	w			+	+	+	+	5	+	nd	
	K70	KB 4.NM NS10	+	-	-	+	+	+	-	ng	+	+	
<i>Wickerhamomyces anomalus</i>	K41	KA 0.CM S1	-	-	-	+	+	+	+	3	+	nd	
	K42	KB 0.CM S5	+	+	+	+	+	+	+	ng	+	nd	
<i>Metschnikowia aff. fructicola</i>	K43	KA 4.CM S1	-	-	-	-	-	-	-	ng	+	+	
<i>Hanseniaspora uvarum</i>	E11	EA 1.W NS1	-	-	-	+	+	+	+	3	+	nd	
	E12	EA 1.W NS2								4	+	nd	
	E13	EA 1.W NS3								ng	+	nd	
	E14	EA 1.W NS4	vw	vw	vw	+	+	+	+	3	+	nd	
<i>Wickerhamomyces anomalus</i>	E37	EA 0.NM NS8	-	-	-	+	+	+	+	3	+	nd	
<i>Lachancea thermotolerans</i>	O1	OA 0.CM NS1	-	-	-	+	+	+	+	3	+	+	
	O2	OB 0.CM NS2								4	+	nd	
	O3	OB 0.CM NS3								4	+	nd	
	O4	OB 0.CM NS5								ng	+	nd	
	O6	OA 4.CM NS3								4	+	nd	
	O8	OB 4.CM NS3								ng	+	nd	
	O9	OB 4.CM NS4	-	-	-	+	+	+	+	3	+	nd	
	O11	OA 0.CM NS2								ng	+	nd	
	O13	OA 0.CM NS4								ng	+	nd	
	O14	OA 0.CM NS5								5	+	nd	
	O15	OA 0.CM NS6								5	+	nd	
	O16	OA 0.CM NS7								5	+	nd	
	O17	OA 0.CM NS8								5	+	nd	
	O18	OA 0.CM NS9								5	+	nd	
	O19	OA 0.CM NS10								5	+	nd	
	O20	OB 0.CM NS1								5	+	nd	
	O21	OB 0.CM NS4								5	+	nd	
	O22	OB 0.CM NS6								5	+	nd	
	O23	OA 4.CM NS1								ng	+	nd	
	O25	OA 4.CM NS6								5	+	nd	
	O26	OA 4.CM NS7								5	+	nd	
	O27	OA 4.CM NS8								5	+	nd	
	O28	OA 4.CM NS9								5	+	nd	
	O29	OA 4.CM NS10								5	+	nd	
	O30	OB 4.CM NS1								5	+	nd	
	O31	OB 4.CM NS2								5	+	nd	
	O32	OB 4.CM NS6								5	+	nd	
	O33	OB 4.CM NS7								5	+	nd	
	O34	OB 4.CM NS8								5	+	nd	
	O35	OB 4.CM NS9								6	+	nd	
	O36	OB 4.CM NS10								5	+	nd	
<i>Wickerhamomyces anomalus</i>	O37	OA 4CM S3	+	+	w	+	+	+	+	3	nd	+	
<i>Hanseniaspora opuntiae</i>	O5	OA 4.CM NS2	vw	vw	vw	+	+	+	+	3	+	nd	
	O7	OA 4.CM NS4	+	+	+	+	+	+	+	4	+	+	
<i>Metschnikowia pulcherrima</i>	B10	BB 0.CM NS2	w	w	-	+	+	+	+	4	+	nd	
	B12	BB 0.CM NS4								5	+	nd	

Table D.1 (continued)

Non-Saccharomyces	Name		Alcohol			SO ₂			H ₂ S			Identification
			Tolerance (%)			Tolerance (mg/L)			Production			
			10	13	15	50	100	150	200			ITS
<i>Metschnikowia pulcherrima</i>	B15	BB 0.CM NS7								5	+	nd
	B30	BB 4.CM NS5								5	+	nd
<i>Metschnikowia aff. fructicola</i>	B33	BB 4.CM NS8	-	-	-	+	+	+	+	3	+	+
<i>Metschnikowia aff. pulcherrima</i>	B11	BB 0.CM NS3								5	+	+
<i>Solicoccozyma aeria</i>	B4	BA 0.CM NS4	-	-	-	+	w	w	W	5	+	+
<i>Metschnikowia aff. pulcherrima</i>	D2	DA 0.CM NS2	+	+	+	+	+	+	+	ng	+	+
<i>Metschnikowia sinensis</i> (ITS)	D3	DA 0.CM NS3	-	-	-	+	+	+	+	3	+	
<i>Metschnikowia pulcherrima</i> (D1/D2)												+
<i>Starmerella bacillaris</i>	D10	DA 4.CM NS2	w	w	-	+	+	+	+	4	+	+
<i>Metschnikowia pulcherrima</i>	D11	DA 4.CM NS3	-	-	-	+	+	+	+	2	+	nd
	D12	DA 4.CM NS4	-	-	-	+	+	+	+	3	+	nd
	D13	DB 4.CM NS1								5	+	nd
	D14	DB 4.CM NS2								4	+	nd
	D4	DA 0.CM NS4								4	+	nd
	D8	DB 0.CM NS4								4	+	+
<i>Metschnikowia chrysoperlae</i>	D6	DB 0.CM NS2								4	+	nd
<i>Metschnikowia fructicola</i>	D15	DB 4.CM NS3	-	-	-	+	+	+	+	4	+	+
<i>Metschnikowia chrysoperlae</i>	14 (Ç)		-	-	-	+	+	+	+	ng	+	nd
<i>Metschnikowia aff. fructicola</i>	M1 (Ç)		+	+	+	+	+	+	+	ng	nd	+

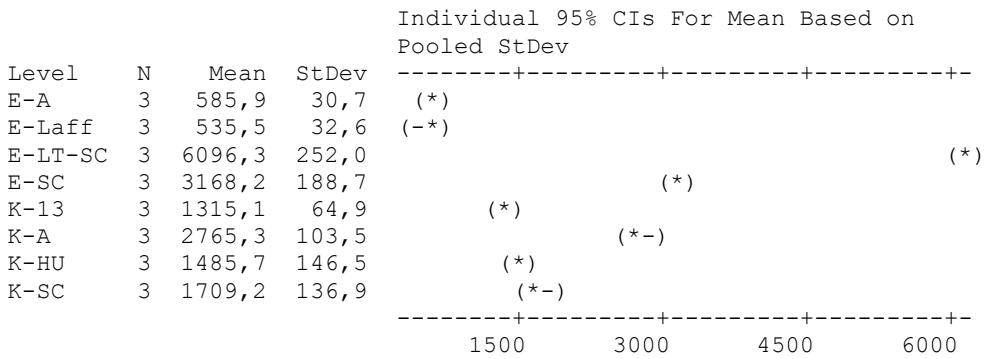
KA: A Parallel of Wine Made from Kalecik Karası Grape , KB: B Parallel of Wine Made from Kalecik Karası Grape, OA: A Parallel of Wine Made from Öküzgözü Grape, OB: B Parallel of Wine Made from Öküzgözü Grape, BA: A Parallel of Wine Made from Boğazkere Grape, BB: B Parallel of Wine Made from Boğazkere Grape, DA: A Parallel of Wine Made from Dimitri Grape, DB: B Parallel of Wine Made from Dimitri Grape, EA: A Parallel of Wine Made from Emir Grape, EB: B Parallel of Wine Made from Emir Grape; 0CM: 0. Day of Cold Maceration, 4CM: 4. Day of Cold Maceration, 2NM: 2. Day of Normal Maceration, 4NM: 4. Day of Normal Maceration, 6NM: 6. Day of Normal Maceration, 1W: 1. Week, 2W: 2. Week, 3W: 3. Week, 4W: 4. Week; NS: Non-Saccharomyces ; + positive growth, w weak growth, - negative growth; 1: white, 2: cream, 3: light brown, 4: brown, 5: dark brown, 6: black. Ç*: Isolated by Çağrı Çavdaroglu nd: Not determined

E. Statistical Analysis of Aroma Compounds for the Emir and Kalecik Karası Wines

One-way ANOVA: 1-Propanol versus Group

Source	DF	SS	MS	F	P
Group	7	70044176	10006311	512,25	0,000
Error	16	312545	19534		
Total	23	70356721			

S = 139,8 R-Sq = 99,56% R-Sq(adj) = 99,36%



Pooled StDev = 139,8

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	6096,3	A
E-SC	3	3168,2	B
K-A	3	2765,3	C
K-SC	3	1709,2	D
K-HU	3	1485,7	D
K-13	3	1315,1	D
E-A	3	585,9	E
E-Laff	3	535,5	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper			
--						
E-Laff	-445,8	-50,4	345,0		(*)	
E-LT-SC	5115,0	5510,4	5905,8			(*-)
E-SC	2186,9	2582,3	2977,7			(-*)

K-13	333,8	729,2	1124,6	(*-)	
K-A	1784,0	2179,4	2574,8	(*-)	
K-HU	504,3	899,7	1295,1	(*)	
K-SC	727,9	1123,3	1518,7	(-*)	
--				+-----+-----+-----+	
		-6000	-3000	0	3000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper	+-----+-----+-----+-----	
--				+-----+-----+-----+-----	
E-LT-SC	5165,5	5560,8	5956,2		
(-*)					
E-SC	2237,3	2632,7	3028,1	(-*)	
K-13	384,2	779,6	1175,0	(-*)	
K-A	1834,4	2229,8	2625,2	(*)	
K-HU	554,8	950,2	1345,6	(*)	
K-SC	778,3	1173,7	1569,1	(*)	
--				+-----+-----+-----+-----	
		-6000	-3000	0	3000

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	+-----+-----+-----+-----	
--				+-----+-----+-----+-----	
E-SC	-3323,5	-2928,1	-2532,7	(*)	
K-13	-5176,6	-4781,2	-4385,8	(*)	
K-A	-3726,4	-3331,0	-2935,6	(*)	
K-HU	-5006,1	-4610,7	-4215,3	(-*)	
K-SC	-4782,5	-4387,1	-3991,7	(*)	
--				+-----+-----+-----+-----	
		-6000	-3000	0	3000

Group = E-SC subtracted from:

Group	Lower	Center	Upper	+-----+-----+-----+-----	
--				+-----+-----+-----+-----	
K-13	-2248,5	-1853,1	-1457,7	(*)	
K-A	-798,3	-402,9	-7,5	(-*)	
K-HU	-2078,0	-1682,6	-1287,2	(-*)	
K-SC	-1854,4	-1459,0	-1063,6	(*)	
--				+-----+-----+-----+-----	
		-6000	-3000	0	3000

Group = K-13 subtracted from:

Group	Lower	Center	Upper	+-----+-----+-----+-----	
K-A	1054,8	1450,2	1845,6	(*)	
K-HU	-224,8	170,6	566,0	(-*)	
K-SC	-1,3	394,1	789,5	(*-)	
--				+-----+-----+-----+-----	
		-6000	-3000	0	3000

Group = K-A subtracted from:

Group	Lower	Center	Upper				
--							
K-HU	-1675,1	-1279,7	-884,3		(-*)		
K-SC	-1451,5	-1056,1	-660,7		(*-)		
--							
				-6000	-3000	0	3000

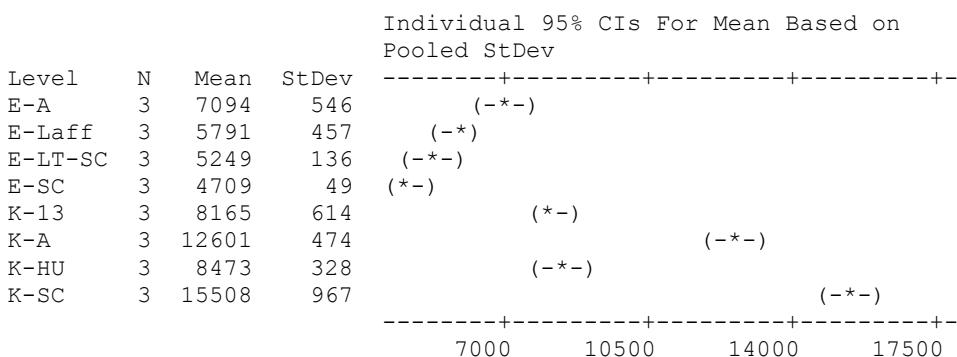
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	-171,8	223,5	618,9		(-*)		
				-6000	-3000	0	3000

One-way ANOVA: Isobutyl Alcohol versus Group

Source	DF	SS	MS	F	P
Group	7	300833685	42976241	158,33	0,000
Error	16	4342980	271436		
Total	23	305176665			

S = 521,0 R-Sq = 98,58% R-Sq(adj) = 97,95%



Pooled StDev = 521

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-SC	3	15508	A
K-A	3	12601	B
K-HU	3	8473	C
K-13	3	8165	C
E-A	3	7094	C D
E-Laff	3	5791	D E
E-LT-SC	3	5249	E
E-SC	3	4709	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-2777	-1303	171	(-*)
E-LT-SC	-3319	-1845	-371	(-*)
E-SC	-3859	-2385	-911	(--*)
K-13	-403	1070	2544	(--*)
K-A	4033	5506	6980	(-*)
K-HU	-95	1379	2853	(-*)
K-SC	6940	8414	9888	(-*)

-----+-----+-----+-----+
-7000 0 7000 14000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper	
E-LT-SC	-2016	-542	932	(-*)
E-SC	-2556	-1082	392	(---)
K-13	899	2373	3847	(-*)
K-A	5335	6809	8283	(-*)
K-HU	1208	2682	4156	(-*)
K-SC	8243	9717	11191	(-*)

-----+-----+-----+-----+
-7000 0 7000 14000

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	
E-SC	-2014	-540	934	(-*)
K-13	1442	2915	4389	(-*)
K-A	5878	7352	8825	(--*)
K-HU	1750	3224	4698	(-*)
K-SC	8785	10259	11733	(-*)

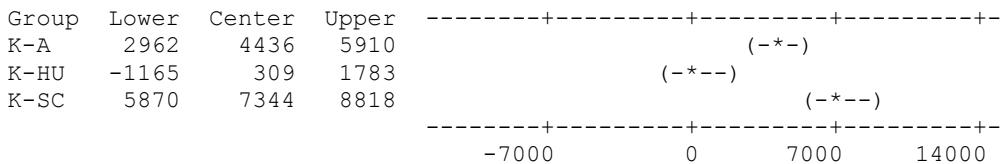
-----+-----+-----+-----+
-7000 0 7000 14000

Group = E-SC subtracted from:

Group	Lower	Center	Upper	
K-13	1981	3455	4929	(-*)
K-A	6417	7891	9365	(-*)
K-HU	2290	3764	5238	(-*)
K-SC	9325	10799	12273	(---)

-----+-----+-----+-----+
-7000 0 7000 14000

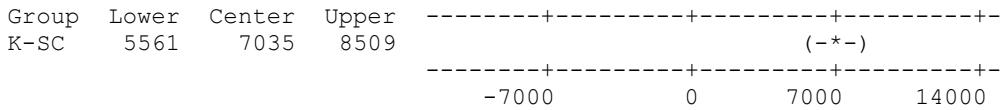
Group = K-13 subtracted from:



Group = K-A subtracted from:



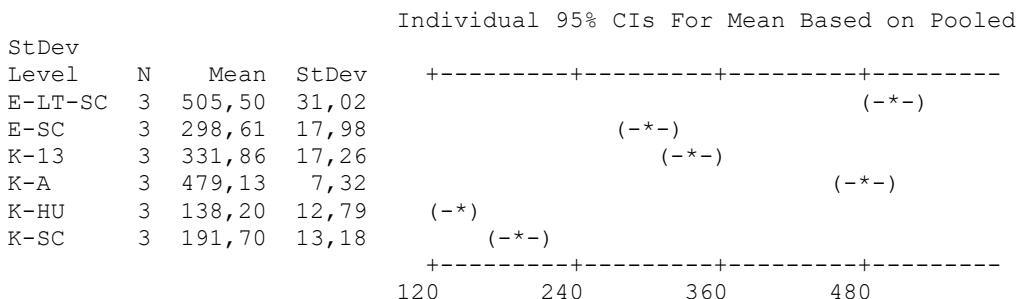
Group = K-HU subtracted from:



One-way ANOVA: 1-Butanol versus Group

Source	DF	SS	MS	F	P
Group	5	329215	65843	200,11	0,000
Error	12	3948	329		
Total	17	333164			

S = 18,14 R-Sq = 98,81% R-Sq(adj) = 98,32%



Pooled StDev = 18,14

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	505,50	A
K-A	3	479,13	A
K-13	3	331,86	B
E-SC	3	298,61	B

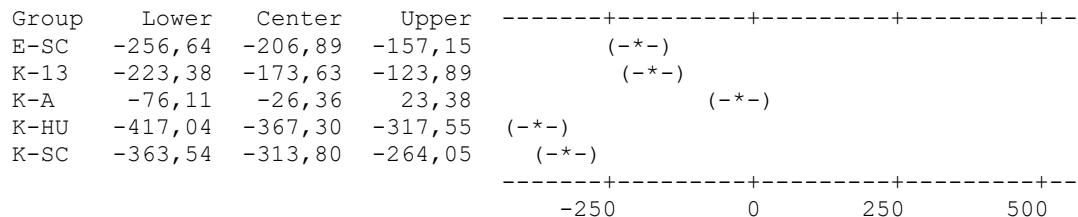
K-SC	3	191,70	C
K-HU	3	138,20	D

Means that do not share a letter are significantly different.

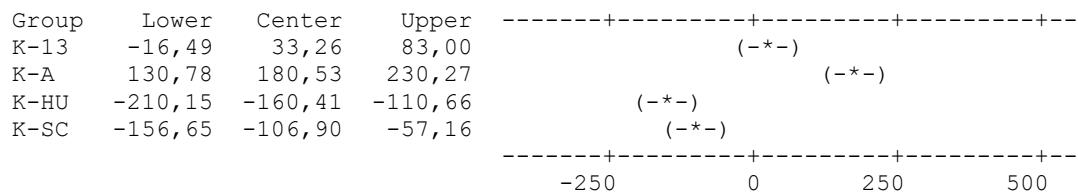
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

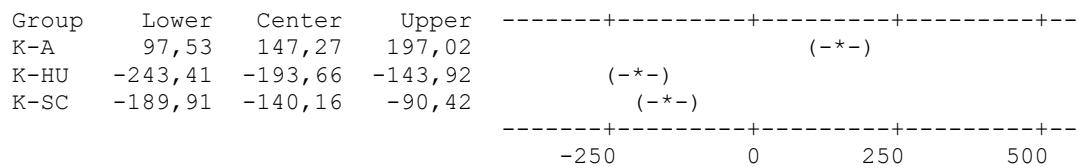
Group = E-LT-SC subtracted from:



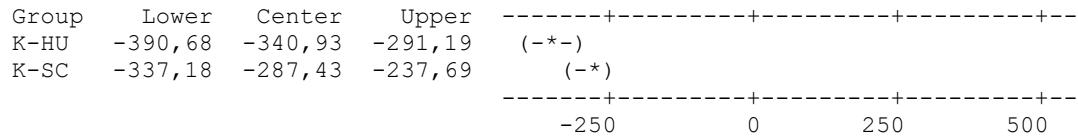
Group = E-SC subtracted from:



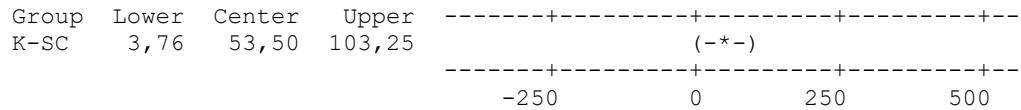
Group = K-13 subtracted from:



Group = K-A subtracted from:



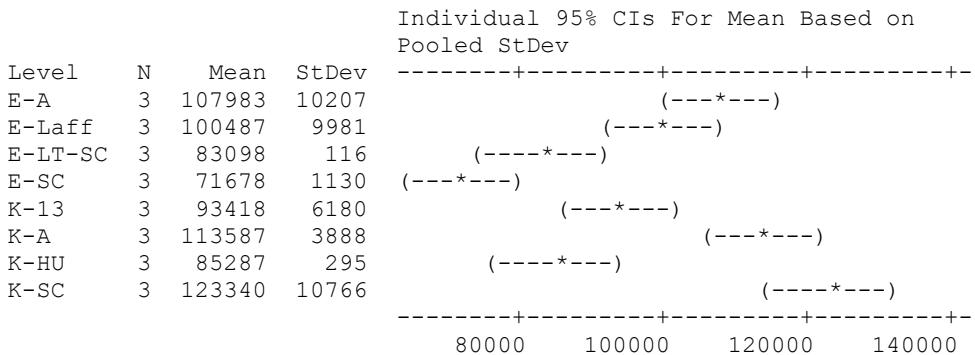
Group = K-HU subtracted from:



One-way ANOVA: Isoamyl Alcohol versus Group

Source	DF	SS	MS	F	P
Group	7	6255623964	893660566	19,09	0,000
Error	16	748818100	46801131		
Total	23	7004442064			

S = 6841 R-Sq = 89,31% R-Sq(adj) = 84,63%



Pooled StDev = 6841

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-SC	3	123340	A
K-A	3	113587	A B
E-A	3	107983	A B C
E-Laff	3	100487	B C D
K-13	3	93418	C D
K-HU	3	85287	D E
E-LT-SC	3	83098	D E
E-SC	3	71678	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

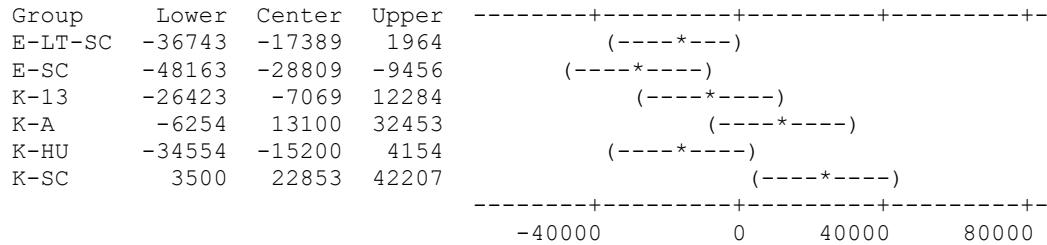
Individual confidence level = 99,68%

Group = E-A subtracted from:

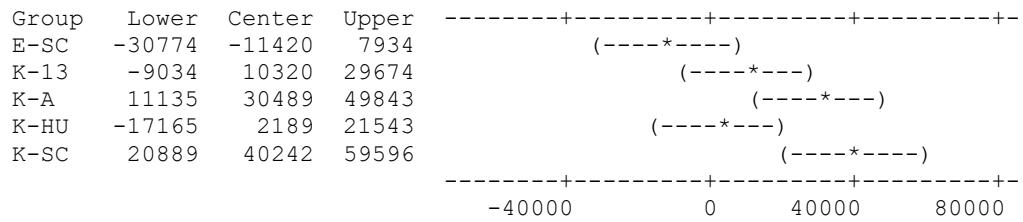
Group	Lower	Center	Upper	
E-Laff	-26850	-7496	11857	(-----*-----)
E-LT-SC	-44239	-24886	-5532	(-----*-----)
E-SC	-55659	-36306	-16952	(-----*-----)
K-13	-33919	-14566	4788	(-----*-----)
K-A	-13750	5603	24957	(-----*-----)
K-HU	-42050	-22697	-3343	(-----*-----)



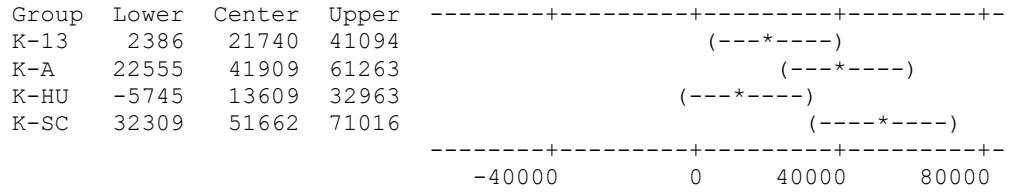
Group = E-Laff subtracted from:



Group = E-LT-SC subtracted from:



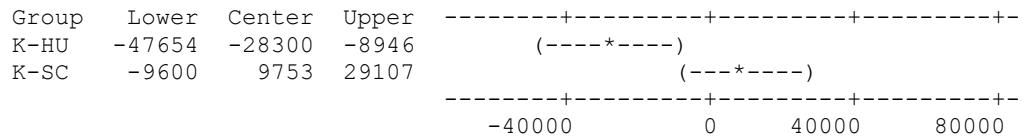
Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



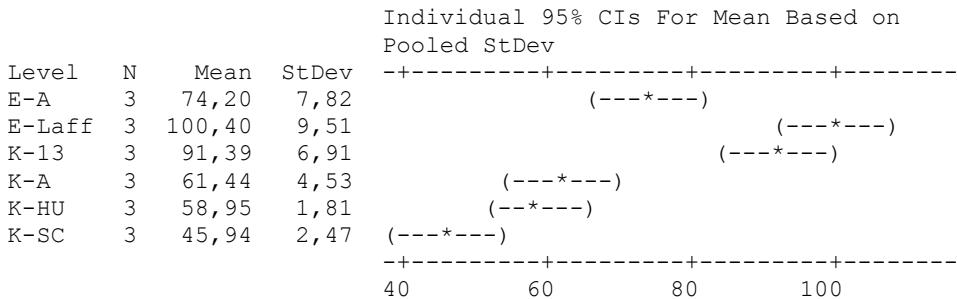
Group = K-HU subtracted from:



One-way ANOVA: 3-Methyl-1-pentanol versus Group

Source	DF	SS	MS	F	P
Group	5	6445,2	1289,0	33,71	0,000
Error	12	458,8	38,2		
Total	17	6904,0			

S = 6,183 R-Sq = 93,35% R-Sq(adj) = 90,59%



Pooled StDev = 6,18

Grouping Information Using Tukey Method

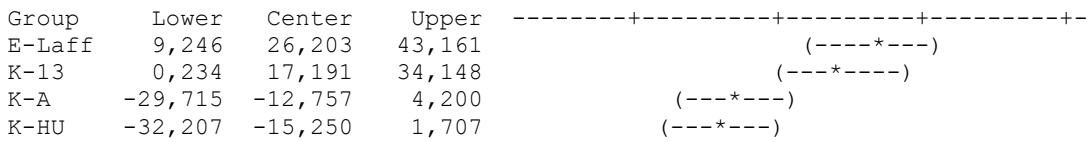
Group	N	Mean	Grouping
E-Laff	3	100,400	A
K-13	3	91,387	A
E-A	3	74,197	B
K-A	3	61,439	B C
K-HU	3	58,947	B C
K-SC	3	45,938	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

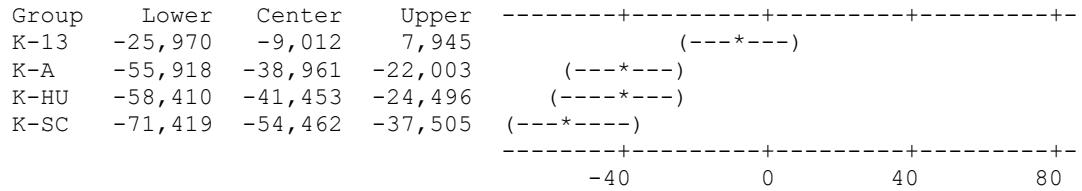
Individual confidence level = 99,43%

Group = E-A subtracted from:

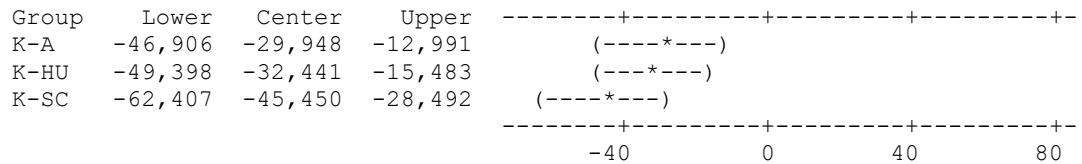




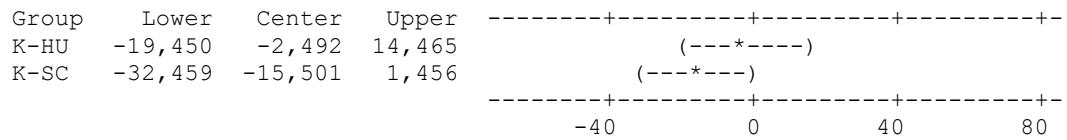
Group = E-Laff subtracted from:



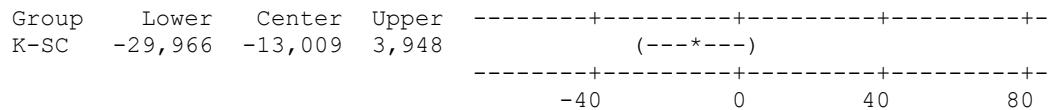
Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:

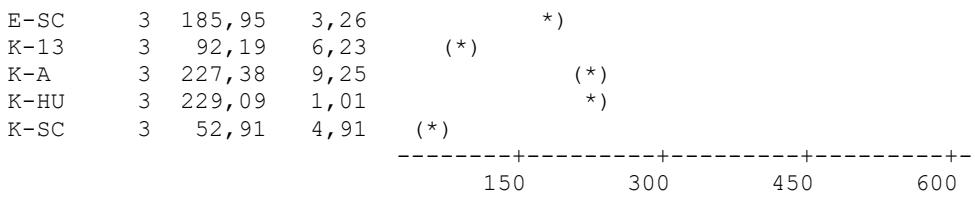


One-way ANOVA: 3-Ethoxy-1-propanol versus Group

Source	DF	SS	MS	F	P
Group	7	531008,1	75858,3	983,61	0,000
Error	16	1234,0	77,1		
Total	23	532242,1			

S = 8,782 R-Sq = 99,77% R-Sq(adj) = 99,67%

Individual 95% CIs For Mean Based on Pooled StDev					
Level	N	Mean	StDev		
E-A	3	38,10	3,78	(*)	
E-Laff	3	41,06	3,75	(*)	
E-LT-SC	3	510,90	20,70		(*)



Pooled StDev = 8,78

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	510,90	A
K-HU	3	229,09	B
K-A	3	227,38	B
E-SC	3	185,95	C
K-13	3	92,19	D
K-SC	3	52,91	E
E-Laff	3	41,06	E
E-A	3	38,10	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

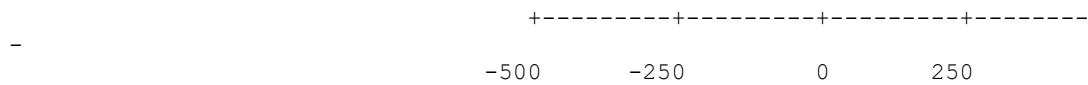
Individual confidence level = 99,68%

Group = E-A subtracted from:

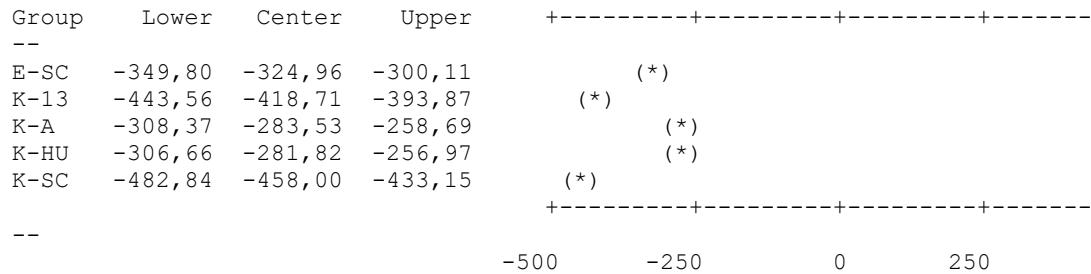
Group	Lower	Center	Upper	
-				-----+-----+-----+
E-Laff	-21,88	2,97	27,81	(*)
E-LT-SC	447,96	472,81	497,65	(*)
(*)				
E-SC	123,01	147,85	172,70	(*)
K-13	29,25	54,10	78,94	(*)
K-A	164,43	189,28	214,12	(*)
K-HU	166,15	190,99	215,84	(*)
K-SC	-10,03	14,81	39,66	(*)
-				-----+-----+-----+
				-500 -250 0 250

Group = E-Laff subtracted from:

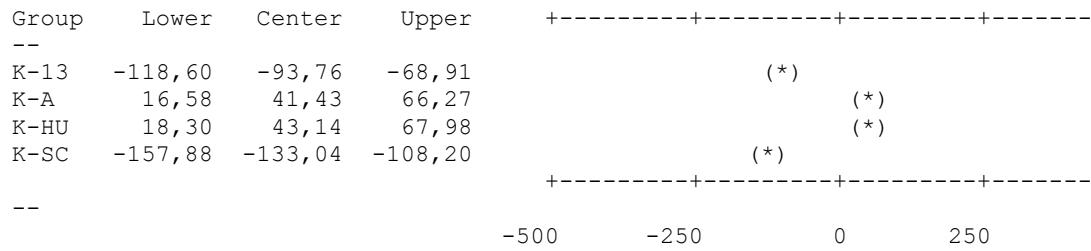
Group	Lower	Center	Upper	
-				-----+-----+-----+
E-LT-SC	445,00	469,84	494,69	(*)
(*)				
E-SC	120,04	144,89	169,73	(*)
K-13	26,29	51,13	75,97	(*)
K-A	161,47	186,31	211,16	(*)
K-HU	163,18	188,03	212,87	(*)
K-SC	-13,00	11,85	36,69	(*)



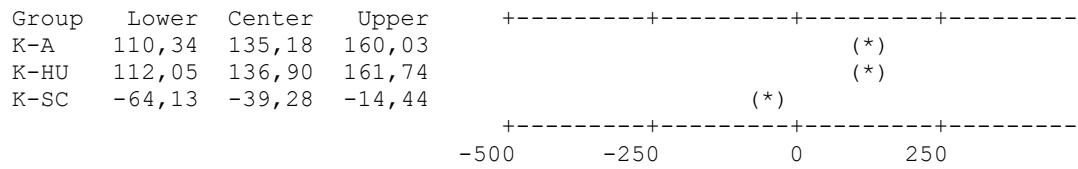
Group = E-LT-SC subtracted from:



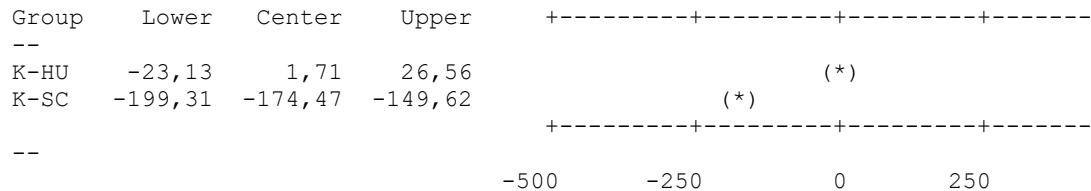
Group = E-SC subtracted from:



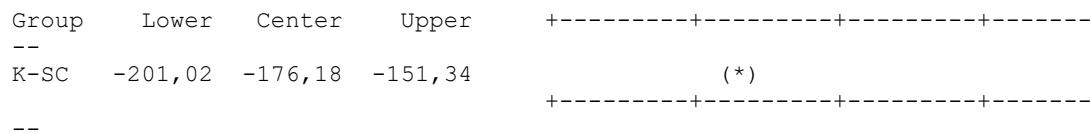
Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:

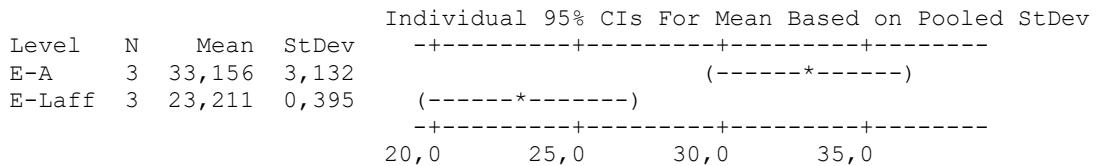


-500 -250 0 250

One-way ANOVA: 3-(Ethylthio)-1-propanol versus Group

Source	DF	SS	MS	F	P
Group	1	148,35	148,35	29,77	0,005
Error	4	19,94	4,98		
Total	5	168,29			

S = 2,232 R-Sq = 88,15% R-Sq(adj) = 85,19%



Pooled StDev = 2,232

Grouping Information Using Tukey Method

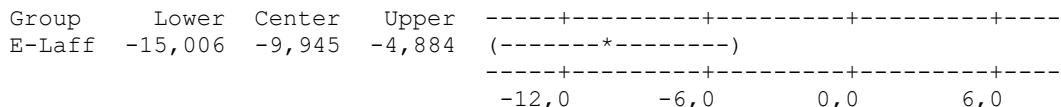
Group	N	Mean	Grouping
E-A	3	33,156	A
E-Laff	3	23,211	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

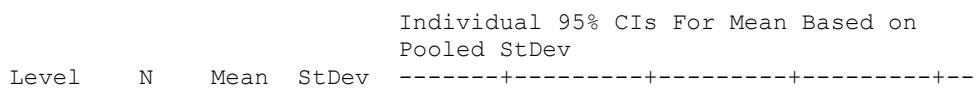
Group = E-A subtracted from:

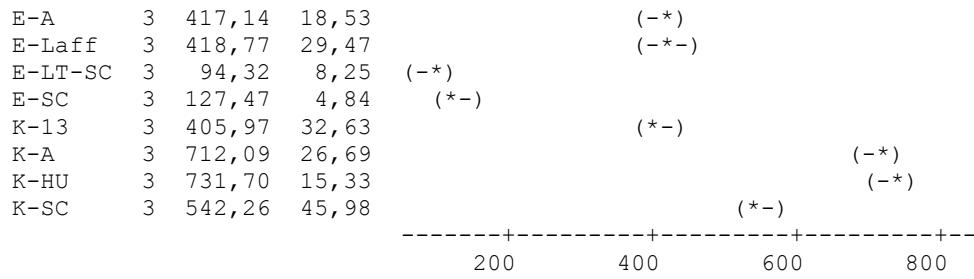


One-way ANOVA: Methionol versus Group

Source	DF	SS	MS	F	P
Group	7	1164781	166397	245,18	0,000
Error	16	10859	679		
Total	23	1175639			

S = 26,05 R-Sq = 99,08% R-Sq(adj) = 98,67%





Pooled StDev = 26,05

Grouping Information Using Tukey Method

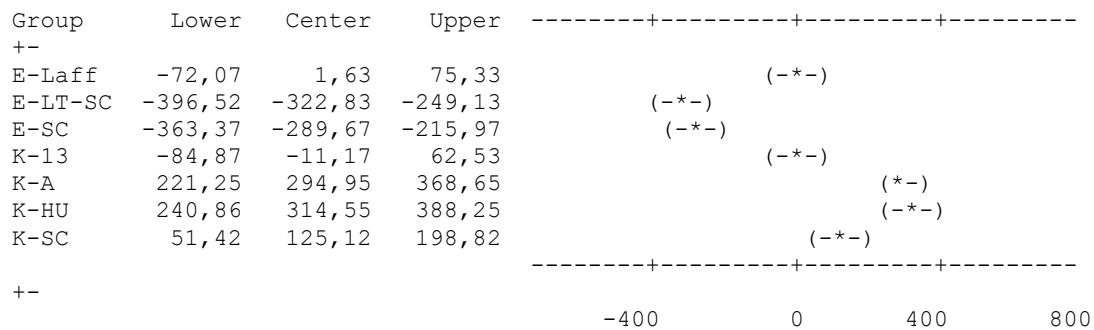
Group	N	Mean	Grouping
K-HU	3	731,70	A
K-A	3	712,09	A
K-SC	3	542,26	B
E-Laff	3	418,77	C
E-A	3	417,14	C
K-13	3	405,97	C
E-SC	3	127,47	D
E-LT-SC	3	94,32	D

Means that do not share a letter are significantly different.

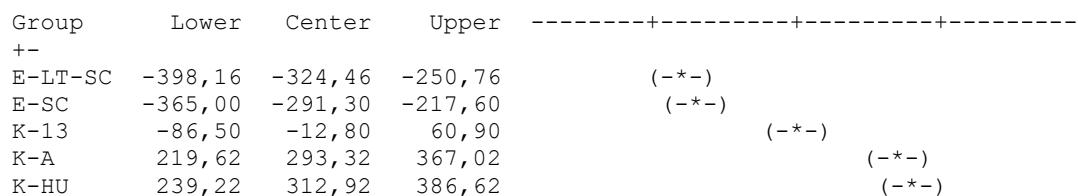
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

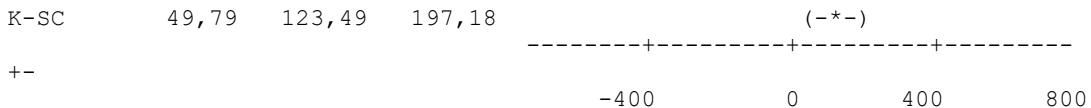
Individual confidence level = 99,68%

Group = E-A subtracted from:

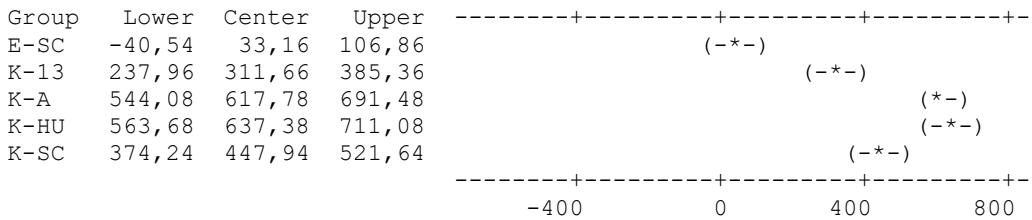


Group = E-Laff subtracted from:

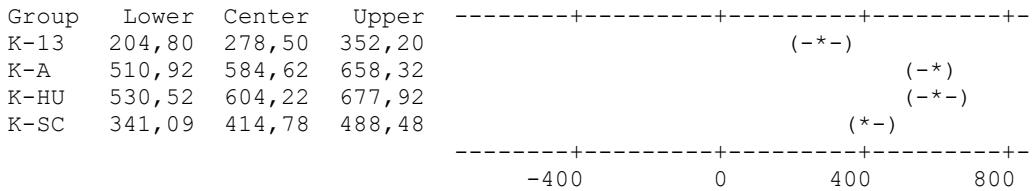




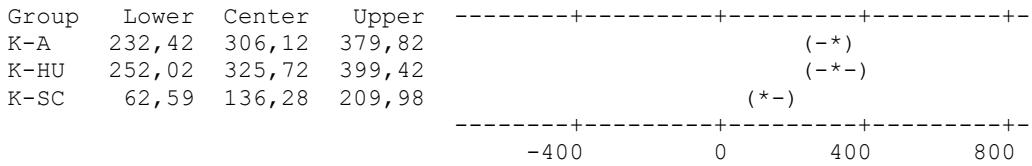
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



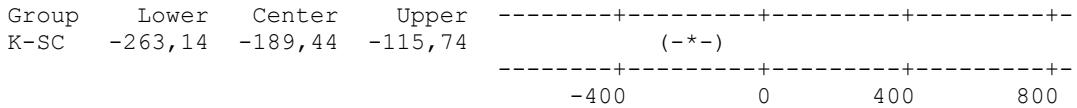
Group = K-13 subtracted from:



Group = K-A subtracted from:



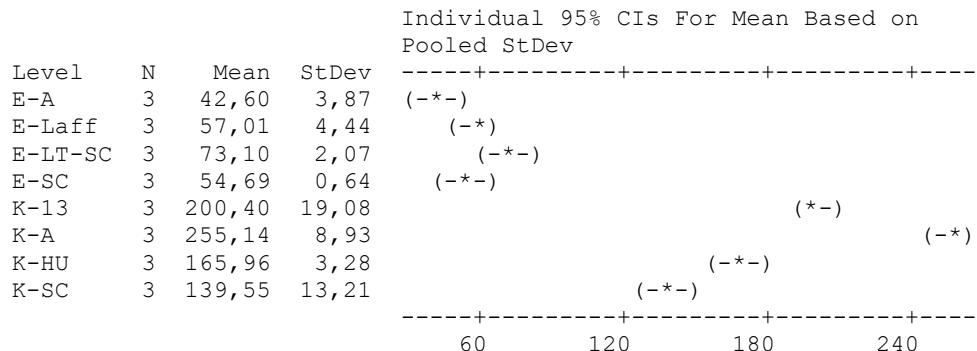
Group = K-HU subtracted from:



One-way ANOVA: Benzyl Alcohol versus Group

Source	DF	SS	MS	F	P
Group	7	130627,5	18661,1	223,31	0,000
Error	16	1337,0	83,6		
Total	23	131964,6			

S = 9,141 R-Sq = 98,99% R-Sq(adj) = 98,54%



Pooled StDev = 9,14

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	255,14	A
K-13	3	200,40	B
K-HU	3	165,96	C
K-SC	3	139,55	D
E-LT-SC	3	73,10	E
E-Laff	3	57,01	E F
E-SC	3	54,69	E F
E-A	3	42,60	F

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

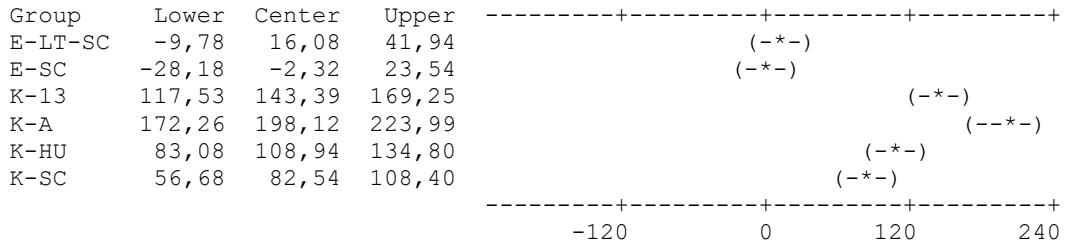
Individual confidence level = 99,68%

Group = E-A subtracted from:

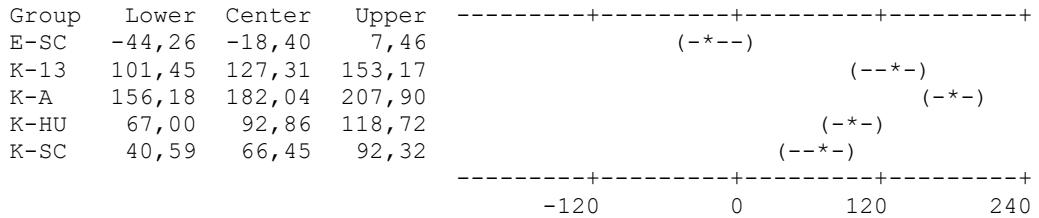
Group	Lower	Center	Upper				
E-Laff	-11,45	14,41	40,27		(-*)		
E-LT-SC	4,63	30,49	56,35		(-*)		
E-SC	-13,77	12,09	37,95		(-*)		
K-13	131,94	157,80	183,66		(-*)		
K-A	186,67	212,53	238,39		(-*)		
K-HU	97,49	123,35	149,21		(-*)		
K-SC	71,08	96,95	122,81		(-*)		

-120 0 120 240

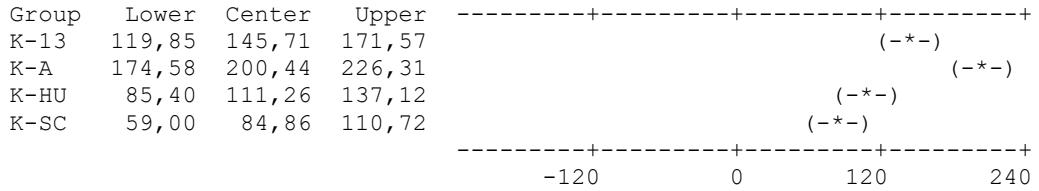
Group = E-Laff subtracted from:



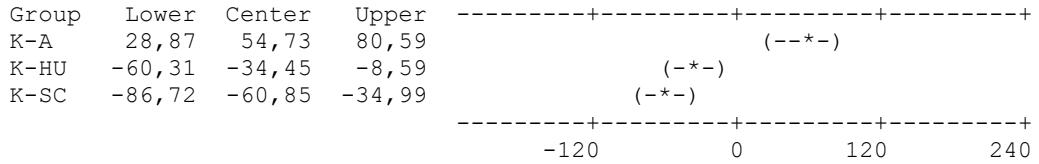
Group = E-LT-SC subtracted from:



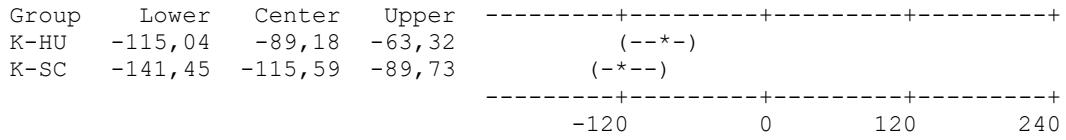
Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



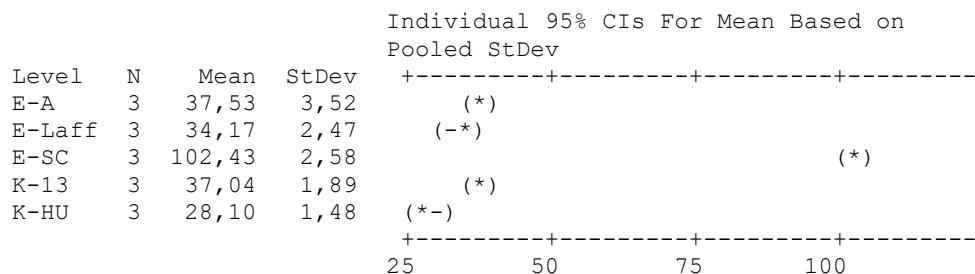
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	-52,27	-26,41	-0,55		(-*)		
				-120	0	120	240

One-way ANOVA: 2-Methyl-2-buten-1-ol versus Group

Source	DF	SS	MS	F	P
Group	4	11339,05	2834,76	458,77	0,000
Error	10	61,79	6,18		
Total	14	11400,84			

S = 2,486 R-Sq = 99,46% R-Sq(adj) = 99,24%



Pooled StDev = 2,49

Grouping Information Using Tukey Method

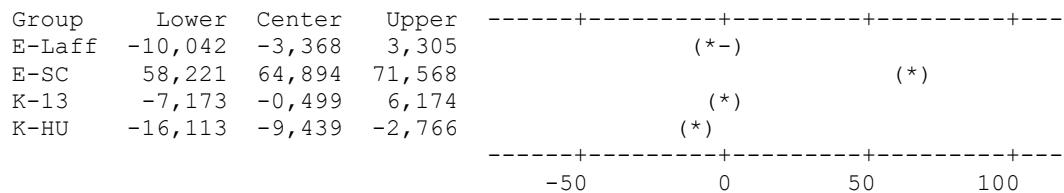
Group	N	Mean	Grouping
E-SC	3	102,429	A
E-A	3	37,535	B
K-13	3	37,036	B
E-Laff	3	34,166	B C
K-HU	3	28,096	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,18%

Group = E-A subtracted from:



Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-SC	61,589	68,262	74,936				(-*)
K-13	-3,804	2,869	9,543				(-*)
K-HU	-12,744	-6,071	0,603				(-*)
				-50	0	50	100

Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	-72,067	-65,393	-58,720	(*)			
K-HU	-81,007	-74,333	-67,660	(*)			
				-50	0	50	100

Group = K-13 subtracted from:

Group	Lower	Center	Upper				
K-HU	-15,614	-8,940	-2,267	(*-)			
				-50	0	50	100

One-way ANOVA: Phenylethyl Alcohol versus Group

Source	DF	SS	MS	F	P
Group	7	785730191	112247170	157,63	0,000
Error	16	11393525	712095		
Total	23	797123716			

S = 843,9 R-Sq = 98,57% R-Sq(adj) = 97,95%

Individual 95% CIs For Mean Based on Pooled StDev						
Level	N	Mean	StDev			
E-A	3	21883	760			(-*)
E-Laff	3	20826	1767			(-*)
E-LT-SC	3	12913	514	(-*)		
E-SC	3	12587	296	(-*)		
K-13	3	21741	511			(-*)
K-A	3	30431	637			(-*)
K-HU	3	26839	527			(-*)
K-SC	3	19790	836			(-*)
				15000	20000	25000
						30000

Pooled StDev = 844

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	30431	A
K-HU	3	26839	B

E-A	3	21883	C
K-13	3	21741	C
E-Laff	3	20826	C
K-SC	3	19790	C
E-LT-SC	3	12913	D
E-SC	3	12587	D

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-3444	-1056	1331	(-*)
E-LT-SC	-11357	-8970	-6582	(-*)
E-SC	-11683	-9295	-6908	(-*)
K-13	-2529	-142	2245	(-*)
K-A	6161	8548	10936	(-*)
K-HU	2569	4957	7344	(-*)
K-SC	-4480	-2093	294	(-*)
				-12000 0 12000 24000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper	
E-LT-SC	-10301	-7913	-5526	(-*)
E-SC	-10626	-8239	-5852	(-*)
K-13	-1473	914	3301	(-*)
K-A	7217	9605	11992	(-*)
K-HU	3625	6013	8400	(-*)
K-SC	-3424	-1037	1351	(-*)
				-12000 0 12000 24000

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	
E-SC	-2713	-326	2061	(-*)
K-13	6440	8827	11215	(-*)
K-A	15131	17518	19905	(-*)
K-HU	11539	13926	16313	(-*)
K-SC	4489	6877	9264	(-*)
				-12000 0 12000 24000

Group = E-SC subtracted from:

Group	Lower	Center	Upper	
K-13	6766	9153	11541	(-*)
K-A	15456	17844	20231	(-*)

K-HU	11865	14252	16639	(-*)
K-SC	4815	7202	9590	(-*)
				-----+-----+-----+-----+-
				-12000 0 12000 24000

Group = K-13 subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-
K-A	6303	8690	11078	(-*)
K-HU	2711	5099	7486	(-*)
K-SC	-4338	-1951	436	(-*)
				-----+-----+-----+-----+-
				-12000 0 12000 24000

Group = K-A subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-
K-HU	-5979	-3592	-1205	(-*)
K-SC	-13029	-10641	-8254	(-*)
				-----+-----+-----+-----+-
				-12000 0 12000 24000

Group = K-HU subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-
K-SC	-9437	-7049	-4662	(-*)
				-----+-----+-----+-----+-
				-12000 0 12000 24000

One-way ANOVA: 3-Methyl-3-butene-1-ol versus Group

Source	DF	SS	MS	F	P
Group	5	8451,5	1690,3	45,85	0,000
Error	12	442,4	36,9		
Total	17	8893,9			

S = 6,072 R-Sq = 95,03% R-Sq(adj) = 92,95%

Individual 95% CIs For Mean Based on Pooled StDev					
Level	N	Mean	StDev	-----+-----+-----+-----+-	
E-A	3	88,65	5,40	(---*---)
E-Laff	3	111,31	7,95	(---*---)
K-13	3	90,62	8,94	(---*---)
K-A	3	134,05	1,19		(---*---
K-HU	3	74,35	5,26	(---*---)
K-SC	3	71,75	4,45	(---*---)
				-----+-----+-----+-----+-	
				80 100 120 140	

Pooled StDev = 6,07

Grouping Information Using Tukey Method

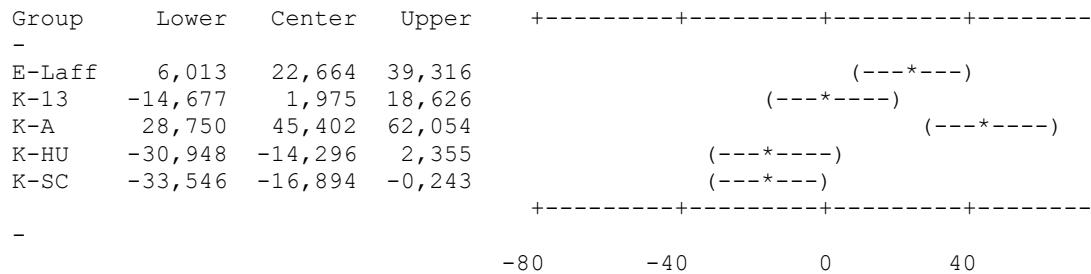
Group	N	Mean	Grouping
K-A	3	134,048	A
E-Laff	3	111,311	B
K-13	3	90,621	C
E-A	3	88,646	C
K-HU	3	74,350	C D
K-SC	3	71,752	D

Means that do not share a letter are significantly different.

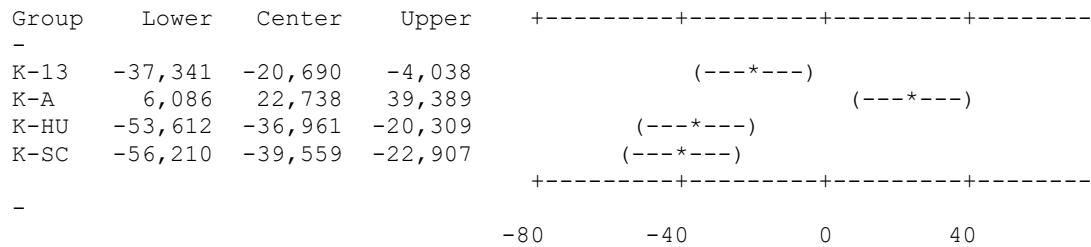
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

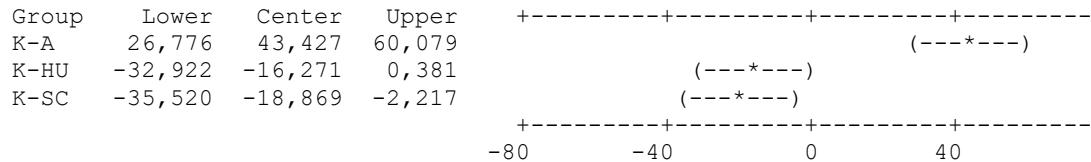
Group = E-A subtracted from:



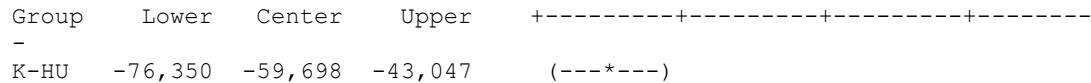
Group = E-Laff subtracted from:

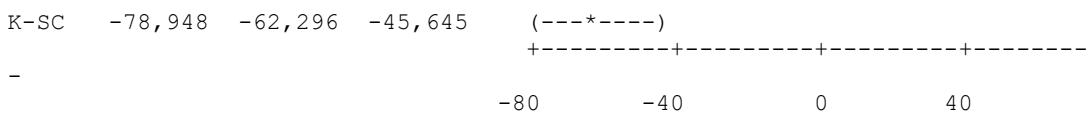


Group = K-13 subtracted from:

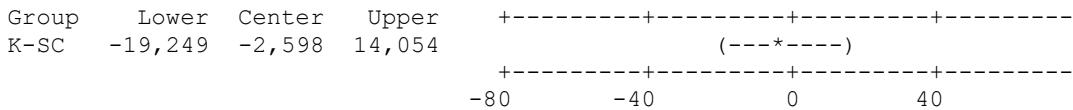


Group = K-A subtracted from:





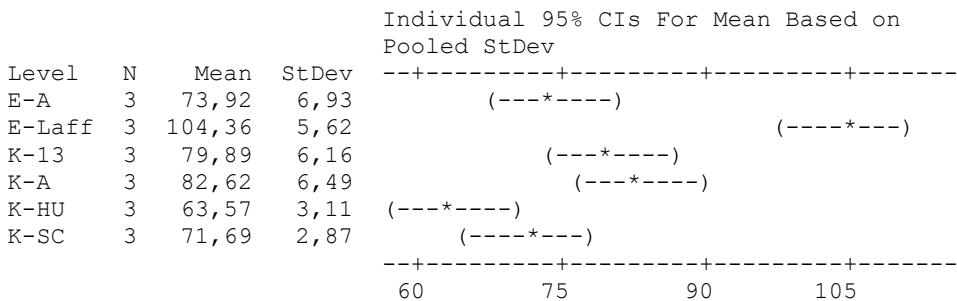
Group = K-HU subtracted from:



One-way ANOVA: 1-Pentanol versus Group

Source	DF	SS	MS	F	P
Group	5	2920,7	584,1	19,73	0,000
Error	12	355,3	29,6		
Total	17	3276,1			

S = 5,442 R-Sq = 89,15% R-Sq(adj) = 84,63%



Pooled StDev = 5,44

Grouping Information Using Tukey Method

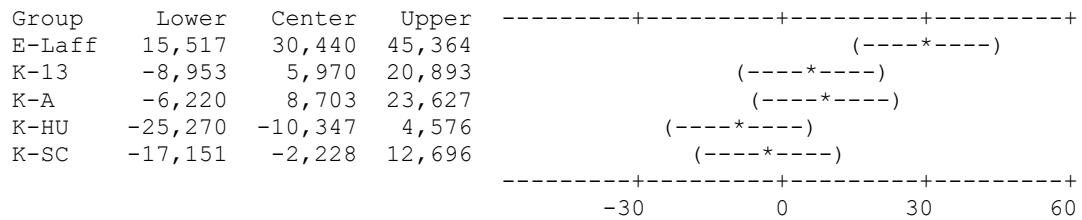
Group	N	Mean	Grouping
E-Laff	3	104,356	A
K-A	3	82,619	B
K-13	3	79,886	B
E-A	3	73,916	B C
K-SC	3	71,688	B C
K-HU	3	63,569	C

Means that do not share a letter are significantly different.

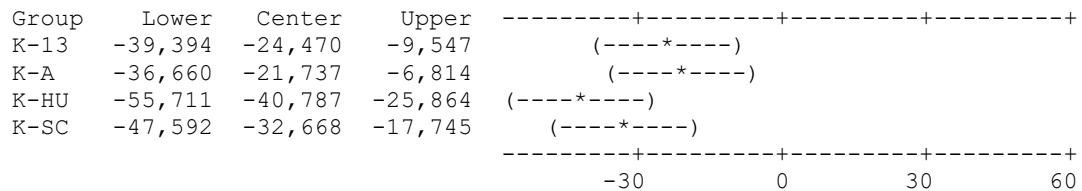
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

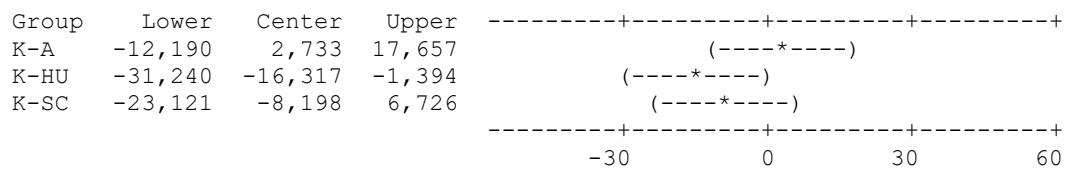
Group = E-A subtracted from:



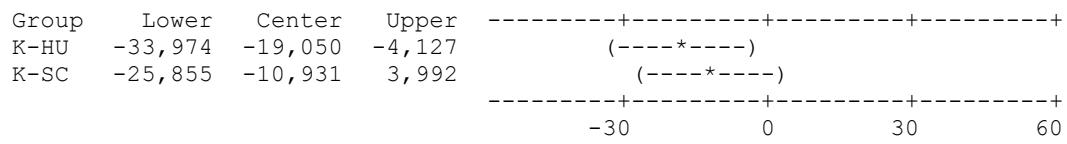
Group = E-Laff subtracted from:



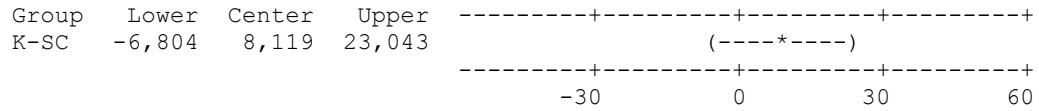
Group = K-13 subtracted from:



Group = K-A subtracted from:



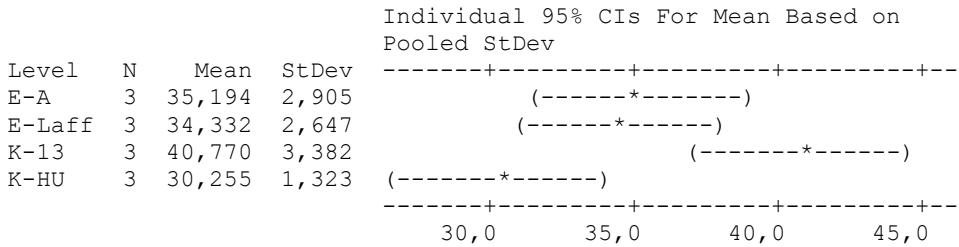
Group = K-HU subtracted from:



One-way ANOVA: 4-Methyl-1-pentanol versus Group

Source	DF	SS	MS	F	P
Group	3	168,66	56,22	7,85	0,009
Error	8	57,26	7,16		
Total	11	225,92			

S = 2,675 R-Sq = 74,65% R-Sq(adj) = 65,15%



Pooled StDev = 2,675

Grouping Information Using Tukey Method

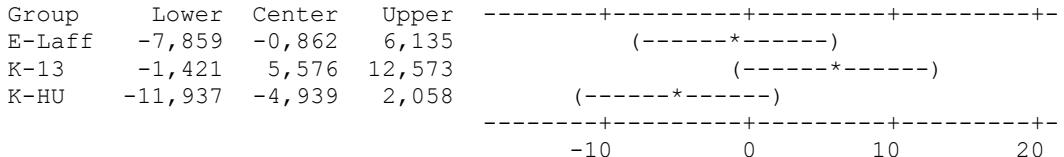
Group	N	Mean	Grouping
K-13	3	40,770	A
E-A	3	35,194	A B
E-Laff	3	34,332	A B
K-HU	3	30,255	B

Means that do not share a letter are significantly different.

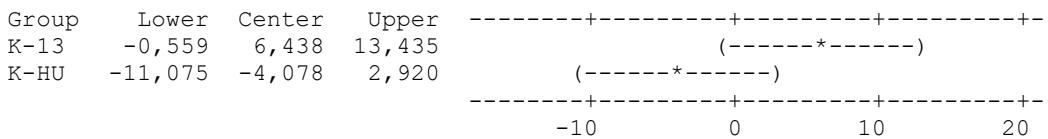
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

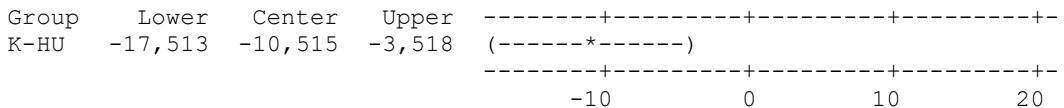
Group = E-A subtracted from:



Group = E-Laff subtracted from:



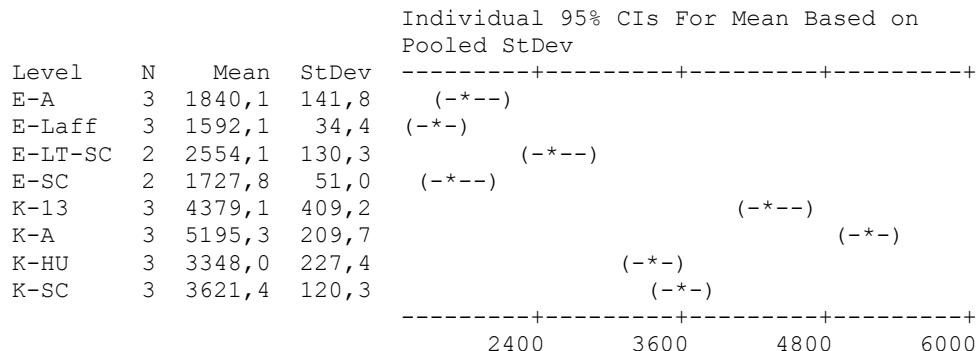
Group = K-13 subtracted from:



One-way ANOVA: 2,3-Butanediol versus Group

Source	DF	SS	MS	F	P
Group	7	35020872	5002982	113,44	0,000
Error	14	617411	44101		
Total	21	35638284			

S = 210,0 R-Sq = 98,27% R-Sq(adj) = 97,40%



Pooled StDev = 210,0

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	5195,3	A
K-13	3	4379,1	B
K-SC	3	3621,4	C
K-HU	3	3348,0	C
E-LT-SC	2	2554,1	D
E-A	3	1840,1	E
E-SC	2	1727,8	E
E-Laff	3	1592,1	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

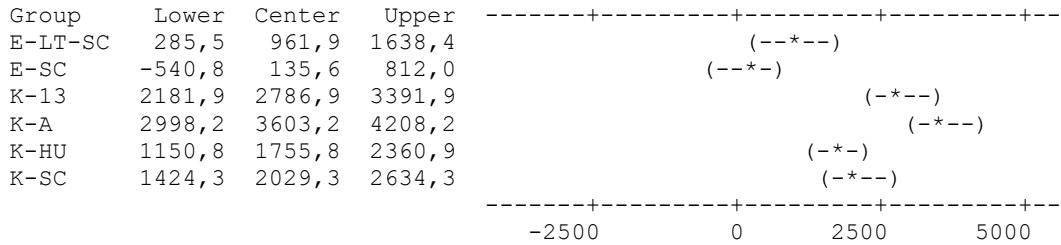
Individual confidence level = 99,67%

Group = E-A subtracted from:

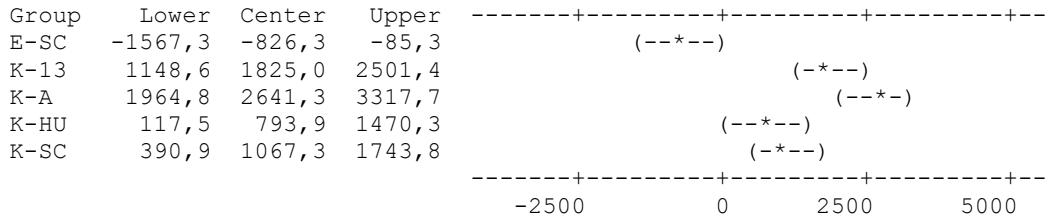
Group	Lower	Center	Upper			
E-Laff	-853,0	-247,9	357,1	(-*)		
E-LT-SC	37,6	714,0	1390,4		(-*)	
E-SC	-788,7	-112,3	564,1			(-*)
K-13	1934,0	2539,0	3144,0			(-*)
K-A	2750,2	3355,3	3960,3			(-*)
K-HU	902,9	1507,9	2112,9		(-*)	
K-SC	1176,3	1781,3	2386,3			(-*)

-2500 0 2500 5000

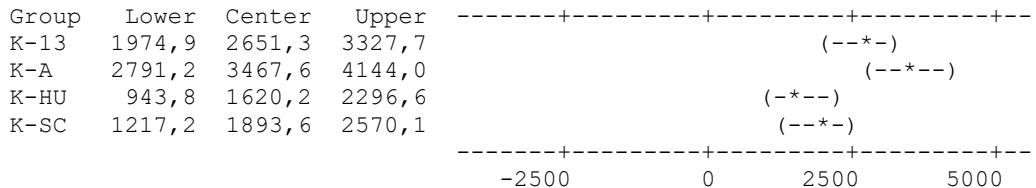
Group = E-Laff subtracted from:



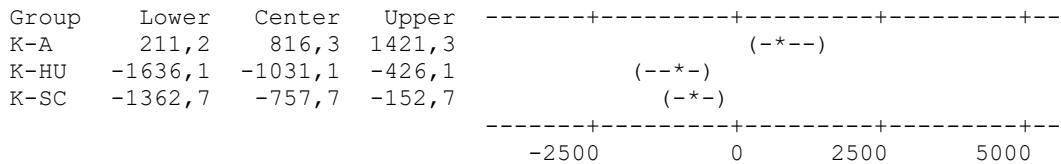
Group = E-LT-SC subtracted from:



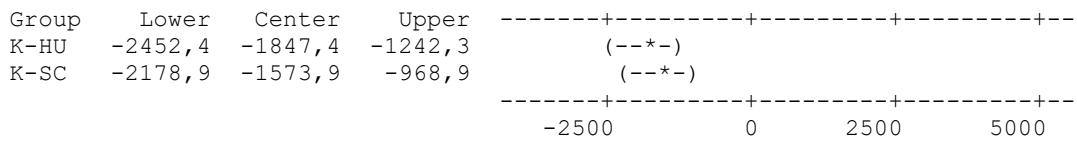
Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



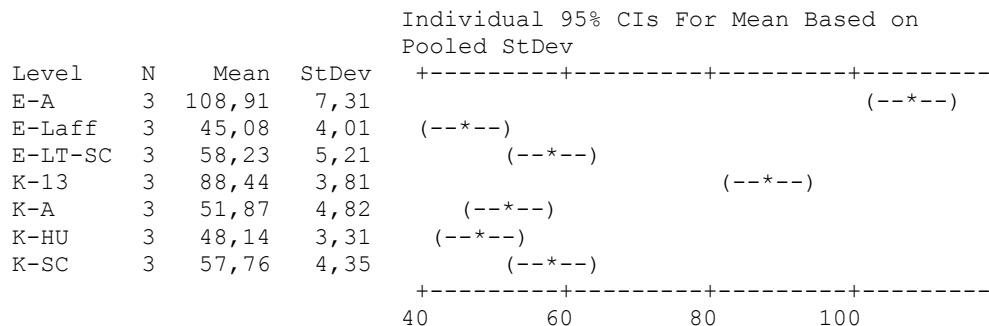
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	-331,6	273,4	878,4		(-*)--)		
	-2500	0	2500			5000	

One-way ANOVA: 1-Heptanol versus Group

Source	DF	SS	MS	F	P
Group	6	10284,0	1714,0	73,05	0,000
Error	14	328,5	23,5		
Total	20	10612,5			

S = 4,844 R-Sq = 96,90% R-Sq(adj) = 95,58%



Pooled StDev = 4,84

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-A	3	108,914	A
K-13	3	88,444	B
E-LT-SC	3	58,229	C
K-SC	3	57,757	C
K-A	3	51,871	C
K-HU	3	48,142	C
E-Laff	3	45,080	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,58%

Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-77,342	-63,834	-50,326	(---*---)
E-LT-SC	-64,192	-50,685	-37,177	(---*---)
K-13	-33,978	-20,470	-6,962	(---*---)

K-A	-70,551	-57,043	-43,535	(---*---
K-HU	-74,279	-60,771	-47,263	(---*--)
K-SC	-64,665	-51,157	-37,649	(--*---
				-----+-----+-----+-----
-				-70 -35 0 35

Group = E-Laff subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----
E-LT-SC	-0,358	13,149	26,657	(---*---
K-13	29,856	43,364	56,872	(--*---
K-A	-6,717	6,791	20,299	(---*---
K-HU	-10,445	3,063	16,570	(---*---
K-SC	-0,831	12,677	26,185	(---*--)
				-----+-----+-----+-----
				-70 -35 0 35

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-----
K-13	16,707	30,214	43,722	(---*--)
K-A	-19,866	-6,359	7,149	(---*---
K-HU	-23,595	-10,087	3,421	(---*---
K-SC	-13,980	-0,472	13,035	(---*---
				-----+-----+-----+-----+-----
				-70 -35 0 35

Group = K-13 subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-----
K-A	-50,081	-36,573	-23,065	(---*--)
K-HU	-53,809	-40,301	-26,793	(--*---
K-SC	-44,195	-30,687	-17,179	(---*---
				-----+-----+-----+-----+-----
				-70 -35 0 35

Group = K-A subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-----
K-HU	-17,236	-3,728	9,780	(---*--)
K-SC	-7,622	5,886	19,394	(---*--)
				-----+-----+-----+-----+-----
				-70 -35 0 35

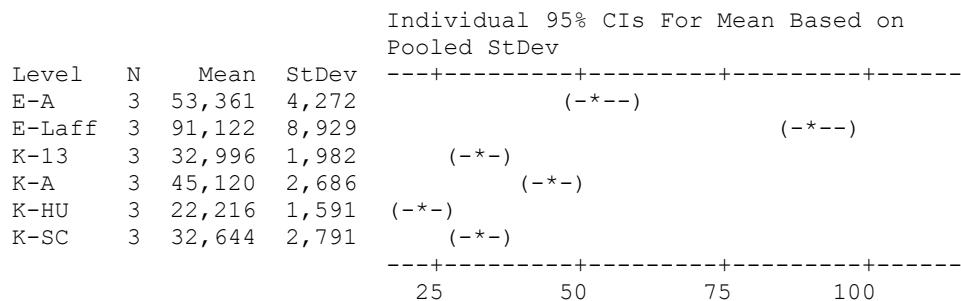
Group = K-HU subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+-----
K-SC	-3,894	9,614	23,122	(---*--)
				-----+-----+-----+-----+-----
				-70 -35 0 35

One-way ANOVA: (Z)-3-Hexene-1-ol versus Group

Source	DF	SS	MS	F	P
Group	5	9011,3	1802,3	90,54	0,000
Error	12	238,9	19,9		
Total	17	9250,2			

S = 4,462 R-Sq = 97,42% R-Sq(adj) = 96,34%



Pooled StDev = 4,462

Grouping Information Using Tukey Method

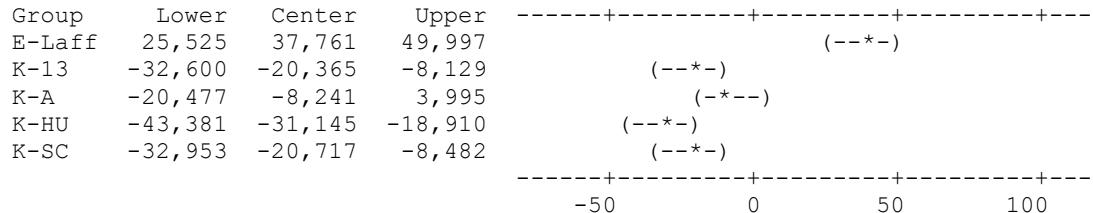
Group	N	Mean	Grouping
E-Laff	3	91,122	A
E-A	3	53,361	B
K-A	3	45,120	B C
K-13	3	32,996	C D
K-SC	3	32,644	D
K-HU	3	22,216	D

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

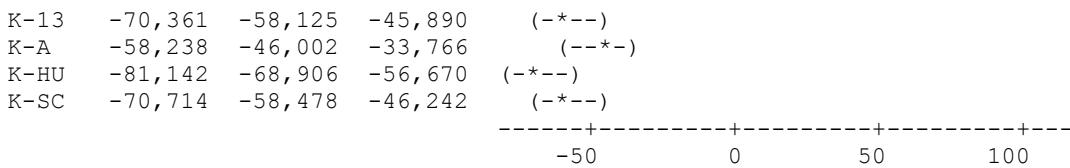
Individual confidence level = 99,43%

Group = E-A subtracted from:

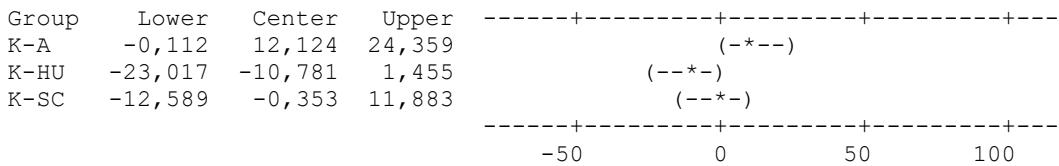


Group = E-Laff subtracted from:

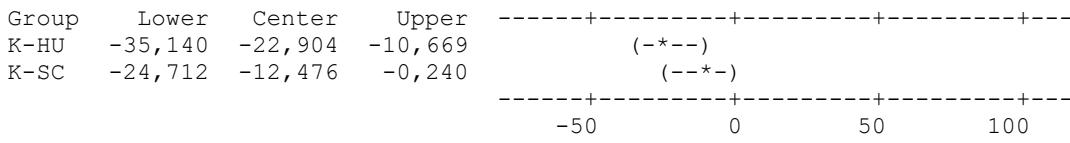
Group Lower Center Upper -----+-----+-----+-----+



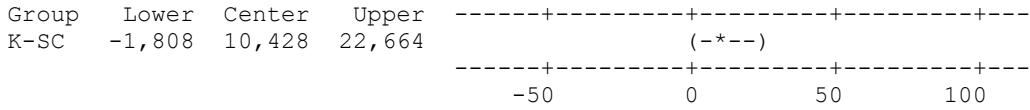
Group = K-13 subtracted from:



Group = K-A subtracted from:



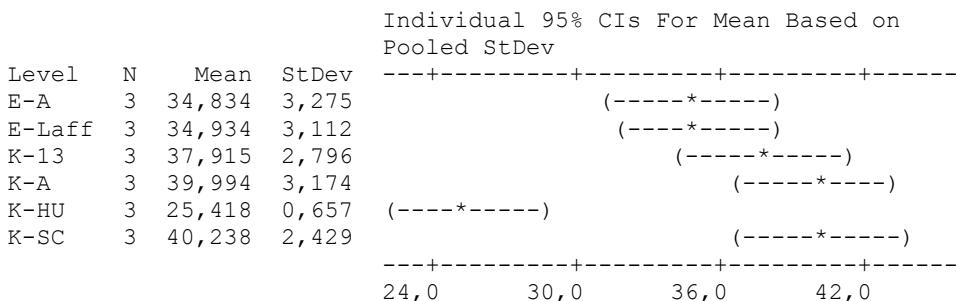
Group = K-HU subtracted from:



One-way ANOVA: (E)-3-Hexene-1-ol versus Group

Source	DF	SS	MS	F	P
Group	5	452, 61	90, 52	12, 17	0, 000
Error	12	89, 26	7, 44		
Total	17	541, 87			

S = 2,727 R-Sq = 83,53% R-Sq(adj) = 76,66%



Pooled StDev = 2,727

Grouping Information Using Tukey Method

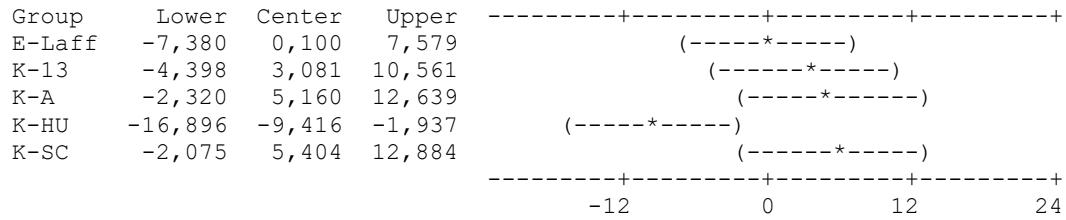
Group	N	Mean	Grouping
K-SC	3	40,238	A
K-A	3	39,994	A
K-13	3	37,915	A
E-Laff	3	34,934	A
E-A	3	34,834	A
K-HU	3	25,418	B

Means that do not share a letter are significantly different.

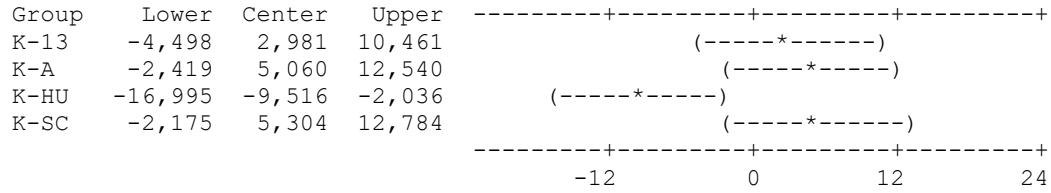
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

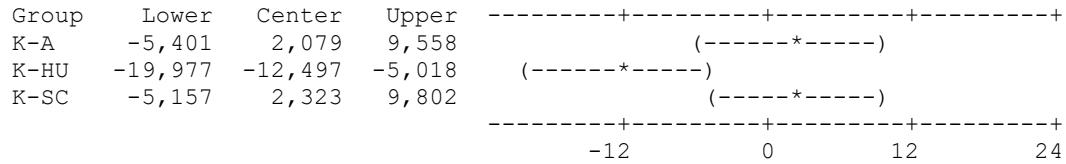
Group = E-A subtracted from:



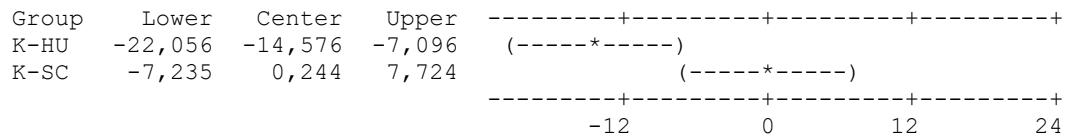
Group = E-Laff subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



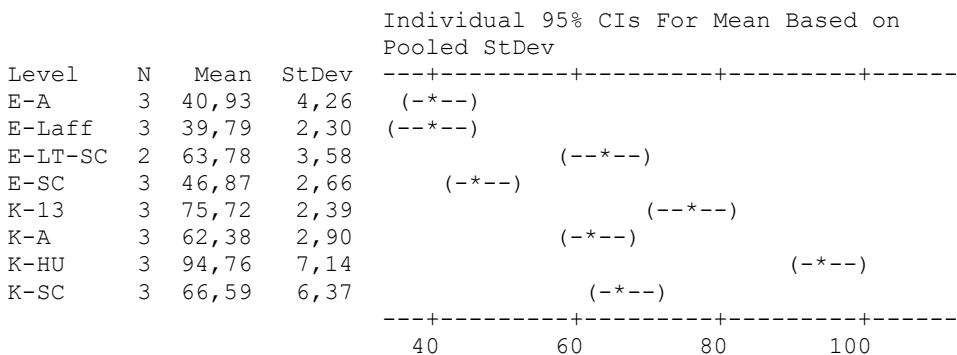
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	7,341	14,820	22,300				(-----*-----)
				-12	0	12	24

One-way ANOVA: 2-(Methylthio)ethanol versus Group

Source	DF	SS	MS	F	P
Group	7	7339,7	1048,5	55,14	0,000
Error	15	285,3	19,0		
Total	22	7625,0			

S = 4,361 R-Sq = 96,26% R-Sq(adj) = 94,51%



Pooled StDev = 4,36

Grouping Information Using Tukey Method

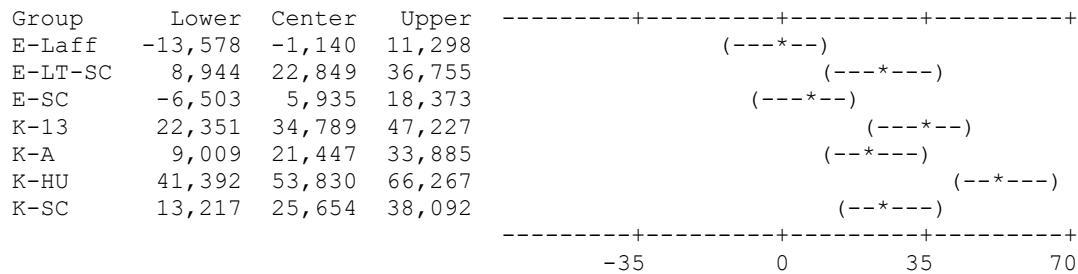
Group	N	Mean	Grouping
K-HU	3	94,764	A
K-13	3	75,723	B
K-SC	3	66,589	B C
E-LT-SC	2	63,784	B C
K-A	3	62,381	C
E-SC	3	46,870	D
E-A	3	40,934	D
E-Laff	3	39,795	D

Means that do not share a letter are significantly different.

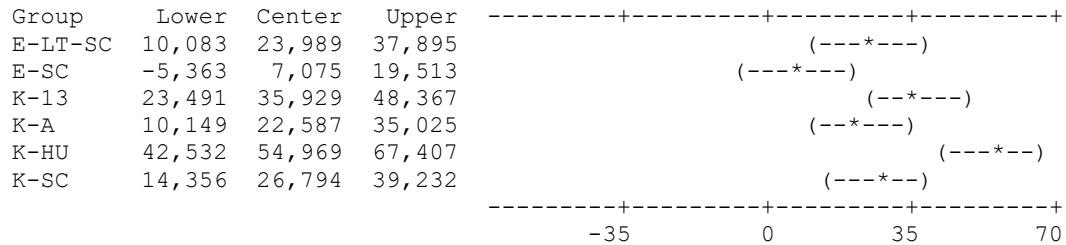
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

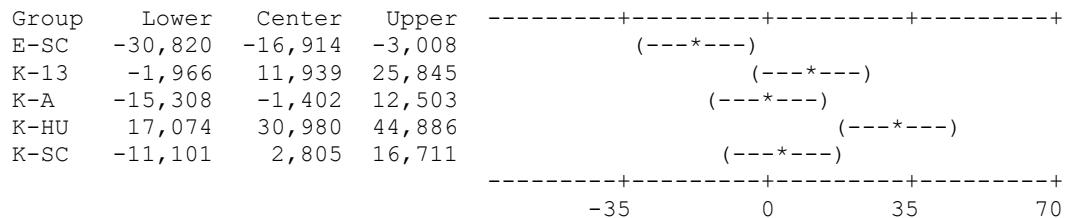
Group = E-A subtracted from:



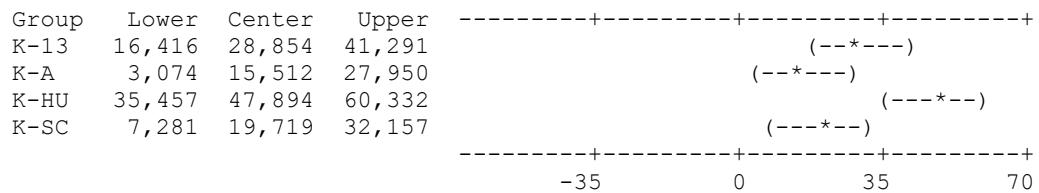
Group = E-Laff subtracted from:



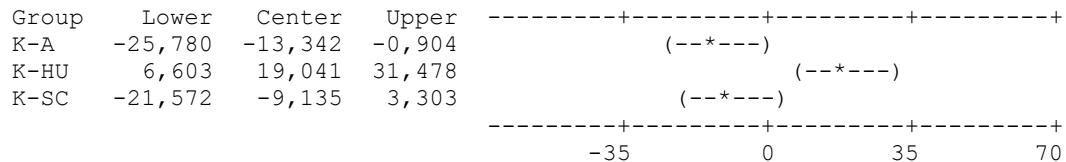
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



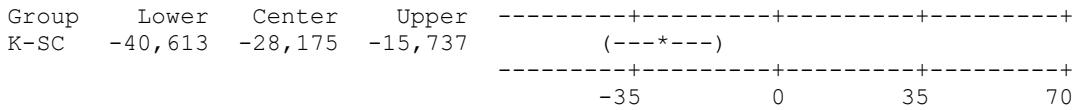
Group = K-13 subtracted from:



Group = K-A subtracted from:



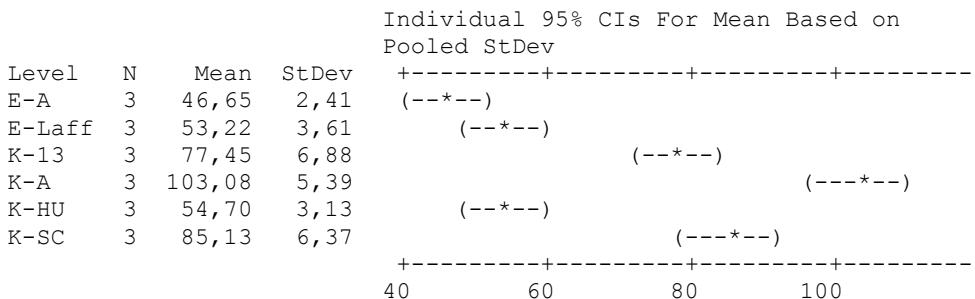
Group = K-HU subtracted from:



One-way ANOVA: 1,2-Propanediol versus Group

Source	DF	SS	MS	F	P
Group	5	7317,8	1463,6	60,34	0,000
Error	12	291,1	24,3		
Total	17	7608,9			

S = 4,925 R-Sq = 96,17% R-Sq(adj) = 94,58%



Pooled StDev = 4,93

Grouping Information Using Tukey Method

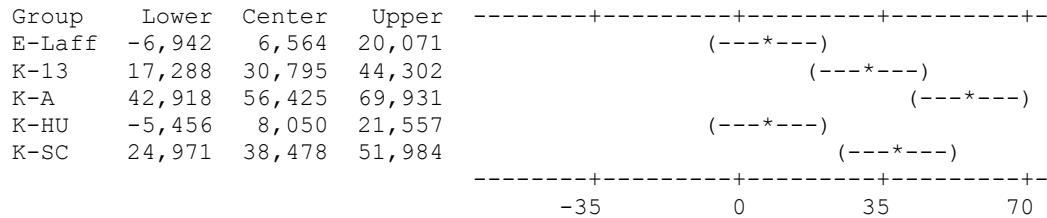
Group	N	Mean	Grouping
K-A	3	103,077	A
K-SC	3	85,129	B
K-13	3	77,447	B
K-HU	3	54,702	C
E-Laff	3	53,216	C
E-A	3	46,652	C

Means that do not share a letter are significantly different.

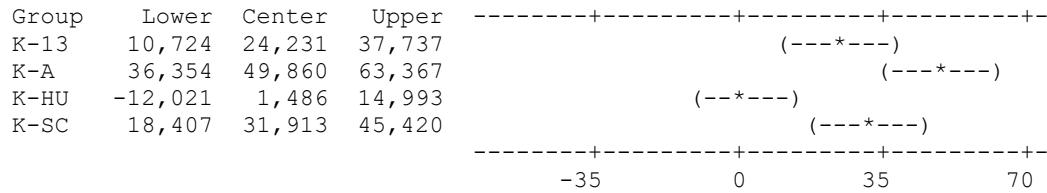
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

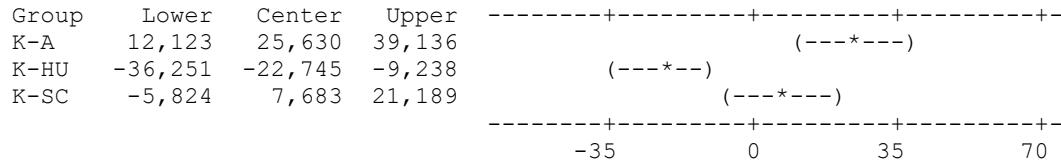
Group = E-A subtracted from:



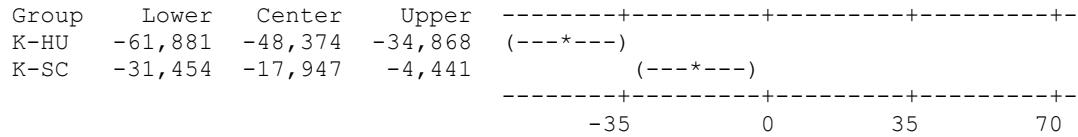
Group = E-Laff subtracted from:



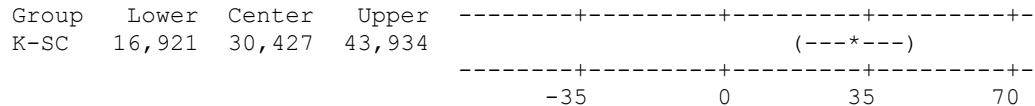
Group = K-13 subtracted from:



Group = K-A subtracted from:



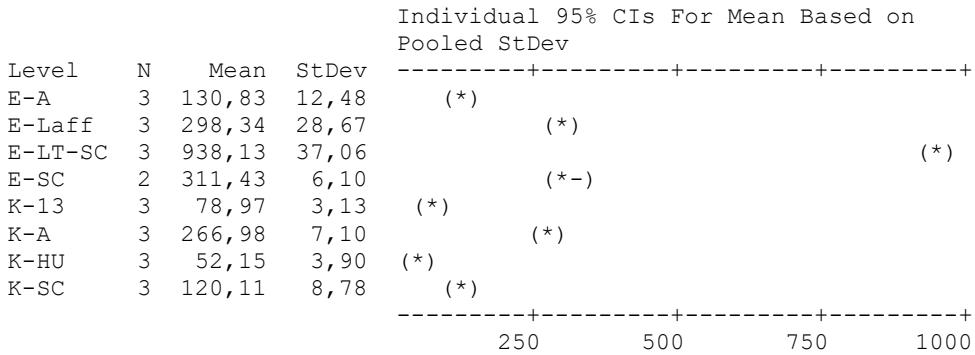
Group = K-HU subtracted from:



One-way ANOVA: 3-Penten-2-ol versus Group

Source	DF	SS	MS	F	P
Group	7	1722234	246033	731,52	0,000
Error	15	5045	336		
Total	22	1727278			

S = 18,34 R-Sq = 99,71% R-Sq(adj) = 99,57%



Pooled StDev = 18,34

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	938,13	A
E-SC	2	311,43	B
E-Laff	3	298,34	B
K-A	3	266,98	B
E-A	3	130,83	C
K-SC	3	120,11	C
K-13	3	78,97	C D
K-HU	3	52,15	D

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	115,21	167,51	219,82			(*)	
E-LT-SC	755,00	807,31	859,61				(*)
E-SC	122,12	180,60	239,08			(-*)	
K-13	-104,16	-51,86	0,45			(*)	
K-A	83,85	136,15	188,46			(*)	
K-HU	-130,99	-78,68	-26,37			(*)	
K-SC	-63,03	-10,72	41,59			(*)	

-----+-----+-----+-----+
 -500 0 500 1000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper			
E-LT-SC	587,49	639,79	692,10			(*)
E-SC	-45,39	13,09	71,57			(*)
K-13	-271,67	-219,37	-167,06			(*)

K-A	-83, 67	-31, 36	20, 94	(*)
K-HU	-298, 50	-246, 19	-193, 89	(*)
K-SC	-230, 54	-178, 23	-125, 93	(*)
	-----+-----+-----+-----			
+	-500 0 500			
1000				

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-685, 19	-626, 71	-568, 23		(*-)		
K-13	-911, 47	-859, 16	-806, 86	(*)			
K-A	-723, 46	-671, 16	-618, 85	(*)			
K-HU	-938, 29	-885, 99	-833, 68	(*)			
K-SC	-870, 33	-818, 03	-765, 72	(*)			

Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	-290, 93	-232, 45	-173, 97		(*)		
K-A	-102, 93	-44, 45	14, 03		(*)		
K-HU	-317, 76	-259, 28	-200, 80		(*)		
K-SC	-249, 80	-191, 32	-132, 84		(*)		
				-500	0	500	1000

Group = K-13 subtracted from:

Group	Lower	Center	Upper				
K-A	135,70	188,01	240,31			(*)	
K-HU	-79,13	-26,82	25,48			(*-)	
K-SC	-11,17	41,14	93,44			(*)	
				-500	0	500	1000

Group = K-A subtracted from:

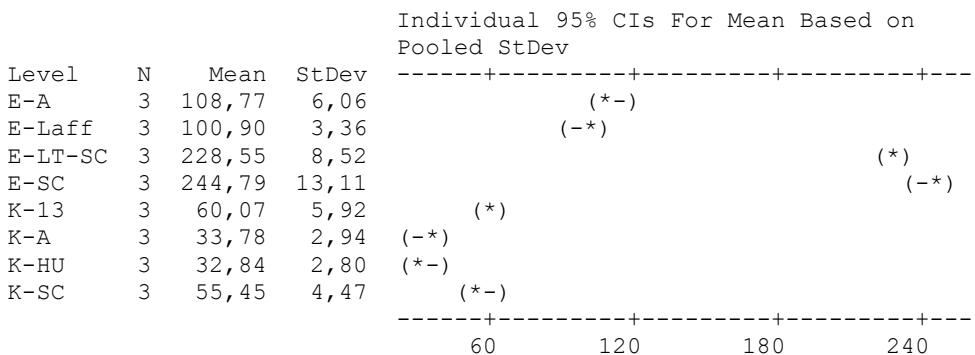
Group	Lower	Center	Upper	(*)		
K-HU	-267,14	-214,83	-162,52			
K-SC	-199,18	-146,87	-94,57	(*)		
			-500	0	500	1000

Group = K-HU subtracted from:

One-way ANOVA: 2-Hexanol versus Group

Source	DF	SS	MS	F	P
Group	7	148534,5	21219,2	466,16	0,000
Error	16	728,3	45,5		
Total	23	149262,8			

S = 6,747 R-Sq = 99,51% R-Sq(adj) = 99,30%



Pooled StDev = 6,75

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-SC	3	244,79	A
E-LT-SC	3	228,55	A
E-A	3	108,77	B
E-Laff	3	100,90	B
K-13	3	60,07	C
K-SC	3	55,45	C
K-A	3	33,78	D
K-HU	3	32,84	D

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	-26,95	-7,86	11,22		(*-)		
E-LT-SC	100,70	119,78	138,87			(-*)	
E-SC	116,93	136,02	155,11			(*-)	
K-13	-67,79	-48,70	-29,61	(-*)			
K-A	-94,08	-74,99	-55,91	(-*)			
K-HU	-95,01	-75,92	-56,84	(-*)			
K-SC	-72,41	-53,32	-34,23	(-*)			

-120 0 120 240

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	108, 56	127, 65	146, 74				(-*)
E-SC	124, 80	143, 88	162, 97				(-* -)
K-13	-59, 92	-40, 84	-21, 75				(-*)
K-A	-86, 22	-67, 13	-48, 04				(*-)
K-HU	-87, 15	-68, 06	-48, 97				(*-)
K-SC	-64, 54	-45, 46	-26, 37				(*-)
				-120	0	120	240

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-2, 85	16, 23	35, 32				(*-)
K-13	-187, 57	-168, 49	-149, 40				(-* -)
K-A	-213, 86	-194, 78	-175, 69				(-*)
K-HU	-214, 80	-195, 71	-176, 62				(-*)
K-SC	-192, 19	-173, 10	-154, 02				(-*)
				-120	0	120	240

Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	-203, 81	-184, 72	-165, 63				(-*)
K-A	-230, 10	-211, 01	-191, 93				(*-)
K-HU	-231, 03	-211, 94	-192, 86				(*-)
K-SC	-208, 42	-189, 34	-170, 25				(*-)
				-120	0	120	240

Group = K-13 subtracted from:

Group	Lower	Center	Upper				
K-A	-45, 38	-26, 29	-7, 21				(-*)
K-HU	-46, 31	-27, 22	-8, 14				(-*)
K-SC	-23, 70	-4, 62	14, 47				(-*)
				-120	0	120	240

Group = K-A subtracted from:

Group	Lower	Center	Upper				
K-HU	-20, 02	-0, 93	18, 16				(-* -)
K-SC	2, 59	21, 67	40, 76				(-*)
				-120	0	120	240

Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	3,52	22,60	41,69		(-*)		
				-120	0	120	240

One-way ANOVA: 1-Hexanol versus Group

Source	DF	SS	MS	F	P
Group	1	934	934	9,12	0,094
Error	2	205	102		
Total	3	1139			

S = 10,12 R-Sq = 82,01% R-Sq(adj) = 73,01%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev				
E-LT-SC	2	191,67	3,30		(-----*-----)		
E-SC	2	161,11	13,93		(-----*-----)		
				150	175	200	225

Pooled StDev = 10,12

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	2	191,67	A
E-SC	2	161,11	A

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-74,10	-30,56	12,99		(-----*-----)		
				-60	-30	0	30

One-way ANOVA: 3-Hexanol versus Group

Source	DF	SS	MS	F	P
Group	1	1329,547	1329,547	1348,79	0,000
Error	4	3,943	0,986		
Total	5	1333,490			

S = 0,9928 R-Sq = 99,70% R-Sq(adj) = 99,63%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev				
E-A	3	22,146	1,056	(*)			
E-Laff	3	51,917	0,925		(-*)		
				30	40	50	60

Pooled StDev = 0,993

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-Laff	3	51,917	A
E-A	3	22,146	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	27,521	29,772	32,023		(-*)		
	0	10	20	30			

One-way ANOVA: 2-Phenoxyethanol versus Group

Source	DF	SS	MS	F	P
Group	1	215,79	215,79	102,76	0,001
Error	4	8,40	2,10		
Total	5	224,19			

S = 1,449 R-Sq = 96,25% R-Sq(adj) = 95,32%

Individual 95% CIs For Mean Based on Pooled StDev

Level	N	Mean	StDev				
E-A	3	34,423	1,381		(----*)----		
E-Laff	3	22,429	1,514	(----*----)			
				20,0	25,0	30,0	35,0

Pooled StDev = 1,449

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-A	3	34,423	A

E-Laff 3 22,429 B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

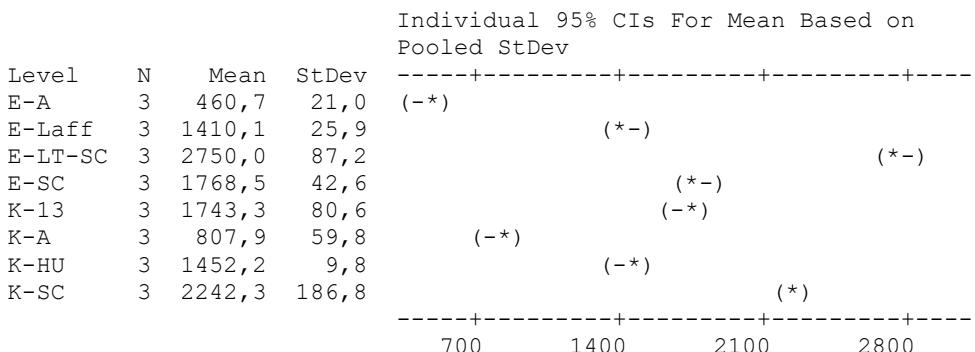
Group = E-A subtracted from:

Group	Lower	Center	Upper				
--							
E-Laff	-15,279	-11,994	-8,709	(-----*-----)			
--				-----+-----+			
				-15,0	-10,0	-5,0	0,0

One-way ANOVA: Isoamyl acetate versus Group

Source	DF	SS	MS	F	P
Group	7	11292154	1613165	232,09	0,000
Error	16	111208	6951		
Total	23	11403362			

S = 83,37 R-Sq = 99,02% R-Sq(adj) = 98,60%



Pooled StDev = 83,4

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	2750,0	A
K-SC	3	2242,3	B
E-SC	3	1768,5	C
K-13	3	1743,3	C
K-HU	3	1452,2	D
E-Laff	3	1410,1	D
K-A	3	807,9	E
E-A	3	460,7	F

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	713,6	949,4	1185,3		(-*)		
E-LT-SC	2053,5	2289,3	2525,2			(-*)	
E-SC	1072,0	1307,9	1543,7			(-*)	
K-13	1046,8	1282,7	1518,5			(-*)	
K-A	111,4	347,2	583,1			(-*)	
K-HU	755,6	991,5	1227,4			(-*)	
K-SC	1545,8	1781,6	2017,5			(-*)	
				-1200	0	1200	2400

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	1104,0	1339,9	1575,7			(-*)	
E-SC	122,6	358,4	594,3			(-*)	
K-13	97,4	333,2	569,1			(-*)	
K-A	-838,1	-602,2	-366,4			(-*)	
K-HU	-193,8	42,0	277,9			(-*)	
K-SC	596,3	832,2	1068,1			(-*)	
				-1200	0	1200	2400

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-1217,3	-981,5	-745,6			(-*)	
K-13	-1242,5	-1006,7	-770,8			(-*)	
K-A	-2178,0	-1942,1	-1706,3			(-*)	
K-HU	-1533,7	-1297,8	-1062,0			(-*)	
K-SC	-743,5	-507,7	-271,8			(-*)	
				-1200	0	1200	2400

Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	-261,1	-25,2	210,7			(-*)	
K-A	-1196,5	-960,7	-724,8			(-*)	
K-HU	-552,2	-316,4	-80,5			(-*)	
K-SC	237,9	473,8	709,6			(-*)	
				-1200	0	1200	2400

Group = K-13 subtracted from:

Group	Lower	Center	Upper				
K-A	-1171,3	-935,5	-699,6		(-*)		
K-HU	-527,0	-291,2	-55,3		(-*)		
K-SC	263,1	499,0	734,8		(-*)		
				-1200	0	1200	2400

Group = K-A subtracted from:

Group	Lower	Center	Upper				
K-HU	408,4	644,3	880,1		(-*)		
K-SC	1198,6	1434,4	1670,3		(-*)		
				-1200	0	1200	2400

Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	554,3	790,1	1026,0		(-*)		
				-1200	0	1200	2400

One-way ANOVA: Isopropyl acetate versus Group

Source	DF	SS	MS	F	P
Group	1	2,6	2,6	0,11	0,757
Error	4	94,2	23,6		
Total	5	96,8			

S = 4,853 R-Sq = 2,66% R-Sq(adj) = 0,00%

Individual 95% CIs For Mean Based on Pooled StDev							
Level	N	Mean	StDev				
E-A	3	70,878	2,376	(-----*-----)			
E-Laff	3	69,567	6,439	(-----*-----)			
				65,0	70,0	75,0	80,0

Pooled StDev = 4,853

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-A	3	70,878	A
E-Laff	3	69,567	A

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

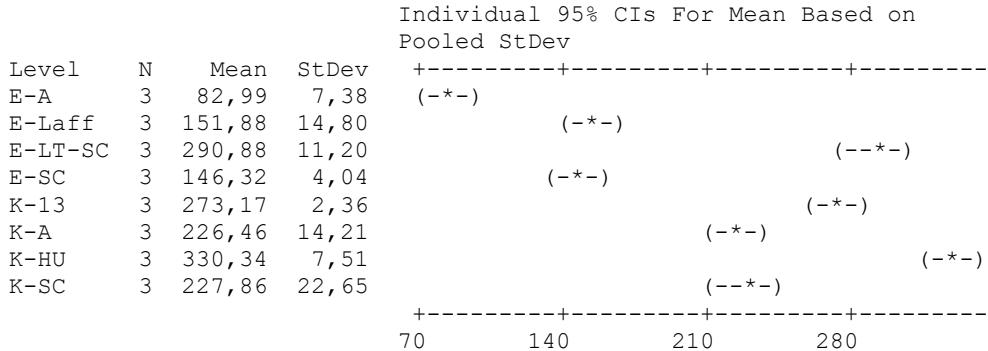
Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-12,314	-1,311	9,691	(-----*-----)
	-12,0	-6,0	0,0	6,0

One-way ANOVA: Phenethyl acetate versus Group

Source	DF	SS	MS	F	P
Group	7	146571	20939	140,46	0,000
Error	16	2385	149		
Total	23	148956			

S = 12,21 R-Sq = 98,40% R-Sq(adj) = 97,70%



Pooled StDev = 12,21

Grouping Information Using Tukey Method

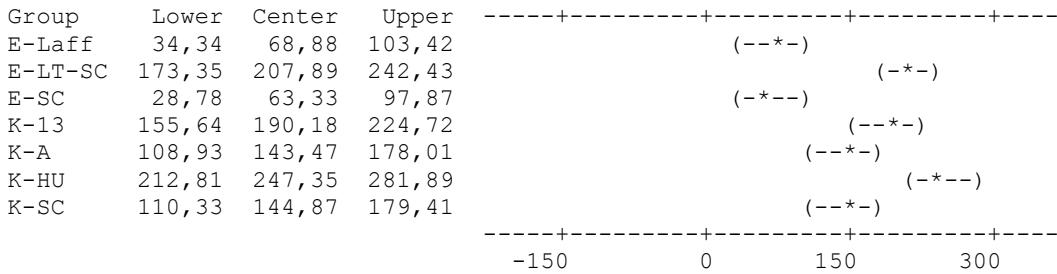
Group	N	Mean	Grouping
K-HU	3	330,34	A
E-LT-SC	3	290,88	B
K-13	3	273,17	B
K-SC	3	227,86	C
K-A	3	226,46	C
E-Laff	3	151,88	D
E-SC	3	146,32	D
E-A	3	82,99	E

Means that do not share a letter are significantly different.

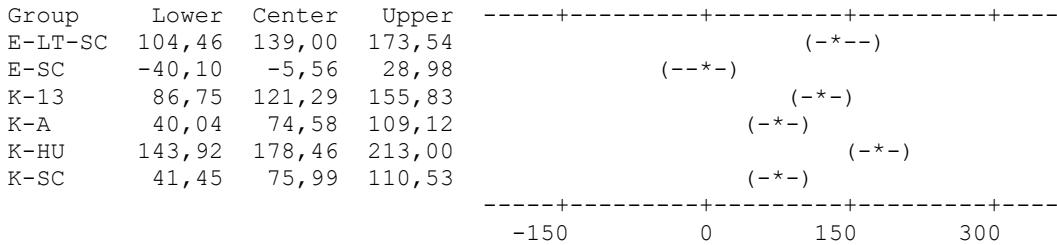
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

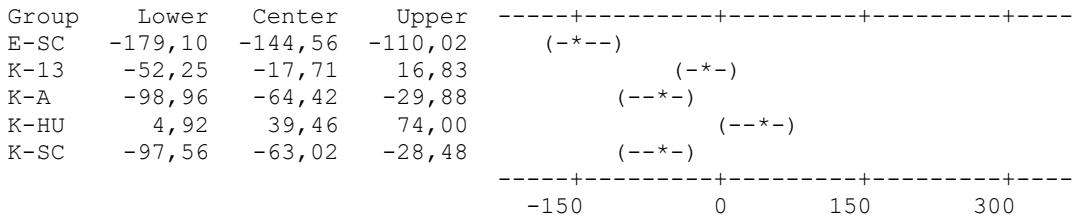
Group = E-A subtracted from:



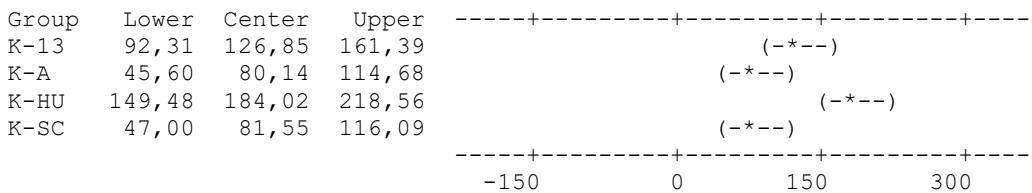
Group = E-Laff subtracted from:



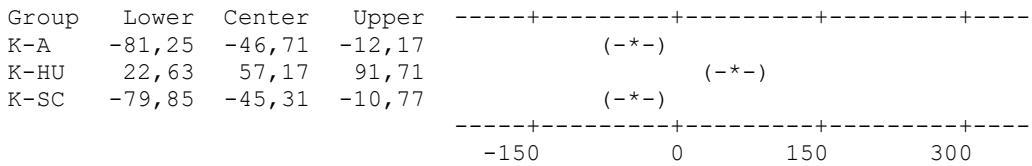
Group = E-LT-SC subtracted from:



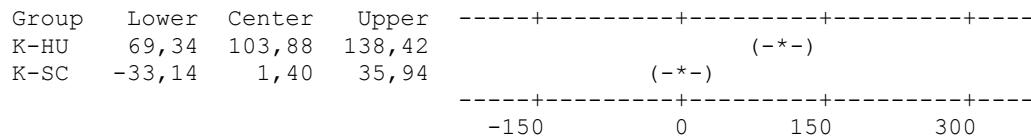
Group = E-SC subtracted from:



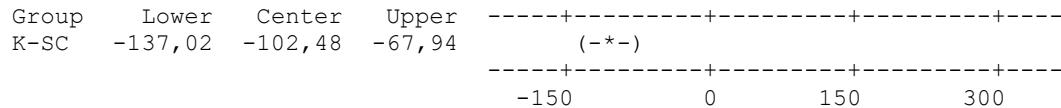
Group = K-13 subtracted from:



Group = K-A subtracted from:



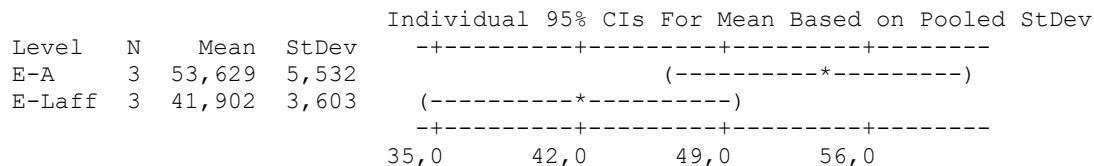
Group = K-HU subtracted from:



One-way ANOVA: Propyl acetate versus Group

Source	DF	SS	MS	F	P
Group	1	206,3	206,3	9,47	0,037
Error	4	87,2	21,8		
Total	5	293,4			

S = 4,668 R-Sq = 70,30% R-Sq(adj) = 62,87%



Pooled StDev = 4,668

Grouping Information Using Tukey Method

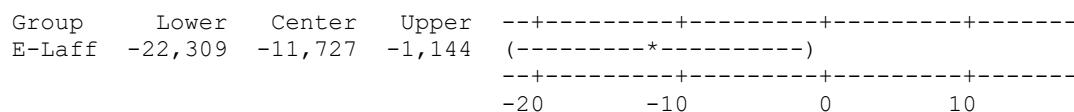
Group	N	Mean	Grouping
E-A	3	53,629	A
E-Laff	3	41,902	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

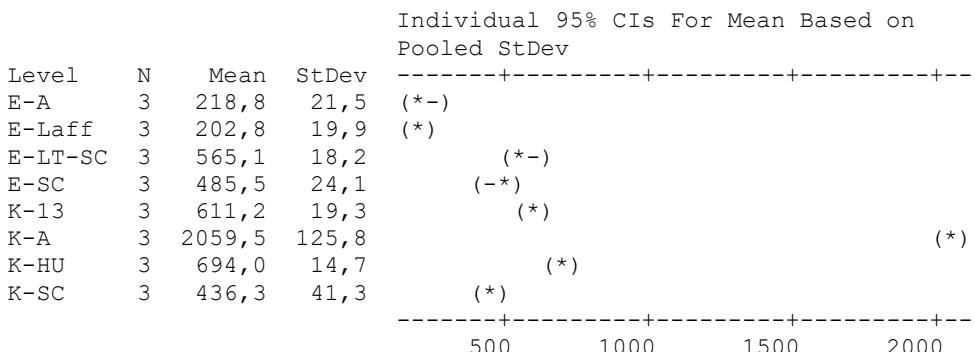
Group = E-A subtracted from:



One-way ANOVA: Ethyl lactate versus Group

Source	DF	SS	MS	F	P
Group	7	7366342	1052335	423,53	0,000
Error	16	39754	2485		
Total	23	7406097			

S = 49,85 R-Sq = 99,46% R-Sq(adj) = 99,23%



Pooled StDev = 49,8

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	2059,5	A
K-HU	3	694,0	B
K-13	3	611,2	B C
E-LT-SC	3	565,1	B C D
E-SC	3	485,5	C D
K-SC	3	436,3	D
E-A	3	218,8	E
E-Laff	3	202,8	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-157,0	-16,0	125,0	(-*)
E-LT-SC	205,4	346,4	487,4	(*-)
E-SC	125,8	266,8	407,8	(-*)

K-13	251,5	392,5	533,5	(*)
K-A	1699,8	1840,8	1981,8	
(*-)				
K-HU	334,2	475,2	616,2	(-*)
K-SC	76,5	217,5	358,5	(*-)
--				
	-2000	-1000	0	1000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper	
--				
E-LT-SC	221,4	362,4	503,4	(-*)
E-SC	141,7	282,8	423,8	(-*)
K-13	267,4	408,5	549,5	(*)
K-A	1715,7	1856,8	1997,8	(-*)
K-HU	350,2	491,2	632,2	(*)
K-SC	92,5	233,5	374,5	(*-)
--				
	-2000	-1000	0	1000

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	
E-SC	-220,6	-79,6	61,4	(*-)
K-13	-94,9	46,1	187,1	(*-)
K-A	1353,4	1494,4	1635,4	(*)
K-HU	-12,2	128,8	269,8	(*-)
K-SC	-269,9	-128,9	12,2	(*-)
	-2000	-1000	0	1000

Group = E-SC subtracted from:

Group	Lower	Center	Upper	
K-13	-15,3	125,7	266,7	(*-)
K-A	1433,0	1574,0	1715,0	(-*)
K-HU	67,4	208,4	349,4	(*)
K-SC	-190,3	-49,2	91,8	(*-)
	-2000	-1000	0	1000

Group = K-13 subtracted from:

Group	Lower	Center	Upper	
K-A	1307,3	1448,3	1589,3	(*-)
K-HU	-58,3	82,7	223,7	(-*)
K-SC	-316,0	-174,9	-33,9	(*-)
	-2000	-1000	0	1000

Group = K-A subtracted from:

Group	Lower	Center	Upper				

K-HU	-1506,6	-1365,6	-1224,6		(*-)		
K-SC	-1764,3	-1623,2	-1482,2		(-*)		

				-2000	-1000	0	1000

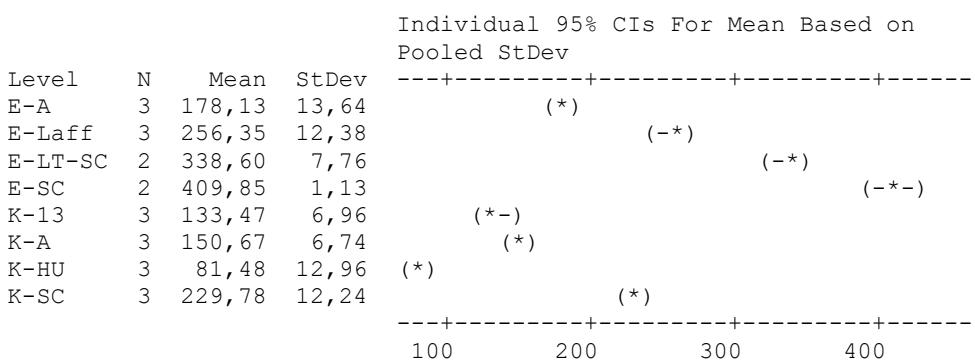
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	-398,7	-257,7	-116,7		(*-)		
				-2000	-1000	0	1000

One-way ANOVA: Ethyl octanoate versus Group

Source	DF	SS	MS	F	P
Group	7	201252	28750	257,45	0,000
Error	14	1563	112		
Total	21	202815			

S = 10,57 R-Sq = 99,23% R-Sq(adj) = 98,84%



Pooled StDev = 10,57

Grouping Information Using Tukey Method

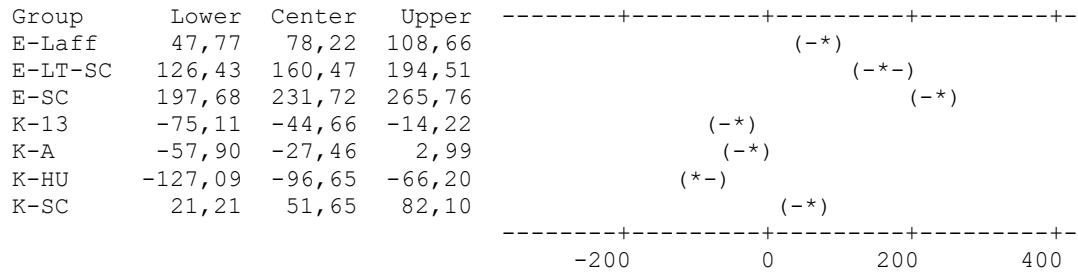
Group	N	Mean	Grouping
E-SC	2	409,85	A
E-LT-SC	2	338,60	B
E-Laff	3	256,35	C
K-SC	3	229,78	C
E-A	3	178,13	D
K-A	3	150,67	D E
K-13	3	133,47	E
K-HU	3	81,48	F

Means that do not share a letter are significantly different.

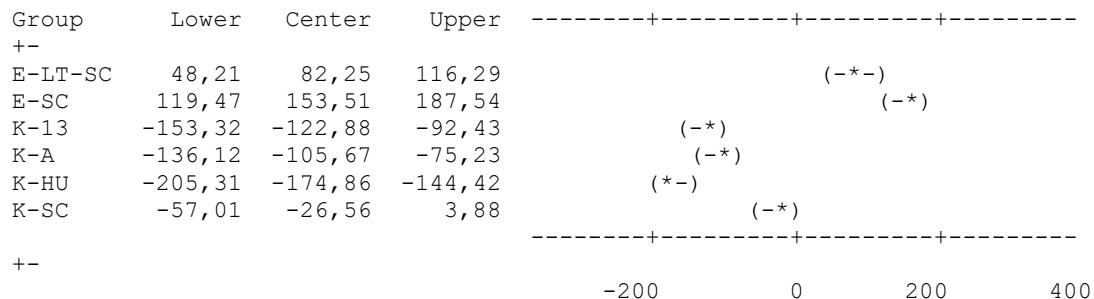
Tukey 95% Simultaneous Confidence Intervals
 All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

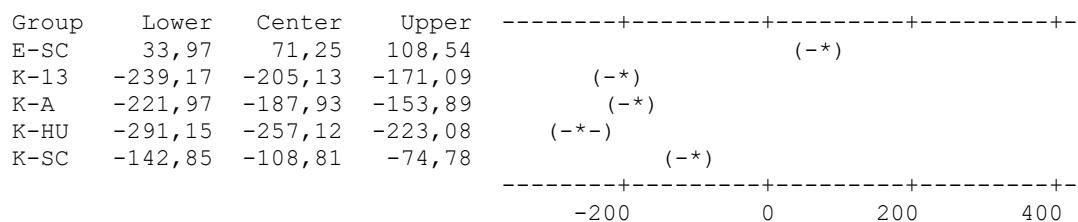
Group = E-A subtracted from:



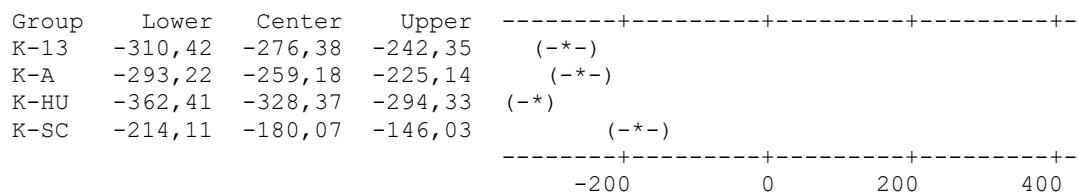
Group = E-Laff subtracted from:



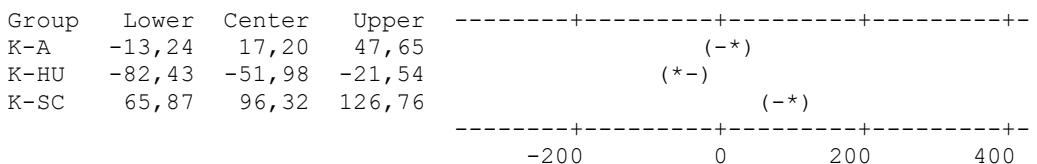
Group = E-LT-SC subtracted from:



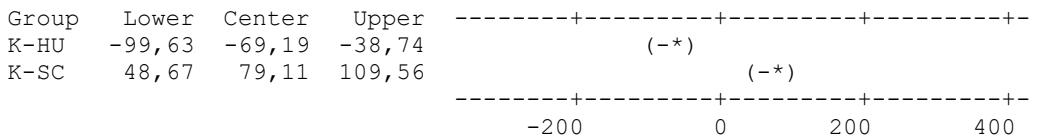
Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



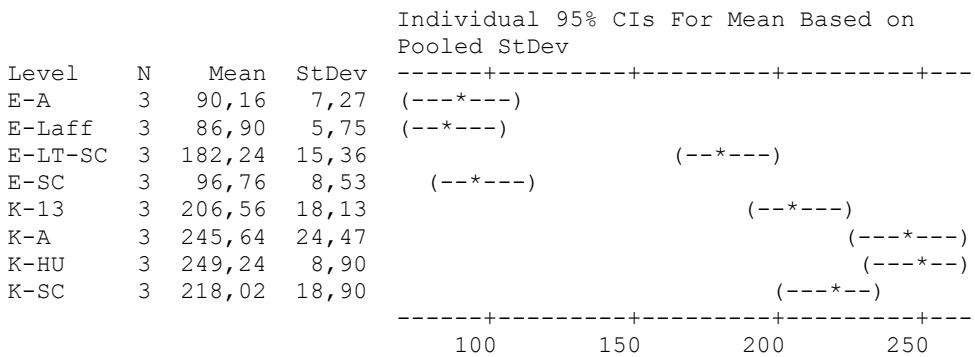
Group = K-HU subtracted from:



One-way ANOVA: Ethyl-3-hydroxybutyrate versus Group

Source	DF	SS	MS	F	P
Group	7	103220	14746	67,09	0,000
Error	16	3517	220		
Total	23	106737			

S = 14,83 R-Sq = 96,71% R-Sq(adj) = 95,26%



Pooled StDev = 14,83

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-HU	3	249,24	A
K-A	3	245,64	A B
K-SC	3	218,02	A B C

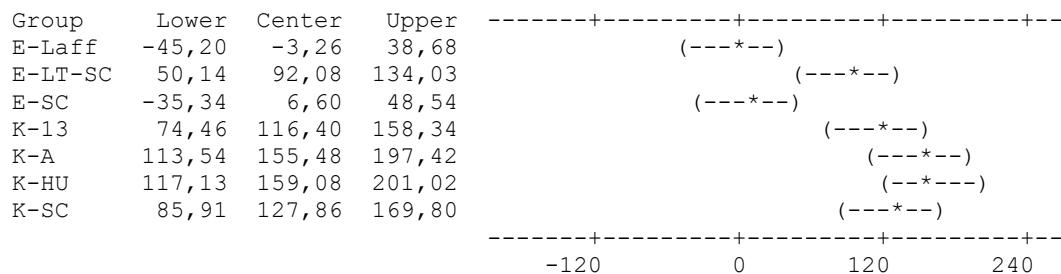
K-13	3	206,56	B C
E-LT-SC	3	182,24	C
E-SC	3	96,76	D
E-A	3	90,16	D
E-Laff	3	86,90	D

Means that do not share a letter are significantly different.

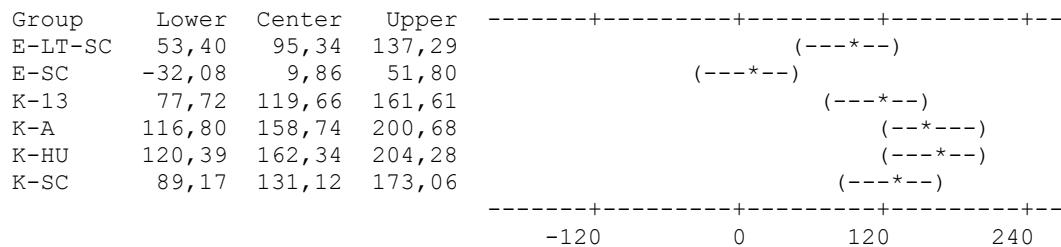
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

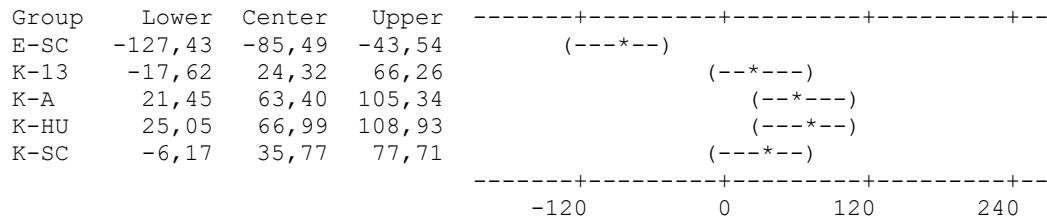
Group = E-A subtracted from:



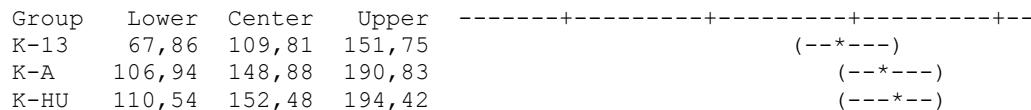
Group = E-Laff subtracted from:

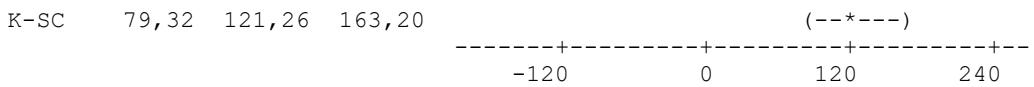


Group = E-LT-SC subtracted from:

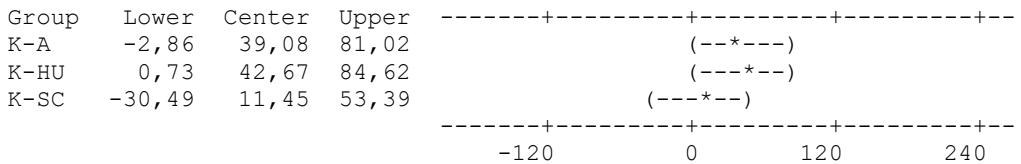


Group = E-SC subtracted from:

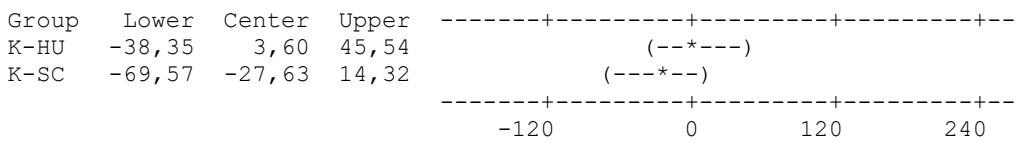




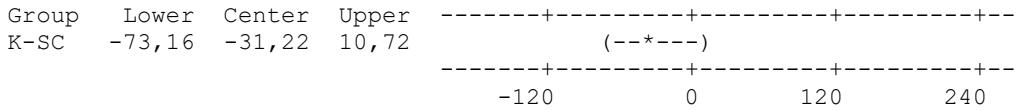
Group = K-13 subtracted from:



Group = K-A subtracted from:



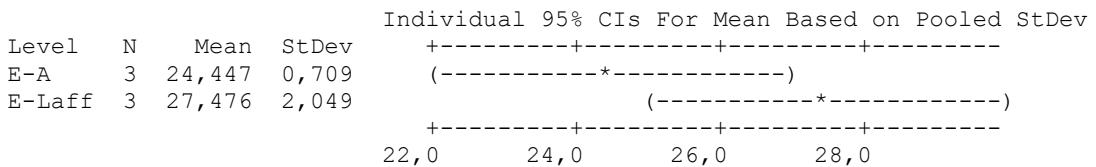
Group = K-HU subtracted from:



One-way ANOVA: Ethyl 9-decenoate versus Group

Source	DF	SS	MS	F	P
Group	1	13,76	13,76	5,86	0,073
Error	4	9,40	2,35		
Total	5	23,17			

S = 1,533 R-Sq = 59,41% R-Sq(adj) = 49,27%



Pooled StDev = 1,533

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-Laff	3	27,476	A
E-A	3	24,447	A

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
 All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

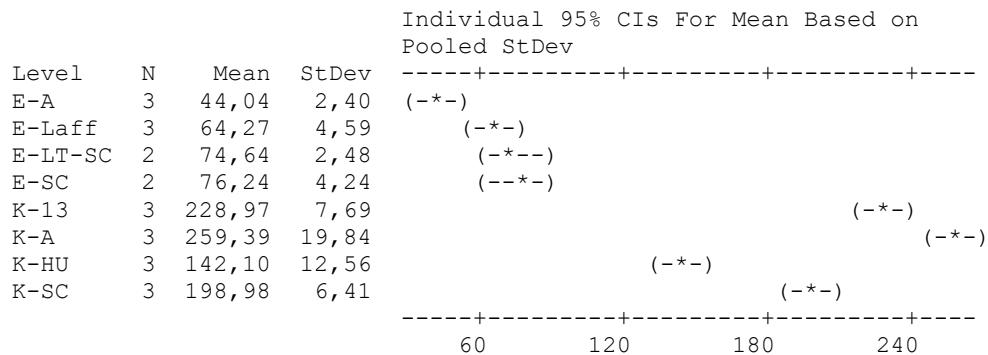
Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-0,446	3,029	6,505	(-----*-----)
	-2,5	0,0	2,5	5,0

One-way ANOVA: Diethyl succinate versus Group

Source	DF	SS	MS	F	P
Group	7	138402,6	19771,8	200,46	0,000
Error	14	1380,8	98,6		
Total	21	139783,4			

S = 9,931 R-Sq = 99,01% R-Sq(adj) = 98,52%



Pooled StDev = 9,93

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	259,39	A
K-13	3	228,97	B
K-SC	3	198,98	C
K-HU	3	142,10	D
E-SC	2	76,24	E
E-LT-SC	2	74,64	E F
E-Laff	3	64,27	E F
E-A	3	44,04	F

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals

All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	-8,38	20,23	48,84		(--*--)		
E-LT-SC	-1,39	30,60	62,59		(--*--)		
E-SC	0,20	32,19	64,18		(--*--)		
K-13	156,32	184,93	213,54		(-*--)		
K-A	186,73	215,34	243,95		(-*--)		
K-HU	69,44	98,05	126,66		(-*--)		
K-SC	126,32	154,94	183,55		(-*--)		
				-120	0	120	240

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	-21,62	10,37	42,35		(--*--)		
E-SC	-20,03	11,96	43,95		(--*--)		
K-13	136,09	164,70	193,31		(--*--)		
K-A	166,50	195,11	223,72		(-*--)		
K-HU	49,21	77,82	106,43		(-*--)		
K-SC	106,09	134,70	163,32		(-*--)		
				-120	0	120	240

Group = E-LT-SC subtracted from:

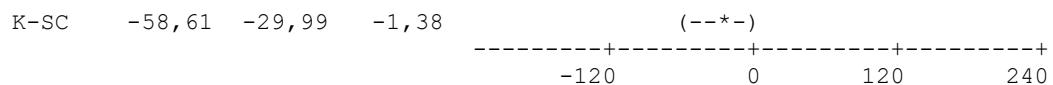
Group	Lower	Center	Upper				
E-SC	-33,45	1,60	36,64		(--*--)		
K-13	122,34	154,33	186,32		(--*--)		
K-A	152,76	184,75	216,74		(-*--)		
K-HU	35,47	67,46	99,45		(--*--)		
K-SC	92,35	124,34	156,33		(-*--)		
				-120	0	120	240

Group = E-SC subtracted from:

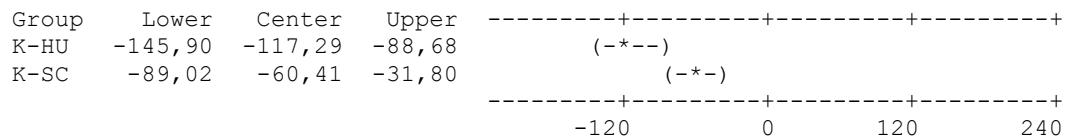
Group	Lower	Center	Upper				
K-13	120,75	152,74	184,73		(--*--)		
K-A	151,16	183,15	215,14		(-*--)		
K-HU	33,87	65,86	97,85		(-*--)		
K-SC	90,75	122,74	154,73		(-*--)		
				-120	0	120	240

Group = K-13 subtracted from:

Group	Lower	Center	Upper				
K-A	1,80	30,41	59,02		(--*--)		
K-HU	-115,49	-86,88	-58,27		(--*--)		



Group = K-A subtracted from:



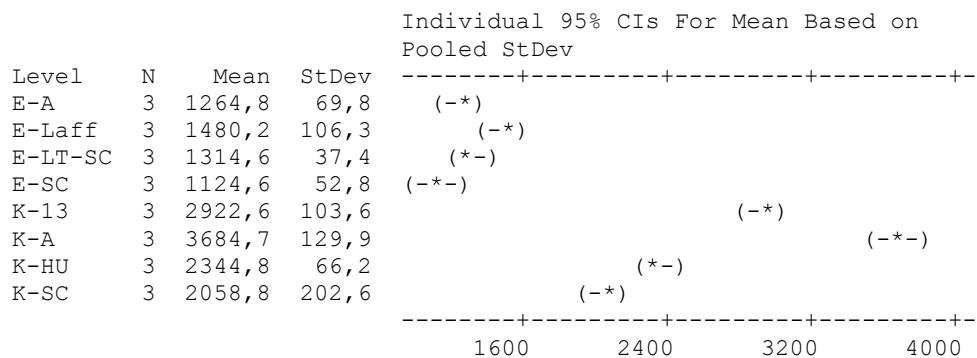
Group = K-HU subtracted from:



One-way ANOVA: Ethyl 4-hydroxybutanoate versus Group

Source	DF	SS	MS	F	P
Group	7	17561306	2508758	214,96	0,000
Error	16	186730	11671		
Total	23	17748036			

S = 108, 0 R-Sq = 98, 95% R-Sq(adj) = 98, 49%



Pooled StDev = 108, 0

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	3684,7	A
K-13	3	2922,6	B
K-HU	3	2344,8	C
K-SC	3	2058,8	C
E-Laff	3	1480,2	D
E-LT-SC	3	1314,6	D E
E-A	3	1264,8	D E

E-SC 3 1124,6 E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	-90,2	215,4	521,0		(-*)		
E-LT-SC	-255,8	49,8	355,4		(-*)		
E-SC	-445,9	-140,2	165,4		(-*)		
K-13	1352,2	1657,8	1963,4		(-*)		
K-A	2114,2	2419,9	2725,5		(-*)		
K-HU	774,4	1080,0	1385,6		(-*)		
K-SC	488,3	793,9	1099,6		(-*)		
				-1500	0	1500	3000

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	-471,2	-165,6	140,0		(-*)		
E-SC	-661,3	-355,6	-50,0		(-*)		
K-13	1136,8	1442,4	1748,0		(-*)		
K-A	1898,8	2204,5	2510,1		(-*)		
K-HU	559,0	864,6	1170,2		(-*)		
K-SC	272,9	578,5	884,2		(-*)		
				-1500	0	1500	3000

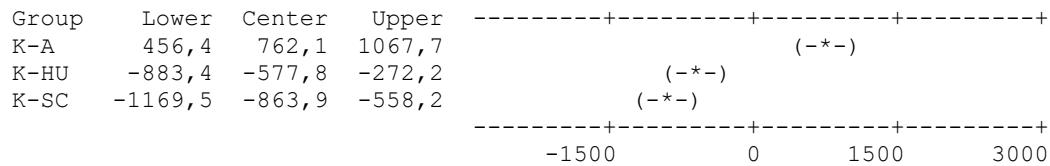
Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-495,7	-190,0	115,6		(-*)		
K-13	1302,4	1608,0	1913,6		(-*)		
K-A	2064,4	2370,1	2675,7		(-*)		
K-HU	724,6	1030,2	1335,8		(-*)		
K-SC	438,5	744,1	1049,8		(-*)		
				-1500	0	1500	3000

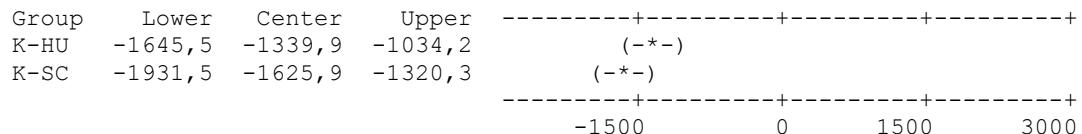
Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	1492,4	1798,0	2103,7		(-*)		
K-A	2254,5	2560,1	2865,7		(-*)		
K-HU	914,6	1220,2	1525,9		(-*)		
K-SC	628,6	934,2	1239,8		(-*)		
				-1500	0	1500	3000

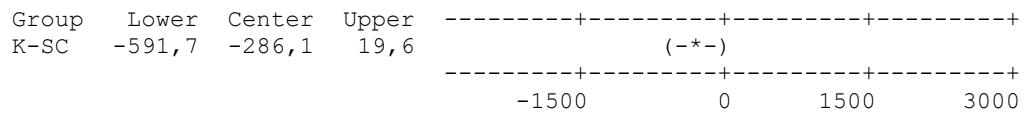
Group = K-13 subtracted from:



Group = K-A subtracted from:



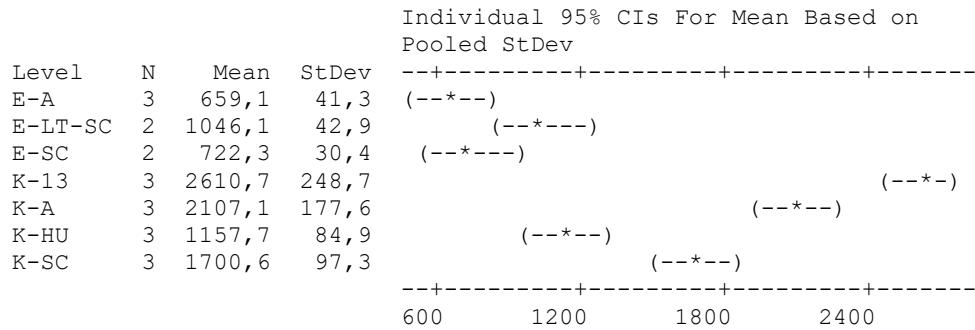
Group = K-HU subtracted from:



One-way ANOVA: Monoethyl succinate versus Group

Source	DF	SS	MS	F	P
Group	6	9018316	1503053	79,68	0,000
Error	12	226370	18864		
Total	18	9244686			

S = 137,3 R-Sq = 97,55% R-Sq(adj) = 96,33%



Pooled StDev = 137,3

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-13	3	2610,7	A
K-A	3	2107,1	B

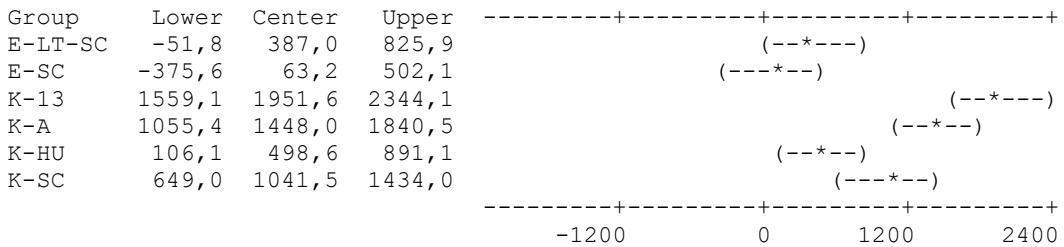
K-SC	3	1700,6	C
K-HU	3	1157,7	D
E-LT-SC	2	1046,1	D E
E-SC	2	722,3	D E
E-A	3	659,1	E

Means that do not share a letter are significantly different.

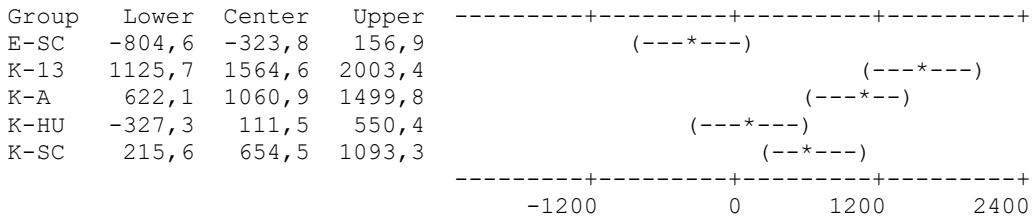
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,56%

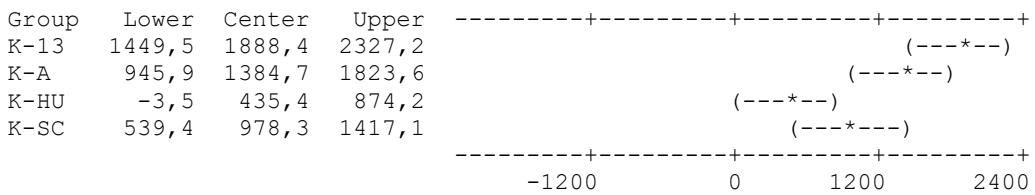
Group = E-A subtracted from:



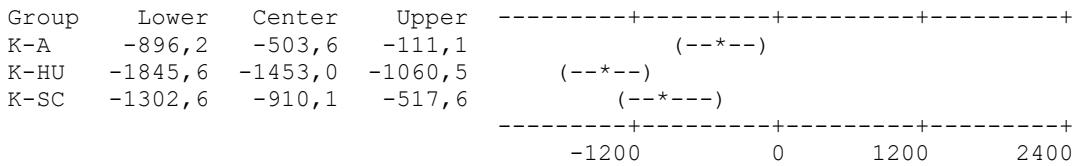
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:

Group	Lower	Center	Upper				
K-HU	-1341,9	-949,4	-556,9		(--*--)		
K-SC	-799,0	-406,5	-13,9		(---*--)		
				-1200	0	1200	2400

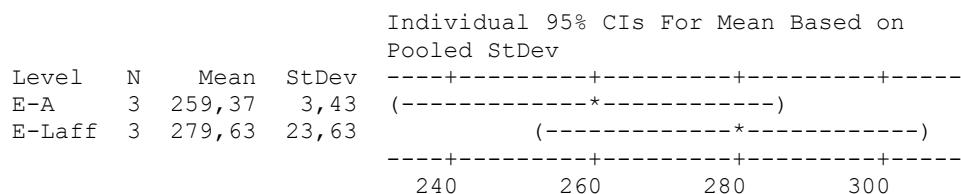
Group = K-HU subtracted from:

Group	Lower	Center	Upper				
K-SC	150,4	542,9	935,4		(---*--)		
				-1200	0	1200	2400

One-way ANOVA: Ethyl butyrate versus Group

Source	DF	SS	MS	F	P
Group	1	616	616	2,16	0,216
Error	4	1140	285		
Total	5	1756			

S = 16,88 R-Sq = 35,06% R-Sq(adj) = 18,82%



Pooled StDev = 16,88

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-Laff	3	279,63	A
E-A	3	259,37	A

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

Group = E-A subtracted from:

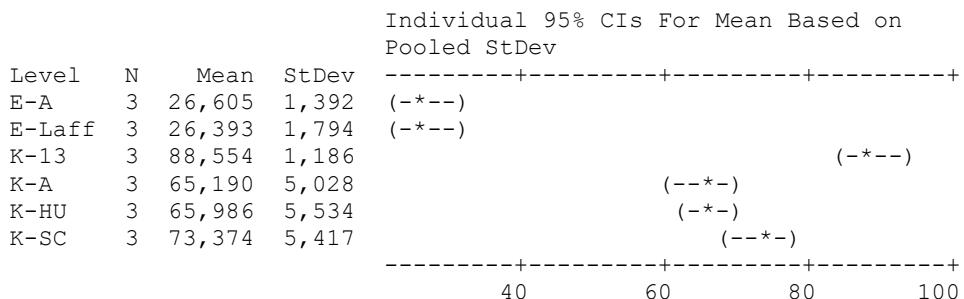
Group	Lower	Center	Upper				
E-Laff	-18,02	20,26	58,54		(-----*-----)		

-25 0 25 50

One-way ANOVA: Ethyl-3-hydroxypropionate versus Group

Source	DF	SS	MS	F	P
Group	5	9808,3	1961,7	128,21	0,000
Error	12	183,6	15,3		
Total	17	9992,0			

S = 3,912 R-Sq = 98,16% R-Sq(adj) = 97,40%



Pooled StDev = 3,912

Grouping Information Using Tukey Method

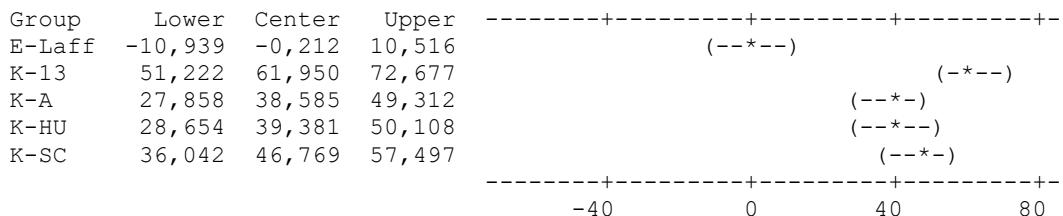
Group	N	Mean	Grouping
K-13	3	88,554	A
K-SC	3	73,374	B
K-HU	3	65,986	B
K-A	3	65,190	B
E-A	3	26,605	C
E-Laff	3	26,393	C

Means that do not share a letter are significantly different.

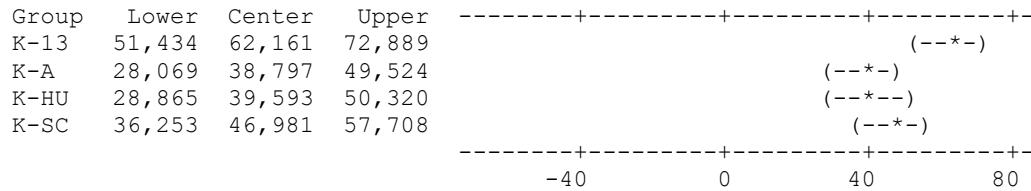
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

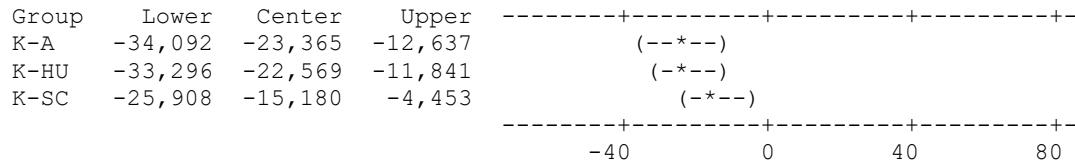
Group = E-A subtracted from:



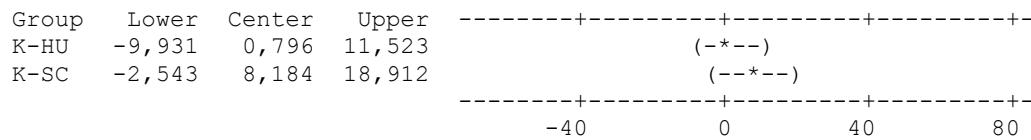
Group = E-Laff subtracted from:



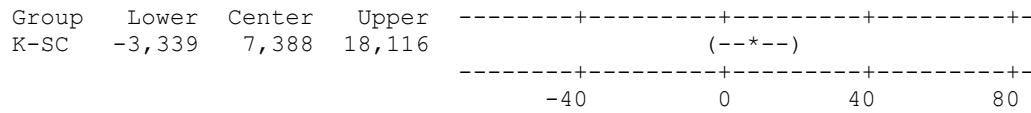
Group = K-13 subtracted from:



Group = K-A subtracted from:



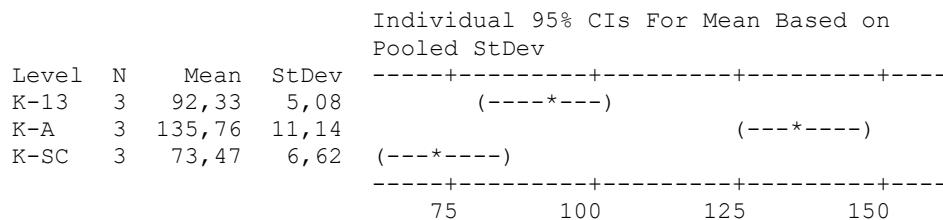
Group = K-HU subtracted from:



One-way ANOVA: Ethyl2-hydroxy3phenylpropanoate versus Group

Source	DF	SS	MS	F	P
Group	2	6121,1	3060,6	47,37	0,000
Error	6	387,7	64,6		
Total	8	6508,8			

S = 8,038 R-Sq = 94,04% R-Sq(adj) = 92,06%



Pooled StDev = 8,04

Grouping Information Using Tukey Method

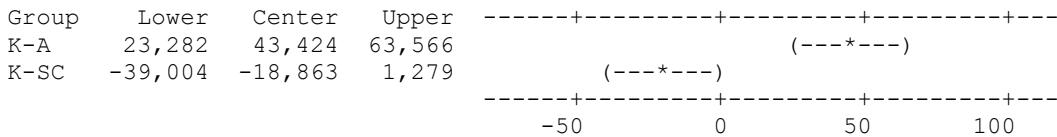
Group	N	Mean	Grouping
K-A	3	135,755	A
K-13	3	92,331	B
K-SC	3	73,468	B

Means that do not share a letter are significantly different.

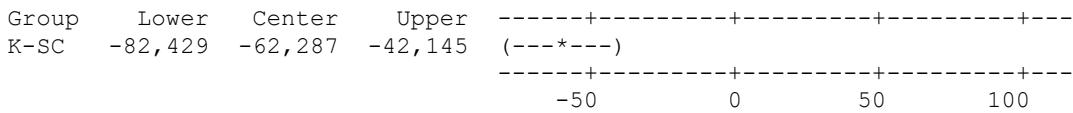
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 97,80%

Group = K-13 subtracted from:



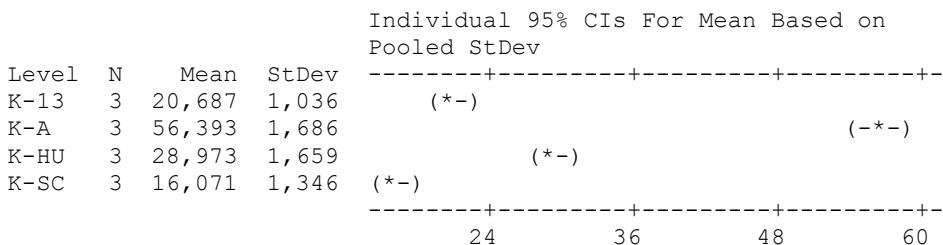
Group = K-A subtracted from:



One-way ANOVA: Methyl 4-hydroxybutanoate versus Group

Source	DF	SS	MS	F	P
Group	3	2931,83	977,28	460,90	0,000
Error	8	16,96	2,12		
Total	11	2948,79			

S = 1,456 R-Sq = 99,42% R-Sq(adj) = 99,21%



Pooled StDev = 1,456

Grouping Information Using Tukey Method

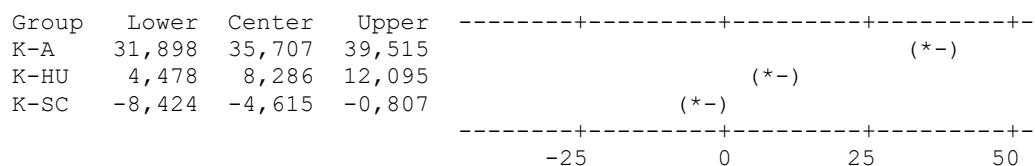
Group	N	Mean	Grouping
K-A	3	56,393	A
K-HU	3	28,973	B
K-13	3	20,687	C
K-SC	3	16,071	D

Means that do not share a letter are significantly different.

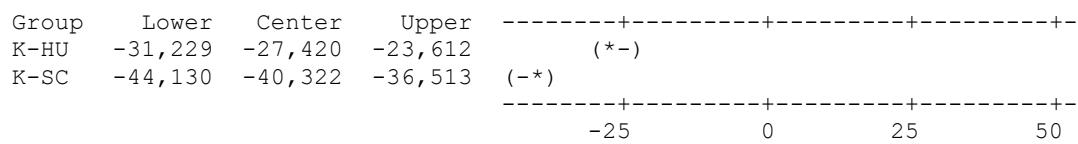
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

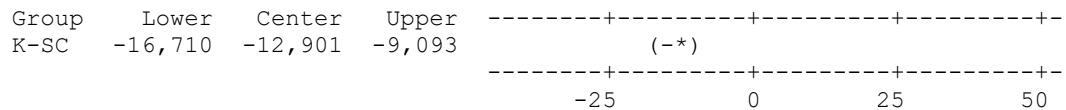
Group = K-13 subtracted from:



Group = K-A subtracted from:



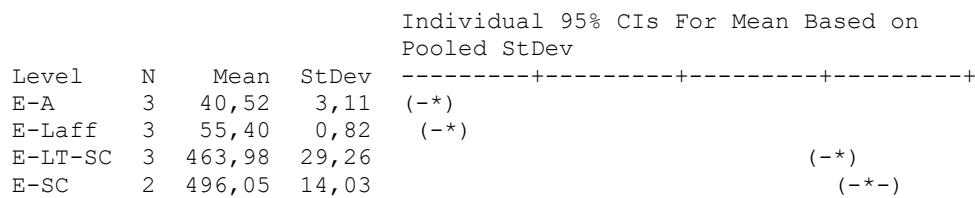
Group = K-HU subtracted from:



One-way ANOVA: Ethyl decanoate versus Group

Source	DF	SS	MS	F	P
Group	3	503145	167715	608,39	0,000
Error	7	1930	276		
Total	10	505075			

S = 16,60 R-Sq = 99,62% R-Sq(adj) = 99,45%



-----+-----+-----+-----+
 150 300 450 600

Pooled StDev = 16,60

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-SC	2	496,05	A
E-LT-SC	3	463,98	A
E-Laff	3	55,40	B
E-A	3	40,52	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,70%

Group = E-A subtracted from:

Group	Lower	Center	Upper			
-						
E-Laff	-29,99	14,87	59,74		(-*)	
E-LT-SC	378,59	423,46	468,32		(-*)	
E-SC	405,38	455,53	505,69		(*-)	
-						
				-500	-250	0
						250

Group = E-Laff subtracted from:

Group	Lower	Center	Upper			
-						
E-LT-SC	363,72	408,58	453,44		(*-)	
E-SC	390,50	440,66	490,81		(*-)	
-						
				-500	-250	0
						250

Group = E-LT-SC subtracted from:

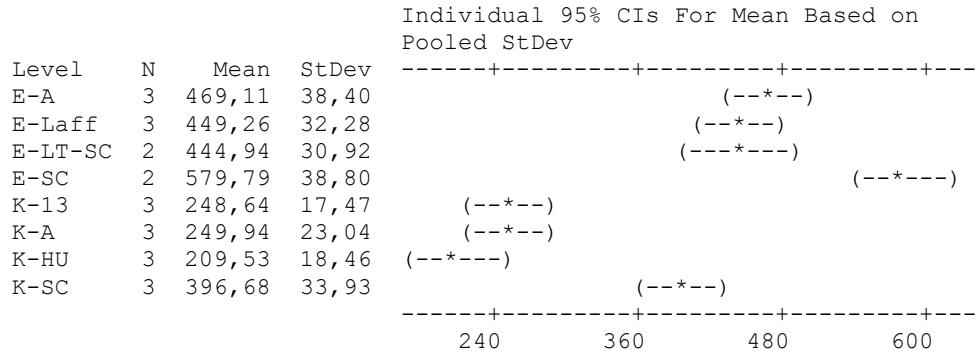
Group	Lower	Center	Upper			
E-SC	-18,08	32,08	82,23		(-*)	
				-500	-250	0
						250

One-way ANOVA: Ethyl hexanoate versus Group

Source	DF	SS	MS	F	P
Group	7	314372	44910	51,75	0,000
Error	14	12150	868		

Total 21 326522

S = 29,46 R-Sq = 96,28% R-Sq(adj) = 94,42%



Grouping Information Using Tukey Method

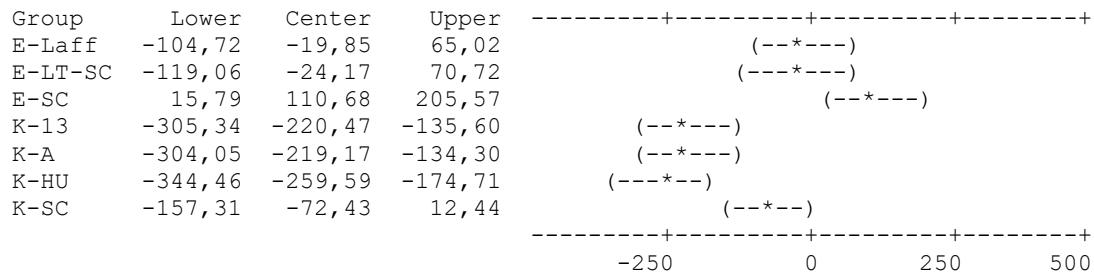
Group	N	Mean	Grouping
E-SC	2	579,79	A
E-A	3	469,11	B
E-Laff	3	449,26	B
E-LT-SC	2	444,94	B
K-SC	3	396,68	B
K-A	3	249,94	C
K-13	3	248,64	C
K-HU	3	209,53	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

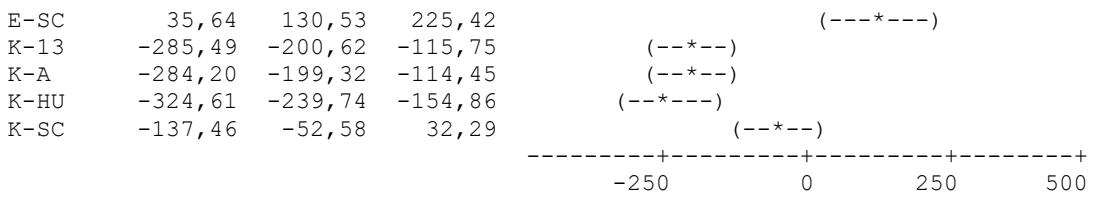
Individual confidence level = 99,67%

Group = E-A subtracted from:

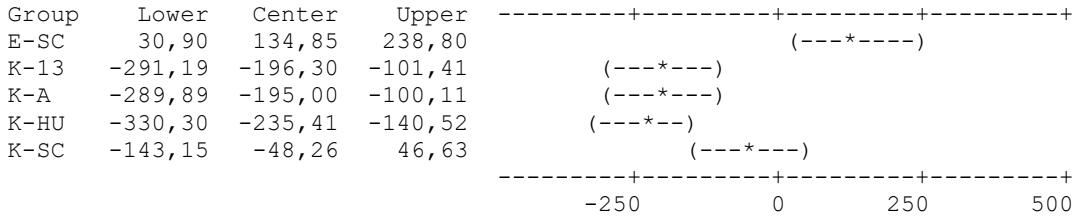


Group = E-Laff subtracted from:

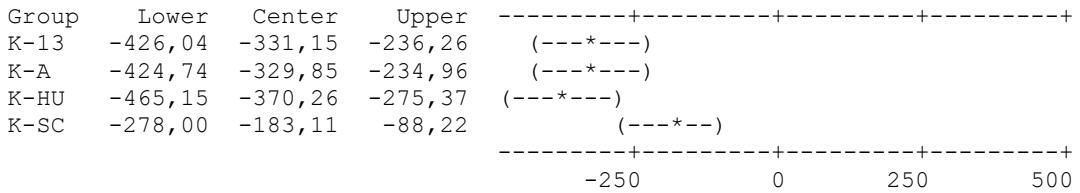
Group	Lower	Center	Upper	95% CI
E-LT-SC	-99,21	-4,32	90,57	(---*--)



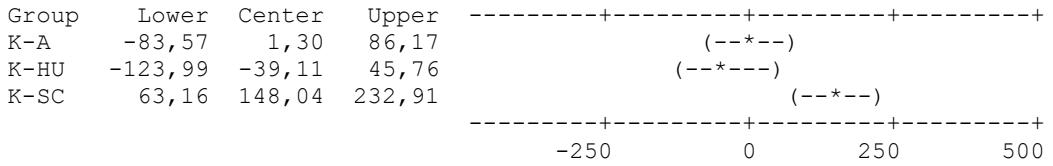
Group = E-LT-SC subtracted from:



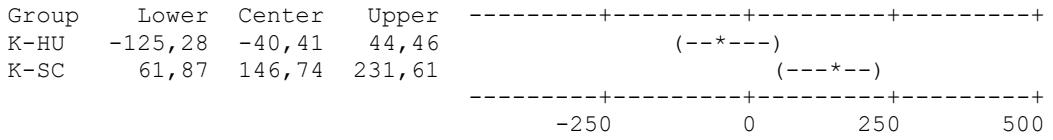
Group = E-SC subtracted from:



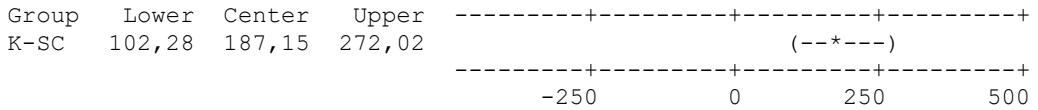
Group = K-13 subtracted from:



Group = K-A subtracted from:



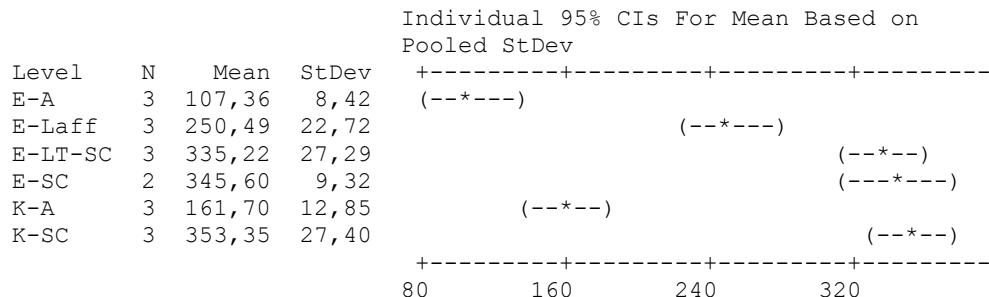
Group = K-HU subtracted from:



One-way ANOVA: Butanoic acid versus Group

Source	DF	SS	MS	F	P
Group	5	156287	31257	75,02	0,000
Error	11	4583	417		
Total	16	160870			

S = 20,41 R-Sq = 97,15% R-Sq(adj) = 95,86%



Pooled StDev = 20,41

Grouping Information Using Tukey Method

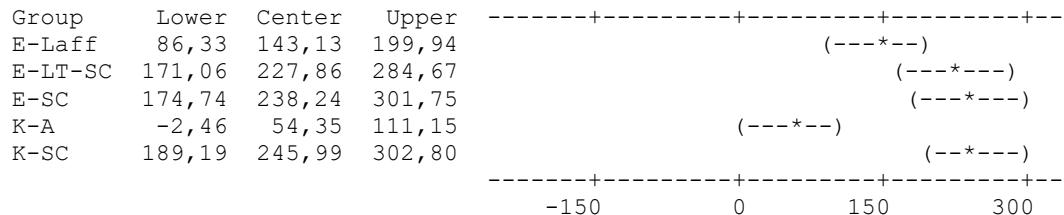
Group	N	Mean	Grouping
K-SC	3	353,35	A
E-SC	2	345,60	A
E-LT-SC	3	335,22	A
E-Laff	3	250,49	B
K-A	3	161,70	C
E-A	3	107,36	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

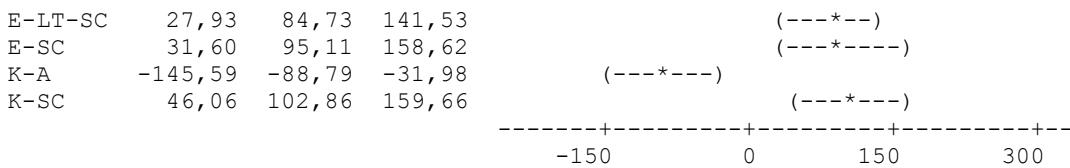
Individual confidence level = 99,42%

Group = E-A subtracted from:

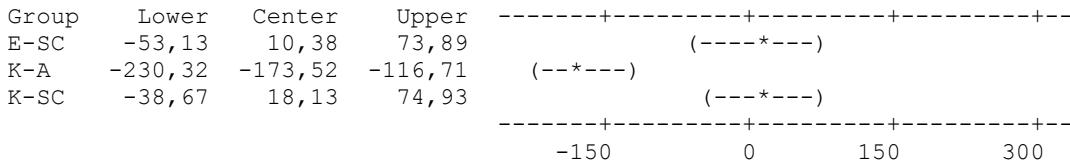


Group = E-Laff subtracted from:

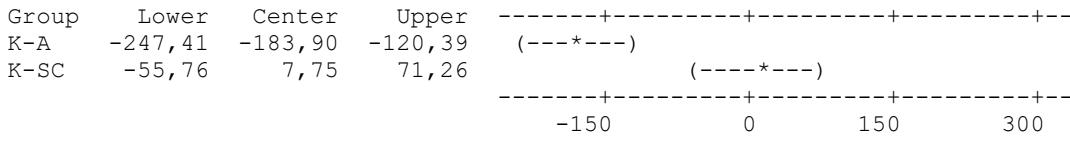
Group Lower Center Upper CI Lower CI Upper



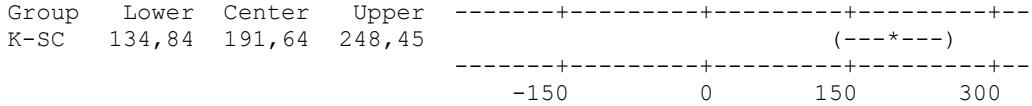
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



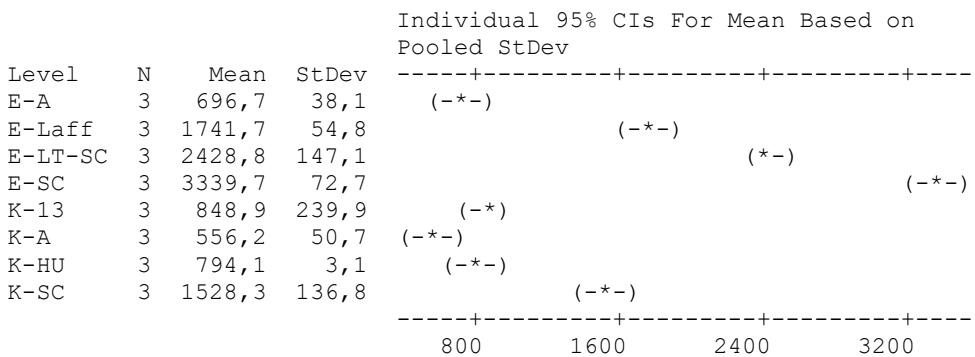
Group = K-A subtracted from:



One-way ANOVA: Hexanoic acid versus Group

Source	DF	SS	MS	F	P
Group	7	20292779	2898968	210, 36	0, 000
Error	16	220495	13781		
Total	23	20513274			

S = 117, 4 R-Sq = 98, 93% R-Sq(adj) = 98, 45%



Pooled StDev = 117,4

Grouping Information Using Tukey Method

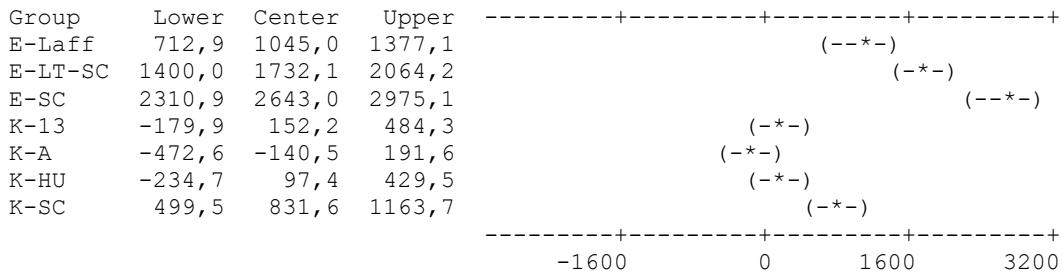
Group	N	Mean	Grouping
E-SC	3	3339,7	A
E-LT-SC	3	2428,8	B
E-Laff	3	1741,7	C
K-SC	3	1528,3	C
K-13	3	848,9	D
K-HU	3	794,1	D
E-A	3	696,7	D
K-A	3	556,2	D

Means that do not share a letter are significantly different.

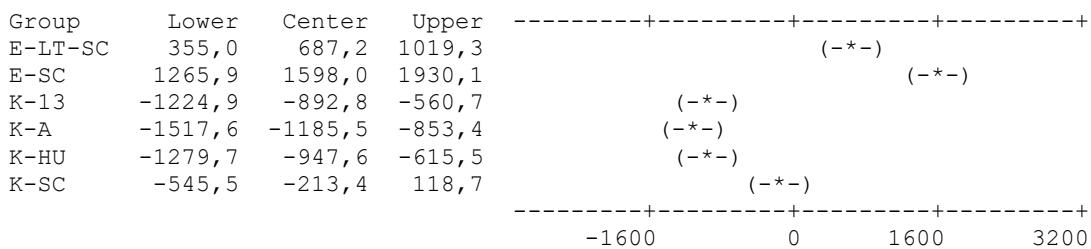
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

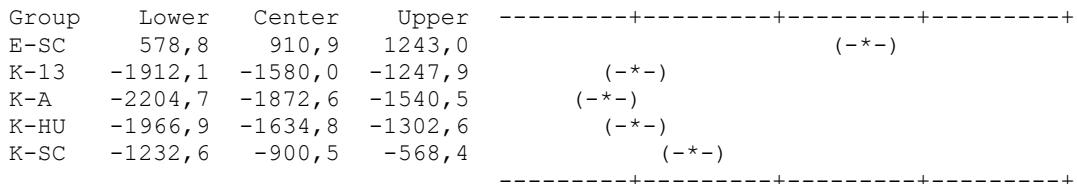
Group = E-A subtracted from:



Group = E-Laff subtracted from:

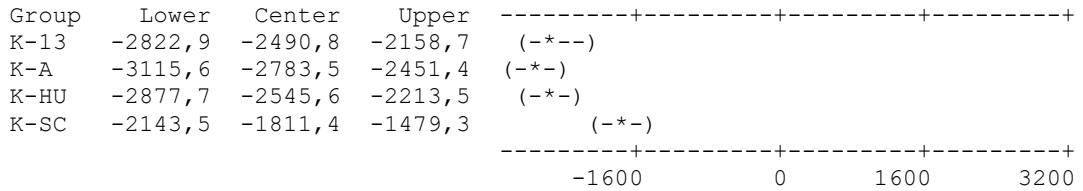


Group = E-LT-SC subtracted from:

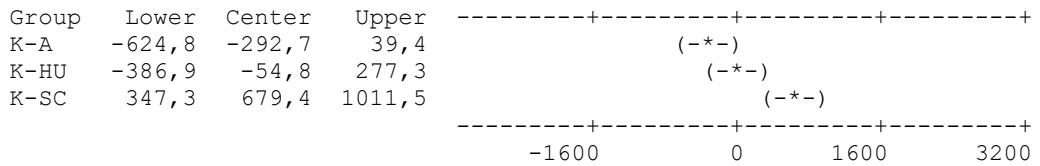


-1600 0 1600 3200

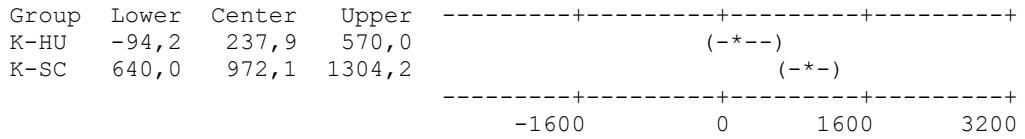
Group = E-SC subtracted from:



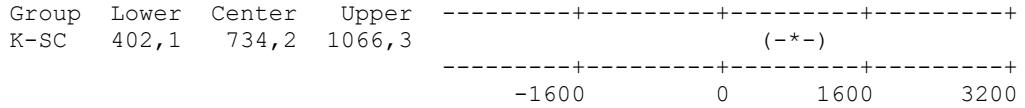
Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:



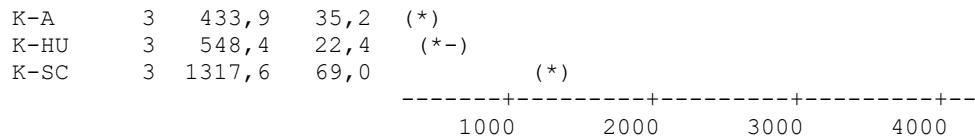
One-way ANOVA: Octanoic acid versus Group

Source	DF	SS	MS	F	P
Group	7	33177837	4739691	623,81	0,000
Error	15	113970	7598		
Total	22	33291806			

S = 87,17 R-Sq = 99,66% R-Sq(adj) = 99,50%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev	
E-A	3	1057,2	50,6	(-*)
E-Laff	3	2251,8	58,5	(-*)
E-LT-SC	3	3300,3	195,0	(*)
E-SC	2	3973,7	113,6	(-*)
K-13	3	527,0	4,3	(*)



Pooled StDev = 87,2

Grouping Information Using Tukey Method

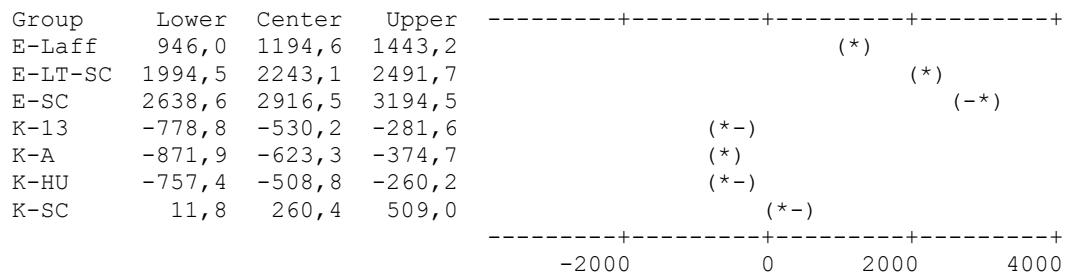
Group	N	Mean	Grouping
E-SC	2	3973,7	A
E-LT-SC	3	3300,3	B
E-Laff	3	2251,8	C
K-SC	3	1317,6	D
E-A	3	1057,2	E
K-HU	3	548,4	F
K-13	3	527,0	F
K-A	3	433,9	F

Means that do not share a letter are significantly different.

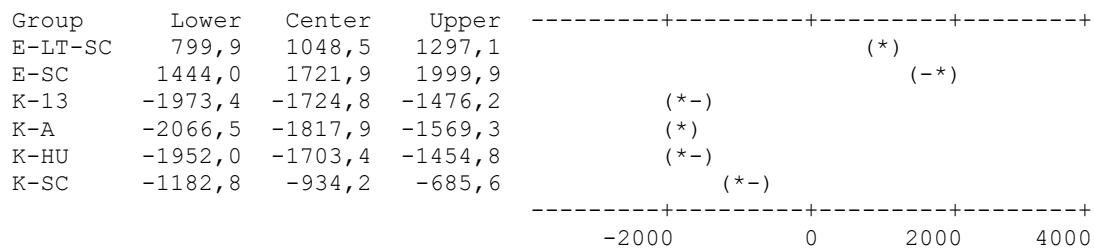
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

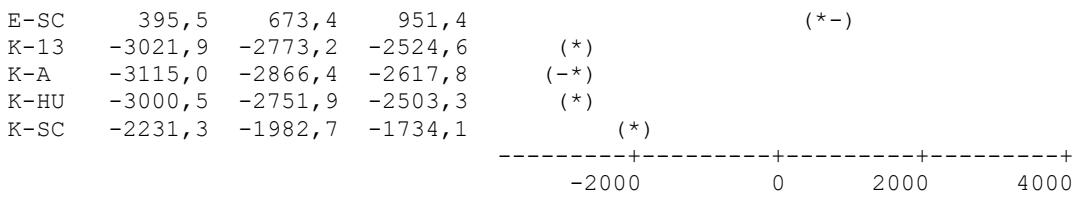


Group = E-Laff subtracted from:

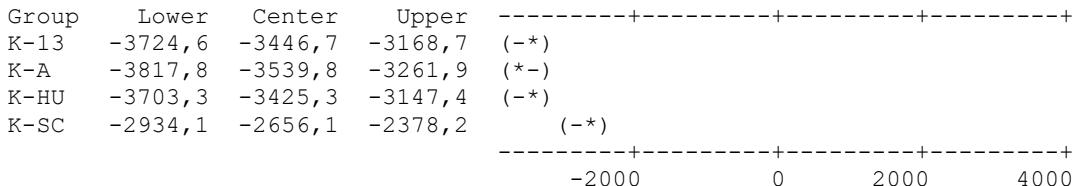


Group = E-LT-SC subtracted from:

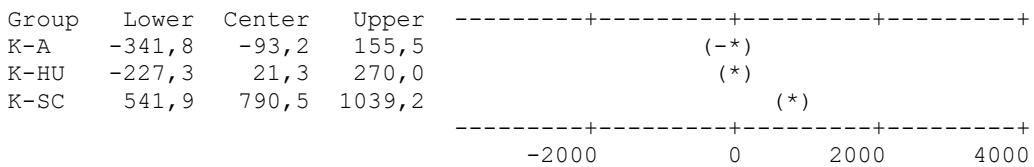
Group Lower Center Upper -----+-----+-----+-----+



Group = E-SC subtracted from:



Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:



One-way ANOVA: Decanoic acid versus Group

Source	DF	SS	MS	F	P
Group	4	9377, 3	2344, 3	33, 17	0, 000
Error	8	565, 5	70, 7		
Total	12	9942, 8			

S = 8,407 R-Sq = 94,31% R-Sq(adj) = 91,47%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev				
E-A	3	127,71	10,38		(---*---)	
E-Laff	3	138,02	9,35		(---*---)	
E-LT-SC	2	132,54	8,44		(---*---)	
E-SC	2	78,99	6,42	(---	*	---)
K-SC	3	77,44	5,60	(---	*	---)
				-----+-----+-----+-----			
				75	100	125	150

Pooled StDev = 8,41

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-Laff	3	138,02	A
E-LT-SC	2	132,54	A
E-A	3	127,71	A
E-SC	2	78,99	B
K-SC	3	77,44	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,14%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	-13,43	10,31	34,05		(---*---)	
E-LT-SC	-21,71	4,83	31,36		(---*---)	
E-SC	-75,26	-48,73	-22,19	(---	*	---)
K-SC	-74,00	-50,27	-26,53	(---	*	---)
				-----+-----+-----+-----			
				-50	0	50	100

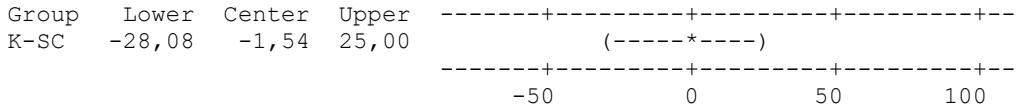
Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	-32,02	-5,48	21,05		(---*---)	
E-SC	-85,58	-59,04	-32,50	(---	*	---)
K-SC	-84,32	-60,58	-36,84	(---	*	---)
				-----+-----+-----+-----			
				-50	0	50	100

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-82,62	-53,55	-24,48	(---	*	---)
K-SC	-81,63	-55,10	-28,56	(---	*	---)
				-----+-----+-----+-----			
				-50	0	50	100

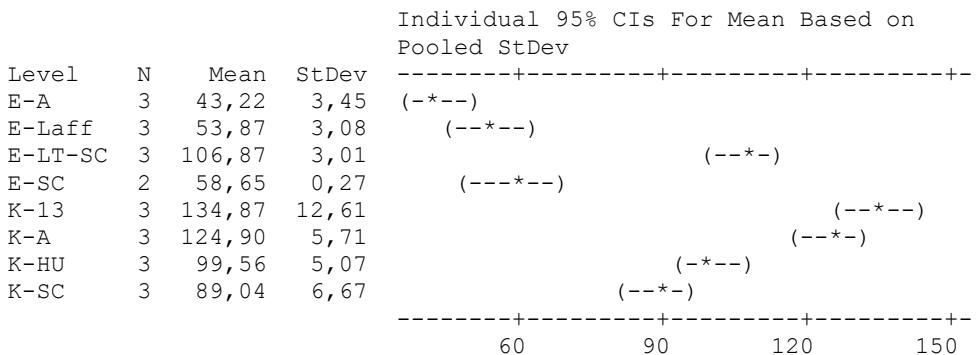
Group = E-SC subtracted from:



One-way ANOVA: Propanoic acid versus Group

Source	DF	SS	MS	F	P
Group	7	23271,0	3324,4	85,33	0,000
Error	15	584,4	39,0		
Total	22	23855,4			

S = 6,242 R-Sq = 97,55% R-Sq(adj) = 96,41%



Pooled StDev = 6,24

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-13	3	134,87	A
K-A	3	124,90	A
E-LT-SC	3	106,87	B
K-HU	3	99,56	B C
K-SC	3	89,04	C
E-SC	2	58,65	D
E-Laff	3	53,87	D
E-A	3	43,22	D

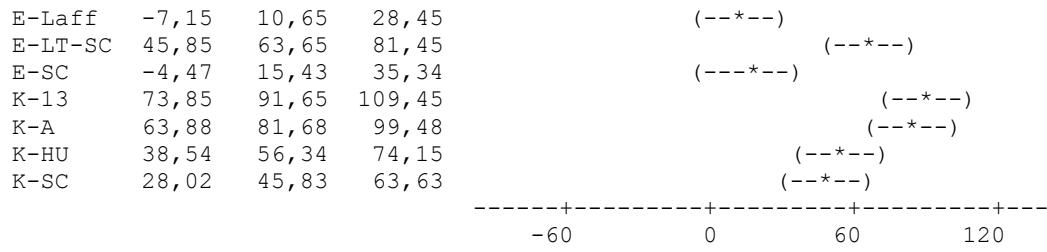
Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

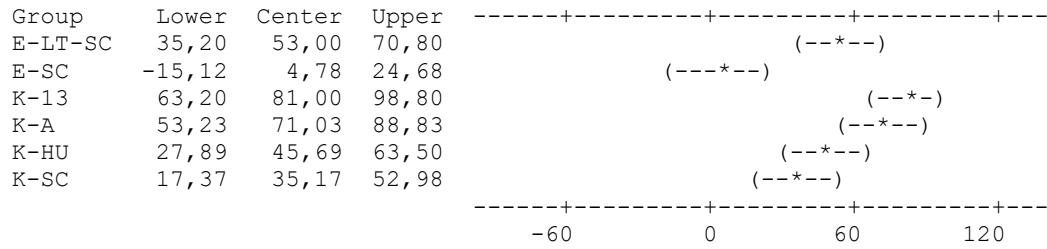
Individual confidence level = 99,67%

Group = E-A subtracted from:

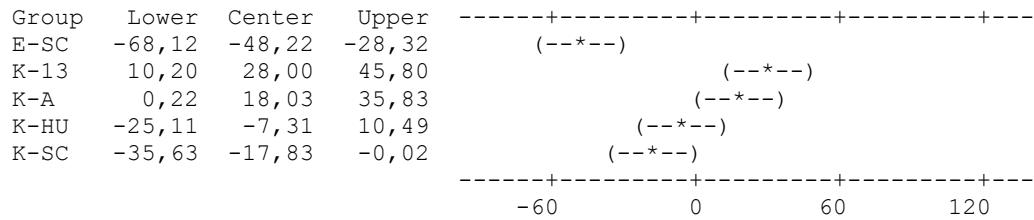




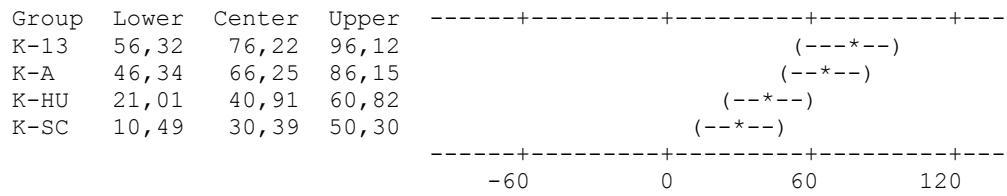
Group = E-Laff subtracted from:



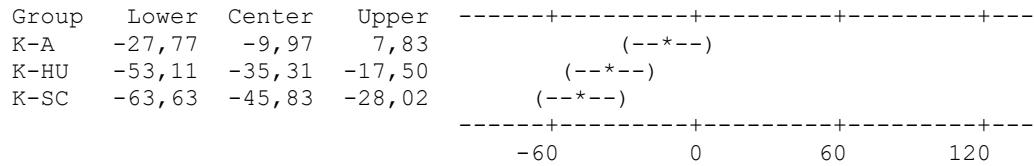
Group = E-LT-SC subtracted from:



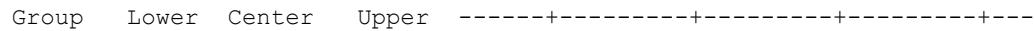
Group = E-SC subtracted from:

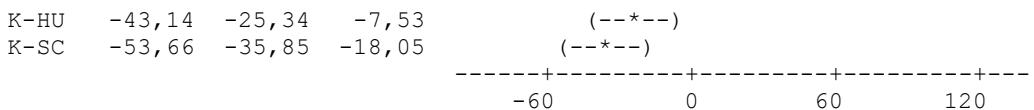


Group = K-13 subtracted from:

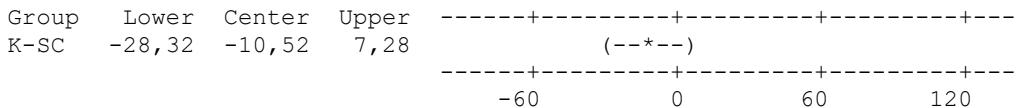


Group = K-A subtracted from:





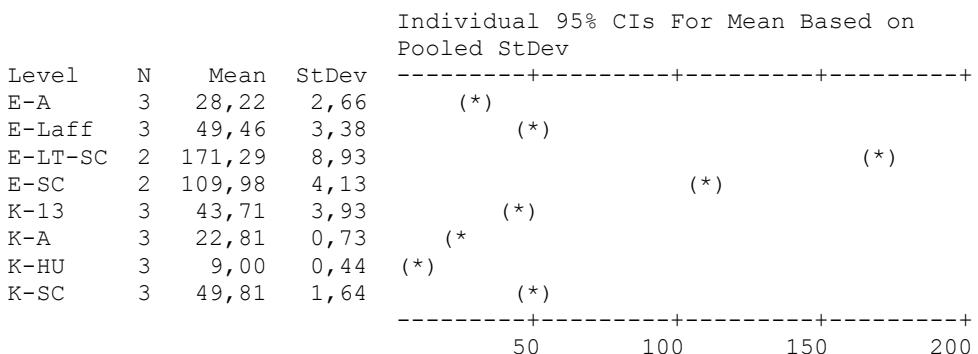
Group = K-HU subtracted from:



One-way ANOVA: Nonanoic acid versus Group

Source	DF	SS	MS	F	P
Group	7	45185,7	6455,1	526,46	0,000
Error	14	171,7	12,3		
Total	21	45357,3			

S = 3,502 R-Sq = 99,62% R-Sq(adj) = 99,43%



Pooled StDev = 3,50

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	2	171,29	A
E-SC	2	109,98	B
K-SC	3	49,81	C
E-Laff	3	49,46	C
K-13	3	43,71	C
E-A	3	28,22	D
K-A	3	22,81	D
K-HU	3	9,00	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	11,16	21,25	31,33		(*)		
E-LT-SC	131,79	143,07	154,35			(*)	
E-SC	70,48	81,76	93,03				(*)
K-13	5,40	15,49	25,58				(*)
K-A	-15,49	-5,41	4,68				(*)
K-HU	-29,31	-19,22	-9,13				(*)
K-SC	11,50	21,59	31,68				(*)
				-100	0	100	200

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
E-LT-SC	110,54	121,82	133,10			(*)	
E-SC	49,23	60,51	71,79				(*)
K-13	-15,84	-5,75	4,33				(*)
K-A	-36,74	-26,65	-16,56				(*)
K-HU	-50,55	-40,47	-30,38				(*)
K-SC	-9,74	0,35	10,43				(*)
				-100	0	100	200

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-73,67	-61,31	-48,96			(*)	
K-13	-138,85	-127,58	-116,30				(*)
K-A	-159,75	-148,47	-137,20				(*)
K-HU	-173,57	-162,29	-151,01				(*)
K-SC	-132,76	-121,48	-110,20				(*)
				-100	0	100	200

Group = E-SC subtracted from:

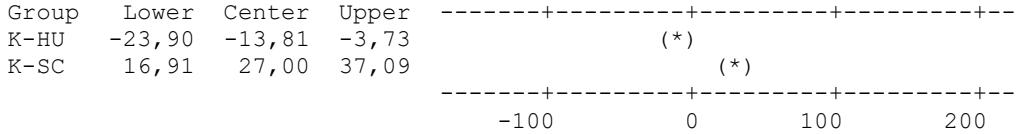
Group	Lower	Center	Upper				
K-13	-77,54	-66,26	-54,99			(*-)	
K-A	-98,44	-87,16	-75,88				(*)
K-HU	-112,26	-100,98	-89,70				(*)
K-SC	-71,44	-60,17	-48,89				(*)
				-100	0	100	200

Group = K-13 subtracted from:

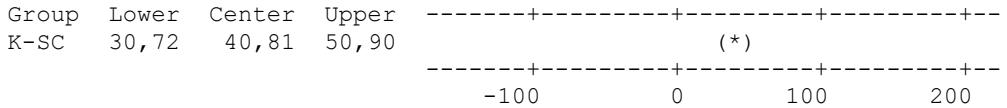
Group	Lower	Center	Upper				
K-A	-30,99	-20,90	-10,81			(*)	
K-HU	-44,80	-34,71	-24,62				(*)
K-SC	-3,99	6,10	16,19				(*)
				-100	0	100	200

-100 0 100 200

Group = K-A subtracted from:



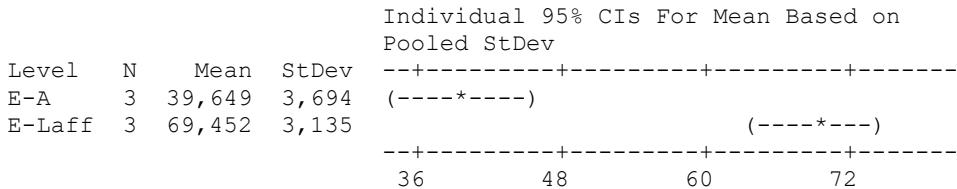
Group = K-HU subtracted from:



One-way ANOVA: Tetradecanoic acid versus Group

Source	DF	SS	MS	F	P
Group	1	1332,4	1332,4	113,52	0,000
Error	4	46,9	11,7		
Total	5	1379,3			

S = 3,426 R-Sq = 96,60% R-Sq(adj) = 95,75%



Pooled StDev = 3,426

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-Laff	3	69,452	A
E-A	3	39,649	B

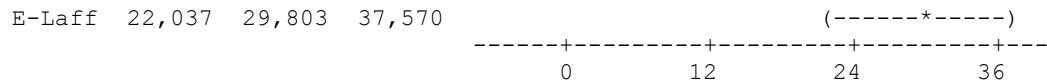
Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

Group = E-A subtracted from:

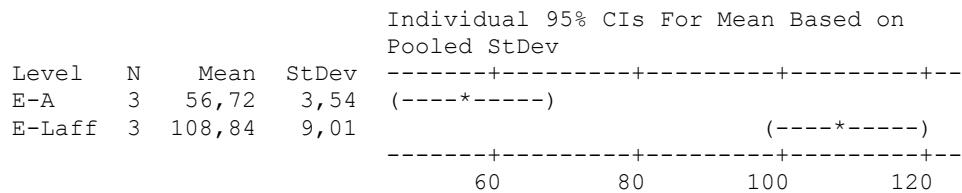




One-way ANOVA: Dodecanoic acid versus Group

Source	DF	SS	MS	F	P
Group	1	4075,6	4075,6	87,00	0,001
Error	4	187,4	46,8		
Total	5	4263,0			

S = 6,844 R-Sq = 95,60% R-Sq(adj) = 94,51%



Pooled StDev = 6,84

Grouping Information Using Tukey Method

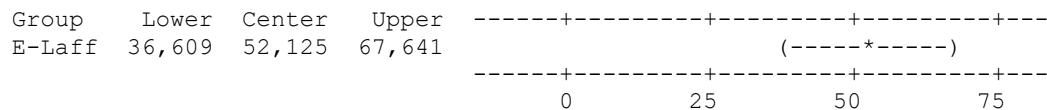
Group	N	Mean	Grouping
E-Laff	3	108,842	A
E-A	3	56,717	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

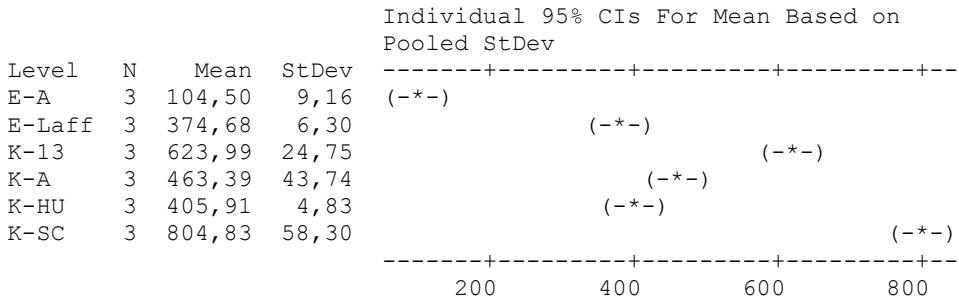
Group = E-A subtracted from:



One-way ANOVA: Pentanoic acid versus Group

Source	DF	SS	MS	F	P
Group	5	847043	169409	167,40	0,000
Error	12	12144	1012		
Total	17	859187			

S = 31,81 R-Sq = 98,59% R-Sq(adj) = 98,00%



Pooled StDev = 31,81

Grouping Information Using Tukey Method

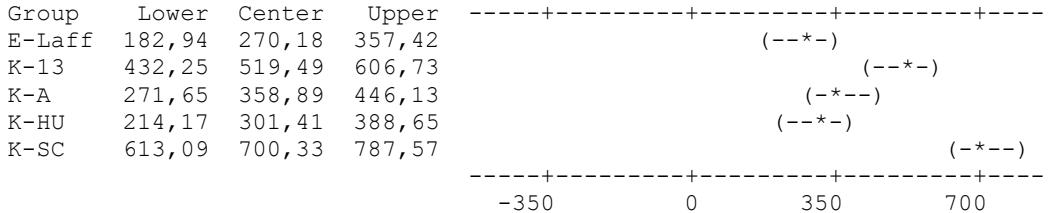
Group	N	Mean	Grouping
K-SC	3	804,83	A
K-13	3	623,99	B
K-A	3	463,39	C
K-HU	3	405,91	C D
E-Laff	3	374,68	D
E-A	3	104,50	E

Means that do not share a letter are significantly different.

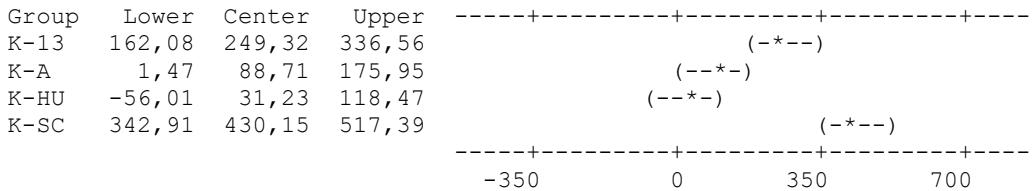
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

Group = E-A subtracted from:

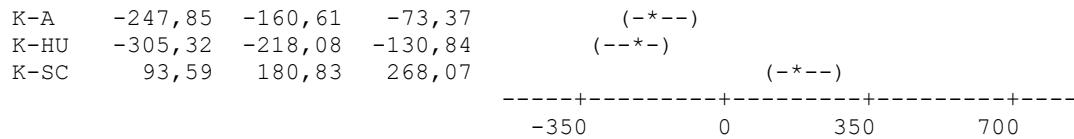


Group = E-Laff subtracted from:

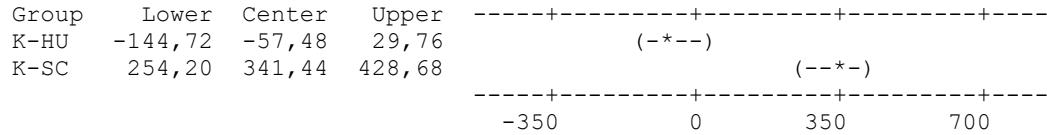


Group = K-13 subtracted from:

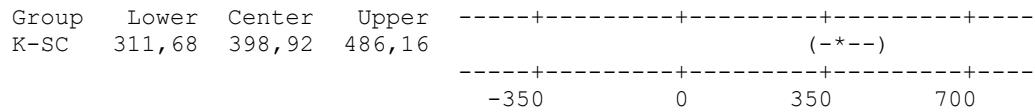
Group	Lower	Center	Upper	
E-A	104,50	270,18	357,42	(--*-)



Group = K-A subtracted from:



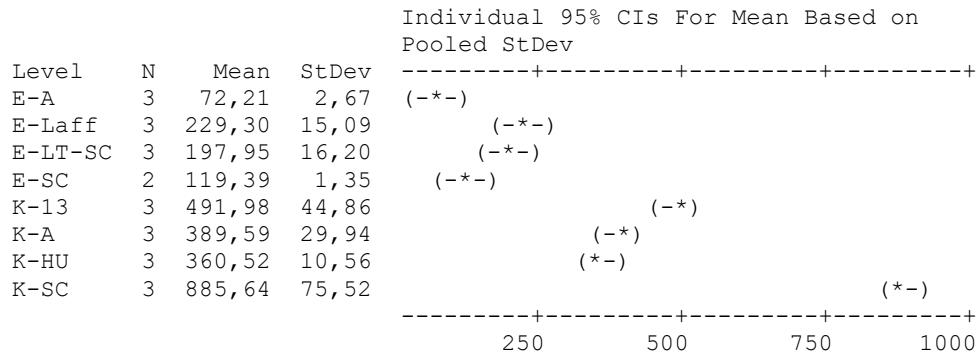
Group = K-HU subtracted from:



One-way ANOVA: Isobutyric acid versus Group

Source	DF	SS	MS	F	P
Group	7	1376941	196706	159,98	0,000
Error	15	18444	1230		
Total	22	1395385			

S = 35,07 R-Sq = 98,68% R-Sq(adj) = 98,06%



Pooled StDev = 35,07

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-SC	3	885,64	A
K-13	3	491,98	B
K-A	3	389,59	C
K-HU	3	360,52	C
E-Laff	3	229,30	D

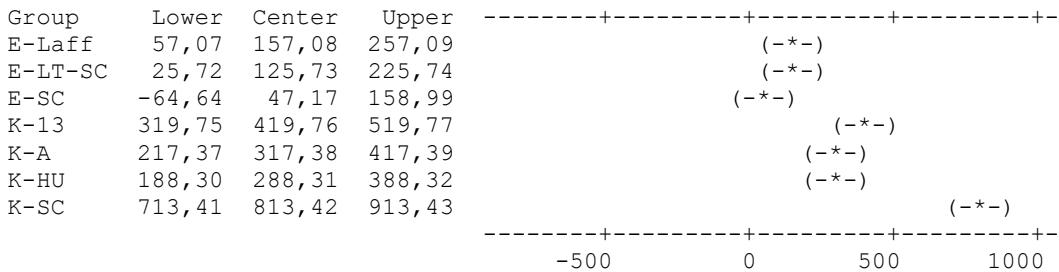
E-LT-SC	3	197, 95	D
E-SC	2	119, 39	D E
E-A	3	72, 21	E

Means that do not share a letter are significantly different.

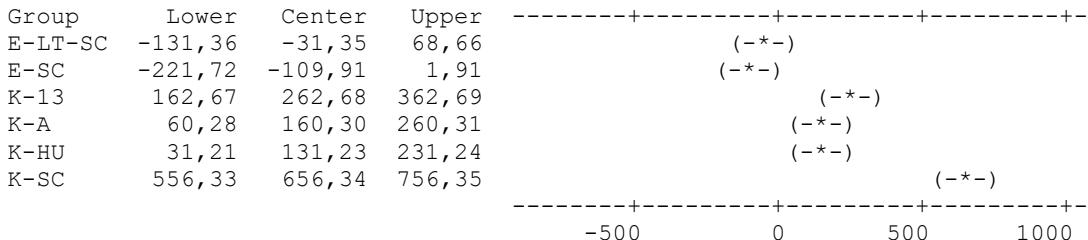
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

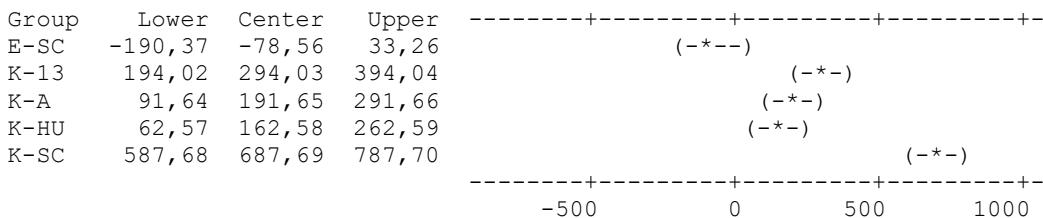
Group = E-A subtracted from:



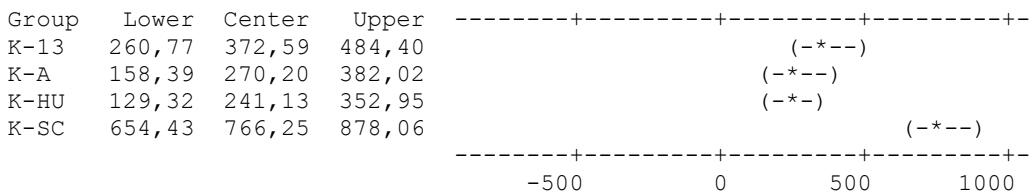
Group = E-Laff subtracted from:



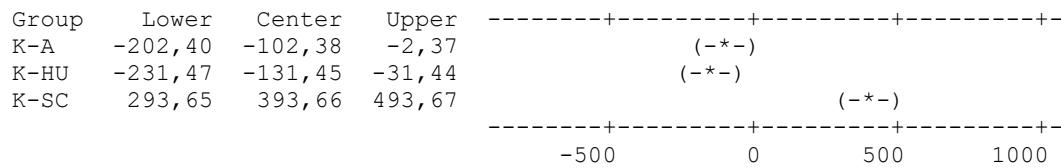
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



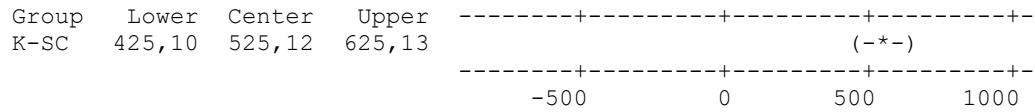
Group = K-13 subtracted from:



Group = K-A subtracted from:



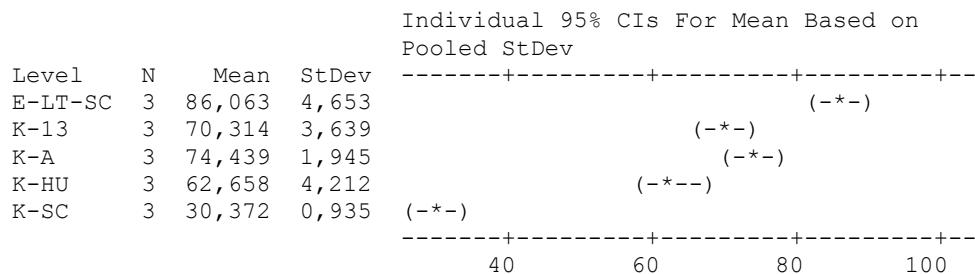
Group = K-HU subtracted from:



One-way ANOVA: Heptanoic acid versus Group

Source	DF	SS	MS	F	P
Group	4	5296,0	1324,0	115,55	0,000
Error	10	114,6	11,5		
Total	14	5410,6			

S = 3,385 R-Sq = 97,88% R-Sq(adj) = 97,04%



Pooled StDev = 3,385

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	86,063	A
K-A	3	74,439	B
K-13	3	70,314	B C

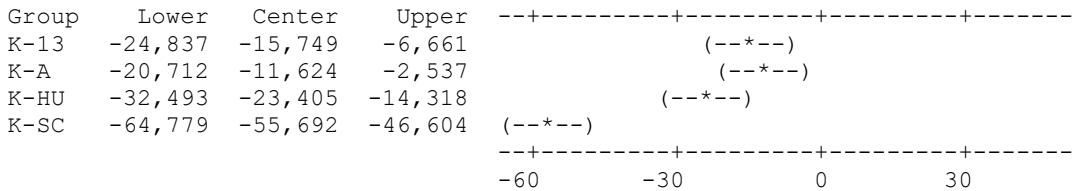
K-HU	3	62,658	C
K-SC	3	30,372	D

Means that do not share a letter are significantly different.

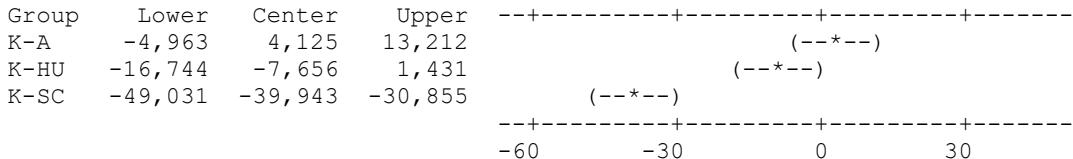
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,18%

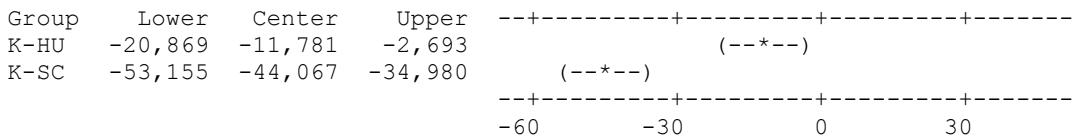
Group = E-LT-SC subtracted from:



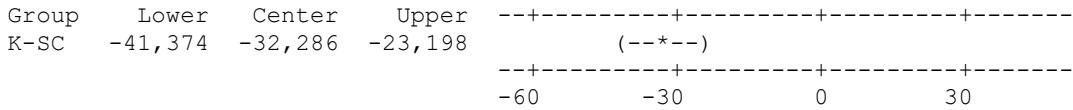
Group = K-13 subtracted from:



Group = K-A subtracted from:



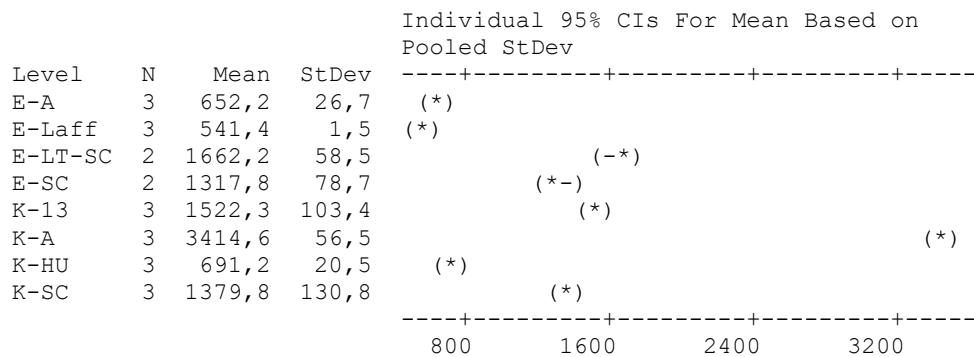
Group = K-HU subtracted from:



One-way ANOVA: Acetic acid versus Group

Source	DF	SS	MS	F	P
Group	7	17767320	2538189	481,16	0,000
Error	14	73852	5275		
Total	21	17841172			

S = 72,63 R-Sq = 99,59% R-Sq(adj) = 99,38%



Pooled StDev = 72,6

Grouping Information Using Tukey Method

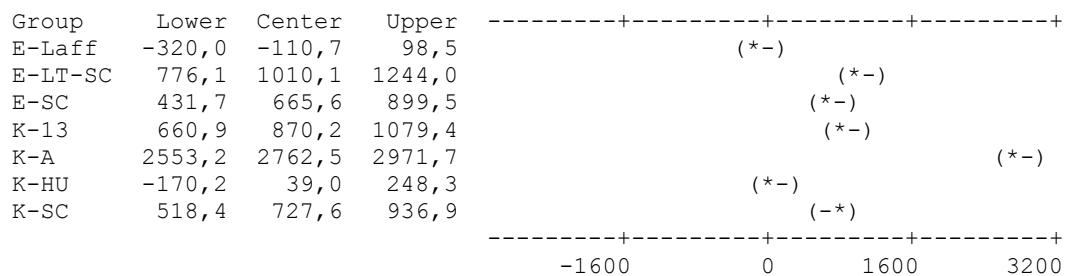
Group	N	Mean	Grouping
K-A	3	3414,6	A
E-LT-SC	2	1662,2	B
K-13	3	1522,3	B C
K-SC	3	1379,8	C
E-SC	2	1317,8	C
K-HU	3	691,2	D
E-A	3	652,2	D
E-Laff	3	541,4	D

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:



Group = E-Laff subtracted from:



K-HU	-59,5	149,8	359,0	(*)
K-SC	629,1	838,4	1047,6	(*-)
				-----+-----+-----+-----+

-1600 0 1600 3200

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
E-SC	-600,8	-344,5	-88,2	(-*)
K-13	-373,8	-139,9	94,1	(*-)
K-A	1518,4	1752,4	1986,3	(-*)
K-HU	-1205,0	-971,0	-737,1	(-*)
K-SC	-516,4	-282,4	-48,5	(*-)
				-----+-----+-----+-----+
				-1600 0 1600 3200

Group = E-SC subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
K-13	-29,4	204,6	438,5	(*-)
K-A	1862,9	2096,9	2330,8	(*-)
K-HU	-860,5	-626,6	-392,6	(*-)
K-SC	-171,9	62,0	296,0	(*-)
				-----+-----+-----+-----+
				-1600 0 1600 3200

Group = K-13 subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
K-A	1683,0	1892,3	2101,5	(*)
K-HU	-1040,4	-831,2	-621,9	(-*)
K-SC	-351,8	-142,6	66,7	(*)
				-----+-----+-----+-----+
				-1600 0 1600 3200

Group = K-A subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
K-HU	-2932,7	-2723,4	-2514,2	(*)
K-SC	-2244,1	-2034,8	-1825,6	(-*)
				-----+-----+-----+-----+
				-1600 0 1600 3200

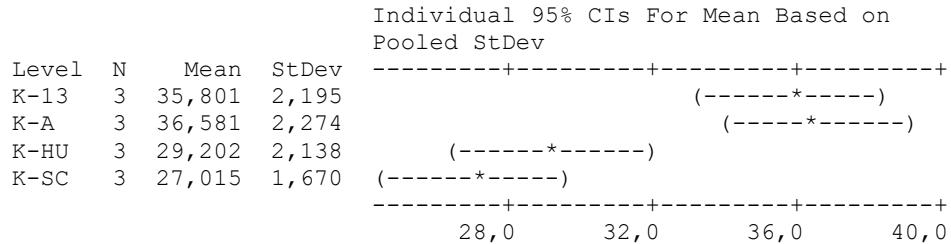
Group = K-HU subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
K-SC	479,4	688,6	897,8	(*)
				-----+-----+-----+-----+
				-1600 0 1600 3200

One-way ANOVA: Diethyl dl-malate versus Group

Source	DF	SS	MS	F	P
Group	3	204,07	68,02	15,69	0,001
Error	8	34,69	4,34		
Total	11	238,76			

S = 2,082 R-Sq = 85,47% R-Sq(adj) = 80,02%



Pooled StDev = 2,082

Grouping Information Using Tukey Method

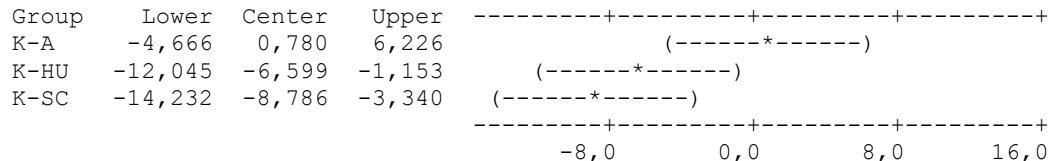
Group	N	Mean	Grouping
K-A	3	36,581	A
K-13	3	35,801	A
K-HU	3	29,202	B
K-SC	3	27,015	B

Means that do not share a letter are significantly different.

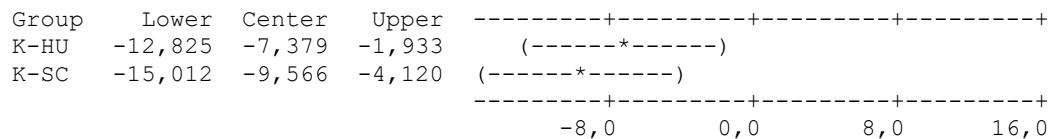
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

Group = K-13 subtracted from:



Group = K-A subtracted from:



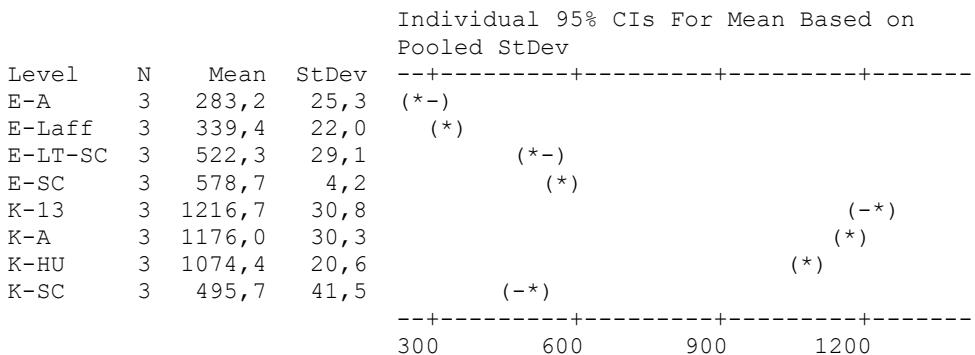
Group = K-HU subtracted from:

Group	Lower	Center	Upper	-----+-----+-----+-----+
K-SC	-7,633	-2,187	3,259	(-----*-----)
				-----+-----+-----+-----+
				-8,0 0,0 8,0 16,0

One-way ANOVA: γ -butyrolactone versus Group

Source	DF	SS	MS	F	P
Group	7	3073759	439108	584,57	0,000
Error	16	12019	751		
Total	23	3085778			

S = 27,41 R-Sq = 99,61% R-Sq(adj) = 99,44%



Pooled StDev = 27,4

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-13	3	1216,7	A
K-A	3	1176,0	A
K-HU	3	1074,4	B
E-SC	3	578,7	C
E-LT-SC	3	522,3	C D
K-SC	3	495,7	D
E-Laff	3	339,4	E
E-A	3	283,2	E

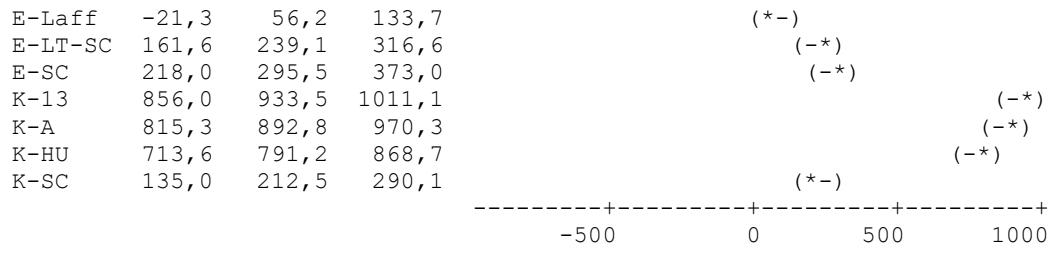
Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

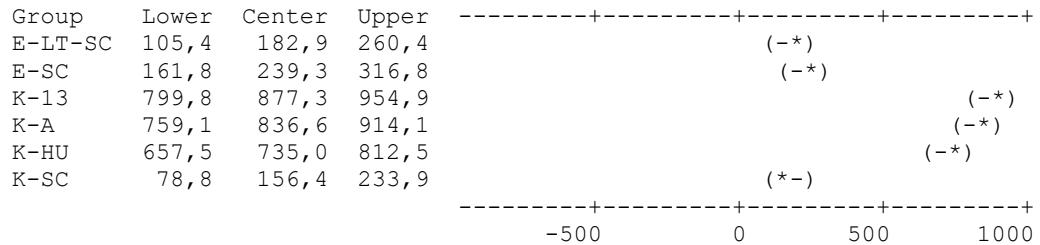
Individual confidence level = 99,68%

Group = E-A subtracted from:

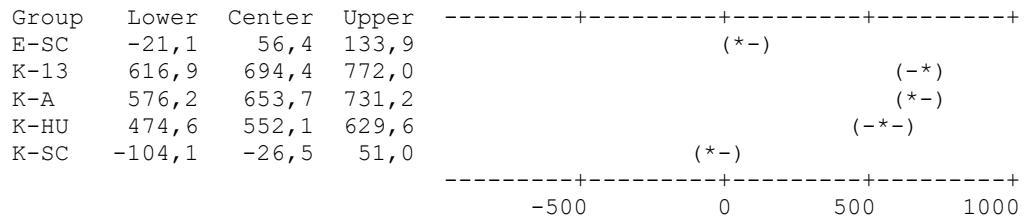
Group	Lower	Center	Upper	-----+-----+-----+-----+



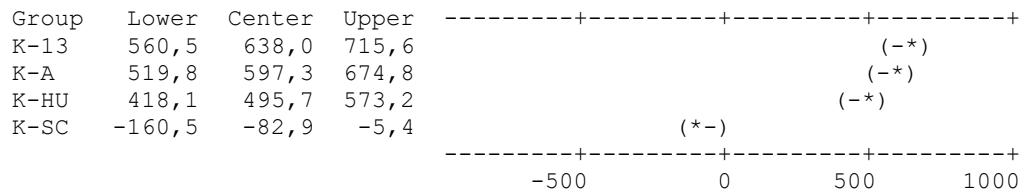
Group = E-Laff subtracted from:



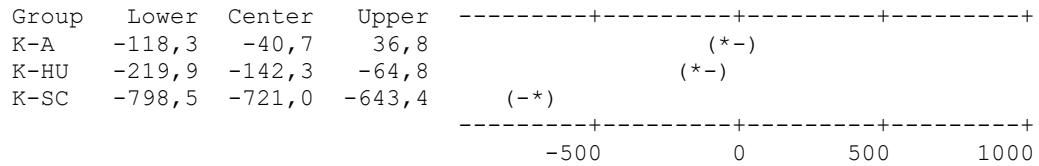
Group = E-LT-SC subtracted from:



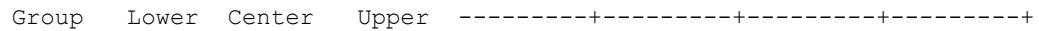
Group = E-SC subtracted from:

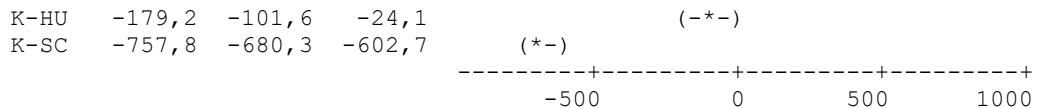


Group = K-13 subtracted from:

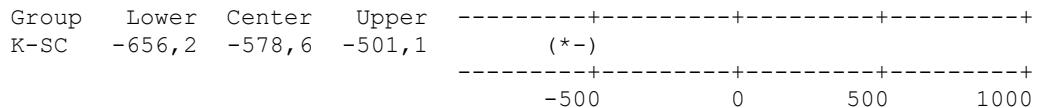


Group = K-A subtracted from:





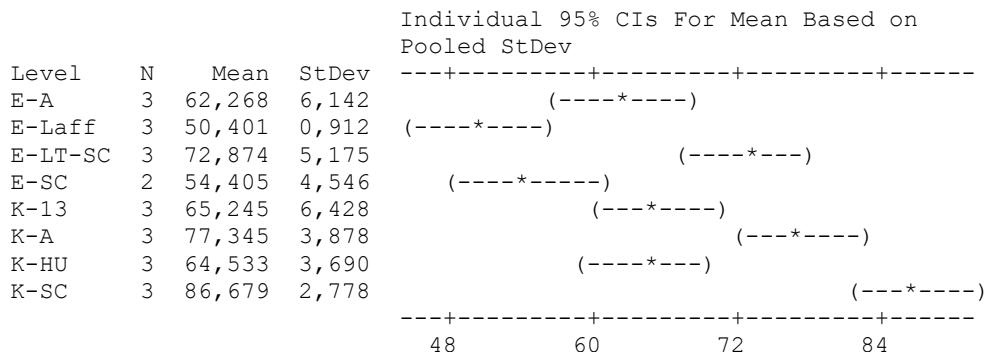
Group = K-HU subtracted from:



One-way ANOVA: Pantolactone versus Group

Source	DF	SS	MS	F	P
Group	7	2823,5	403,4	19,72	0,000
Error	15	306,7	20,4		
Total	22	3130,2			

S = 4,522 R-Sq = 90,20% R-Sq(adj) = 85,63%



Pooled StDev = 4,522

Grouping Information Using Tukey Method

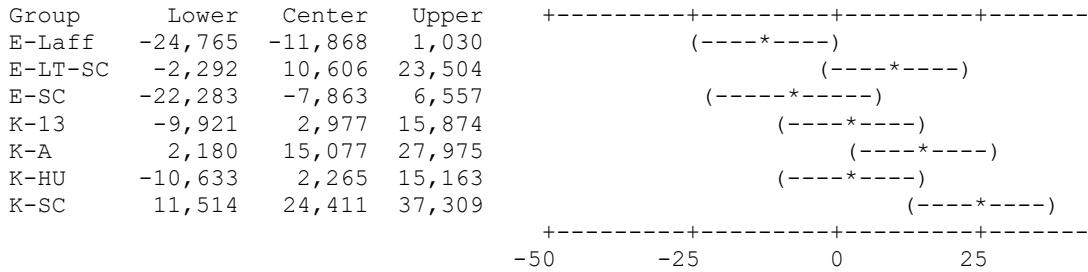
Group	N	Mean	Grouping
K-SC	3	86,679	A
K-A	3	77,345	A B
E-LT-SC	3	72,874	B C
K-13	3	65,245	B C D
K-HU	3	64,533	B C D
E-A	3	62,268	C D E
E-SC	2	54,405	D E
E-Laff	3	50,401	E

Means that do not share a letter are significantly different.

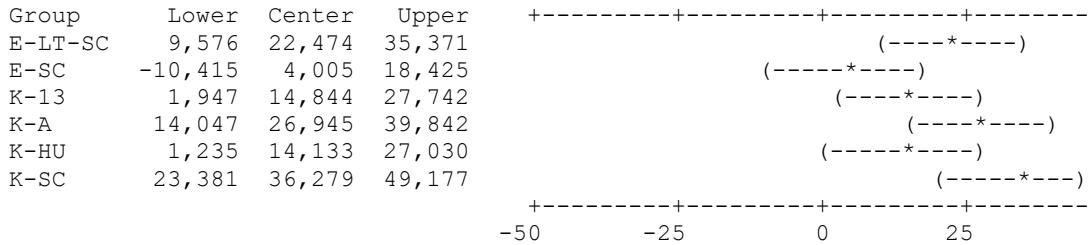
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

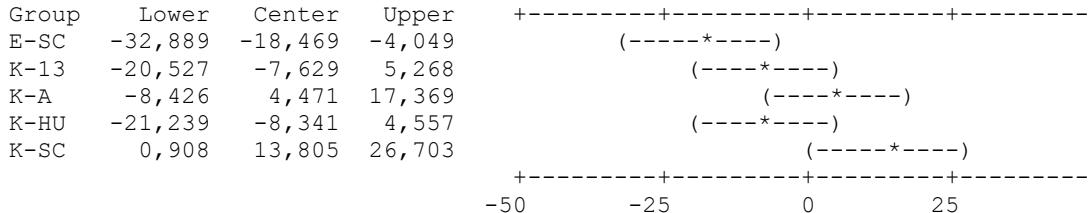
Group = E-A subtracted from:



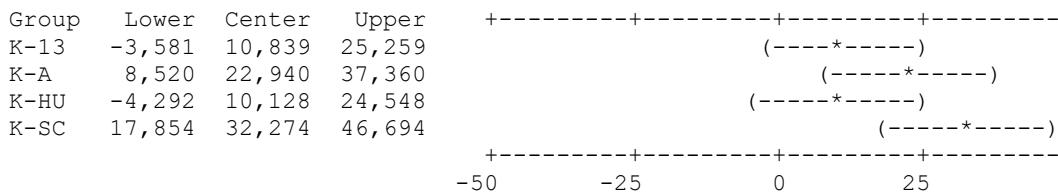
Group = E-Laff subtracted from:



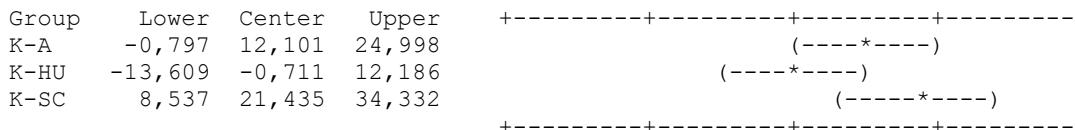
Group = E-LT-SC subtracted from:



Group = E-SC subtracted from:



Group = K-13 subtracted from:



-50 -25 0 25

Group = K-A subtracted from:

Group	Lower	Center	Upper	
K-HU	-25,710	-12,812	0,086	(----*----)
K-SC	-3,564	9,334	22,232	(----*----)
	-50	-25	0	25

Group = K-HU subtracted from:

Group	Lower	Center	Upper	
K-SC	9,249	22,146	35,044	(----*----)
	-50	-25	0	25

One-way ANOVA: 2-Methoxy-4-vinylphenol versus Group

Source	DF	SS	MS	F	P
Group	7	188306,9	26901,0	376,28	0,000
Error	16	1143,9	71,5		
Total	23	189450,8			

S = 8,455 R-Sq = 99,40% R-Sq(adj) = 99,13%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev	
E-A	3	142,38	9,09	(*-)
E-Laff	3	176,57	5,32	(*-)
E-LT-SC	3	283,40	18,50	(*-)
E-SC	3	290,51	5,90	(-*)
K-13	3	79,03	1,40	(*-)
K-A	3	153,11	7,84	(-*)
K-HU	3	52,17	4,50	(*-)
K-SC	3	286,06	0,68	(-*)

70 140 210 280

Pooled StDev = 8,46

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-SC	3	290,51	A
K-SC	3	286,06	A
E-LT-SC	3	283,40	A
E-Laff	3	176,57	B
K-A	3	153,11	B C
E-A	3	142,38	C
K-13	3	79,03	D
K-HU	3	52,17	E

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,68%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
E-Laff	10,27	34,19	58,11			(*-)	
E-LT-SC	117,10	141,02	164,94			(*-)	
E-SC	124,21	148,14	172,06			(*-)	
K-13	-87,26	-63,34	-39,42		(-*)		
K-A	-13,19	10,73	34,65		(-*)		
K-HU	-114,12	-90,20	-66,28		(-*-)		
K-SC	119,77	143,69	167,61			(-*)	
				-150	0	150	300

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
-							
E-LT-SC	82,91	106,83	130,75			(*-)	
E-SC	90,02	113,94	137,86			(-*)	
K-13	-121,45	-97,53	-73,61		(-*)		
K-A	-47,38	-23,46	0,46		(-*)		
K-HU	-148,31	-124,39	-100,47		(-*)		
K-SC	85,58	109,50	133,42			(-*)	
-				-150	0	150	300

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper				
E-SC	-16,81	7,11	31,03			(*-)	
K-13	-228,28	-204,36	-180,44		(-*)		
K-A	-154,21	-130,29	-106,37		(-*)		
K-HU	-255,14	-231,22	-207,30		(-*)		
K-SC	-21,25	2,67	26,59			(-*)	
				-150	0	150	300

Group = E-SC subtracted from:

Group	Lower	Center	Upper				
K-13	-235,40	-211,48	-187,56		(-*)		
K-A	-161,32	-137,40	-113,48		(-*)		
K-HU	-262,26	-238,34	-214,42		(-*)		
K-SC	-28,37	-4,45	19,47			(-*)	
				-150	0	150	300

Group = K-13 subtracted from:

Group	Lower	Center	Upper		
K-A	50,15	74,07	97,99		(--*)
K-HU	-50,78	-26,86	-2,94		(*-)
K-SC	183,11	207,03	230,95		(-*)

-----+-----+-----+-----+
-150 0 150 300

Group = K-A subtracted from:

Group	Lower	Center	Upper		
K-HU	-124,86	-100,93	-77,01		(*-)
K-SC	109,03	132,95	156,87		(-*)

-----+-----+-----+-----+
-150 0 150 300

Group = K-HU subtracted from:

Group	Lower	Center	Upper		
K-SC	209,97	233,89	257,81		(-*)

-----+-----+-----+-----+
-150 0 150 300

One-way ANOVA: 4-vinyl-phenol versus Group

Source	DF	SS	MS	F	P
Group	3	82716	27572	107,17	0,000
Error	8	2058	257		
Total	11	84774			

S = 16,04 R-Sq = 97,57% R-Sq(adj) = 96,66%

Level	N	Mean	StDev	Individual 95% CIs For Mean Based on Pooled StDev	
				(--*)	(--*)
E-LT-SC	3	329,95	28,57		(--*)
E-SC	3	254,27	12,56		(--*)
K-A	3	100,22	4,13	(--*)	
K-SC	3	213,46	6,17		(--*)

-----+-----+-----+-----+
140 210 280 350

Pooled StDev = 16,04

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
E-LT-SC	3	329,95	A
E-SC	3	254,27	B
K-SC	3	213,46	B
K-A	3	100,22	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper					
E-SC	-117,63	-75,68	-33,73				(---*---)	
K-A	-271,68	-229,72	-187,77	(---*---)				
K-SC	-158,44	-116,49	-74,54				(---*---)	
	-240		-120			0		120

Group = E-SC subtracted from:

Group	Lower	Center	Upper					
K-A	-195,99	-154,04	-112,09				(---*---)	
K-SC	-82,76	-40,81	1,14				(---*---)	
	-240		-120			0		120

Group = K-A subtracted from:

Group	Lower	Center	Upper					
K-SC	71,28	113,24	155,19				(---*---)	
	-240		-120			0		120

One-way ANOVA: Phenol versus Group

Source	DF	SS	MS	F	P
Group	1	216,47	216,47	114,16	0,000
Error	4	7,59	1,90		
Total	5	224,06			

S = 1,377 R-Sq = 96,61% R-Sq(adj) = 95,77%

Level	N	Mean	StDev	Individual 95% CIs For Mean Based on Pooled StDev			
				10,0	15,0	20,0	25,0
E-A	3	24,180	1,770				(---*---)
E-Laff	3	12,167	0,811	(---*---)			

Pooled StDev = 1,377

Grouping Information Using Tukey Method

Group N Mean Grouping

E-A	3	24,180	A
E-Laff	3	12,167	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 95,00%

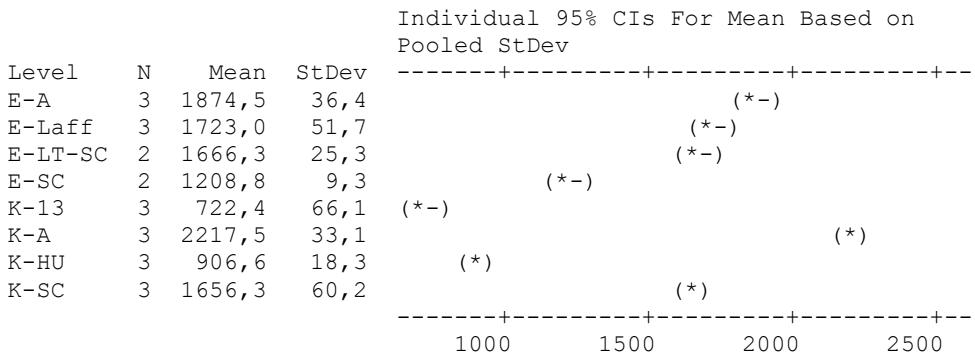
Group = E-A subtracted from:

Group	Lower	Center	Upper	
E-Laff	-15,135	-12,013	-8,891	(-----*-----)
				+-----+-----+-----+
	-15,0	-10,0	-5,0	0,0

One-way ANOVA: Tyrosol versus Group

Source	DF	SS	MS	F	P
Group	7	5282953	754708	383,26	0,000
Error	14	27569	1969		
Total	21	5310522			

S = 44,38 R-Sq = 99,48% R-Sq(adj) = 99,22%



Pooled StDev = 44,4

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	2217,5	A
E-A	3	1874,5	B
E-Laff	3	1723,0	C
E-LT-SC	2	1666,3	C
K-SC	3	1656,3	C
E-SC	2	1208,8	D
K-HU	3	906,6	E
K-13	3	722,4	F

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

Group	Lower	Center	Upper					
-								
E-Laff	-279,4	-151,5	-23,7		(*)			
E-LT-SC	-351,2	-208,2	-65,3		(-*)			
E-SC	-808,7	-665,7	-522,8		(*)			
K-13	-1280,0	-1152,1	-1024,3	(*)				
K-A	215,1	343,0	470,8		(*)			
K-HU	-1095,8	-968,0	-840,1	(*)				
K-SC	-346,1	-218,2	-90,4	(*)				
-								
				-1000	0	1000	2000	

Group = E-Laff subtracted from:

Group	Lower	Center	Upper					
E-LT-SC	-199,7	-56,7	86,2		(*)			
E-SC	-657,1	-514,2	-371,3		(-*)			
K-13	-1128,4	-1000,6	-872,8	(*)				
K-A	366,6	494,5	622,3		(*)			
K-HU	-944,3	-816,5	-688,6	(*)				
K-SC	-194,6	-66,7	61,1	(*)				
-				-1000	0	1000	2000	

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper					
E-SC	-614,1	-457,5	-300,9		(*)			
K-13	-1086,8	-943,9	-800,9	(-*)				
K-A	408,3	551,2	694,1		(-*)			
K-HU	-902,7	-759,7	-616,8	(*)				
K-SC	-152,9	-10,0	132,9	(-*)				
-				-1000	0	1000	2000	

Group = E-SC subtracted from:

Group	Lower	Center	Upper					
K-13	-629,3	-486,4	-343,5		(*)			
K-A	865,8	1008,7	1151,6		(-*)			
K-HU	-445,2	-302,2	-159,3	(*)				
K-SC	304,6	447,5	590,4	(*)				
-				-1000	0	1000	2000	

Group = K-13 subtracted from:

Group	Lower	Center	Upper		
K-A	1367,2	1495,1	1622,9		(*)
K-HU	56,3	184,2	312,0	(*)	
K-SC	806,0	933,9	1061,7	(*-)	

-----+-----+-----+-----+-----+-----
-1000 0 1000 2000

Group = K-A subtracted from:

Group	Lower	Center	Upper		
K-HU	-1438,8	-1310,9	-1183,1	(*)	
K-SC	-689,0	-561,2	-433,4	(*-)	

-----+-----+-----+-----+-----+-----
-1000 0 1000 2000

Group = K-HU subtracted from:

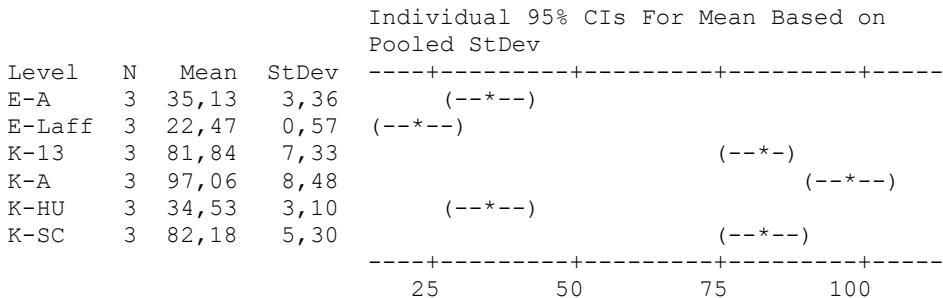
Group	Lower	Center	Upper		
K-SC	621,9	749,7	877,6	(*-)	

-----+-----+-----+-----+-----+-----
-1000 0 1000 2000

One-way ANOVA: Soleron versus Group

Source	DF	SS	MS	F	P
Group	5	15030,2	3006,0	103,02	0,000
Error	12	350,2	29,2		
Total	17	15380,4			

S = 5,402 R-Sq = 97,72% R-Sq(adj) = 96,77%



Pooled StDev = 5,40

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	97,061	A
K-SC	3	82,178	B
K-13	3	81,839	B

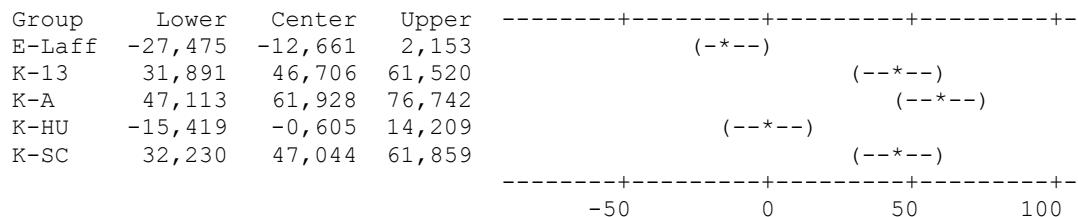
E-A	3	35,134	C
K-HU	3	34,529	C
E-Laff	3	22,473	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

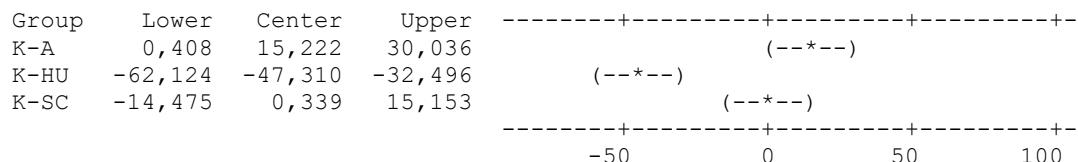
Group = E-A subtracted from:



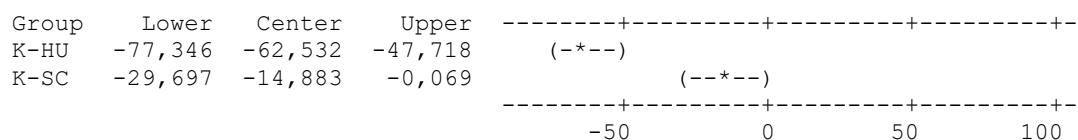
Group = E-Laff subtracted from:



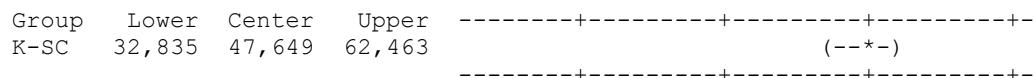
Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:

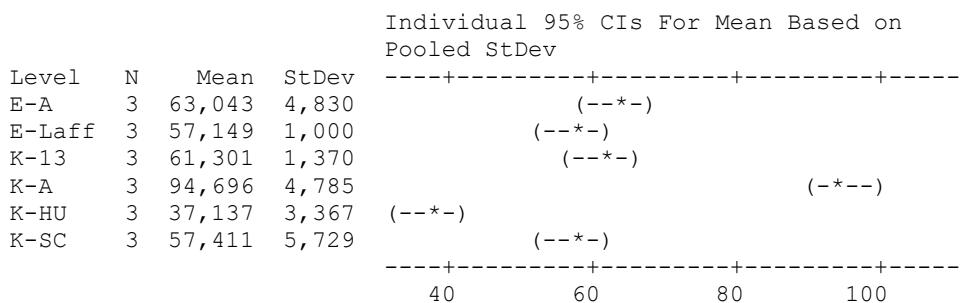


-50 0 50 100

One-way ANOVA: Guaiacol versus Group

Source	DF	SS	MS	F	P
Group	5	5199,3	1039,9	66,91	0,000
Error	12	186,5	15,5		
Total	17	5385,8			

S = 3,942 R-Sq = 96,54% R-Sq(adj) = 95,09%



Pooled StDev = 3,942

Grouping Information Using Tukey Method

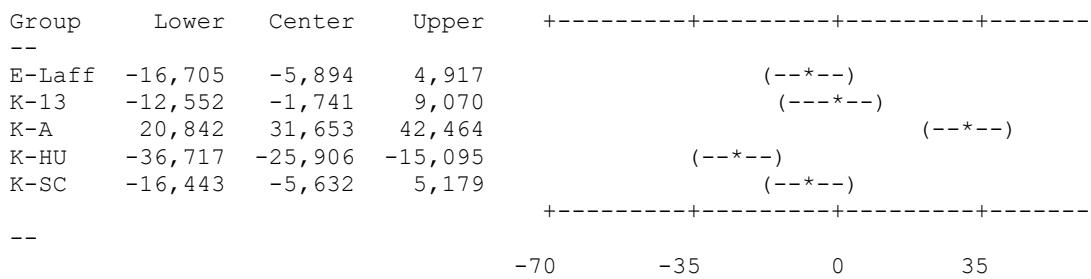
Group	N	Mean	Grouping
K-A	3	94,696	A
E-A	3	63,043	B
K-13	3	61,301	B
K-SC	3	57,411	B
E-Laff	3	57,149	B
K-HU	3	37,137	C

Means that do not share a letter are significantly different.

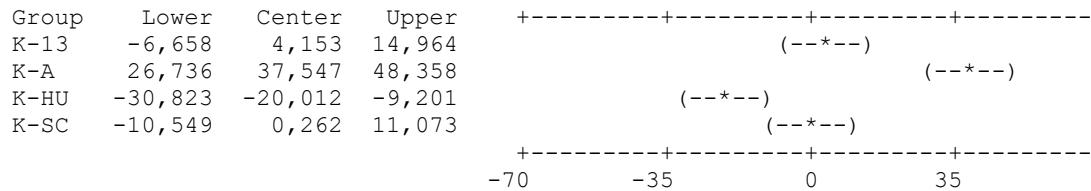
Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,43%

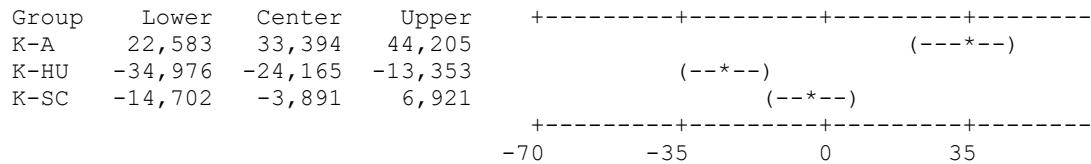
Group = E-A subtracted from:



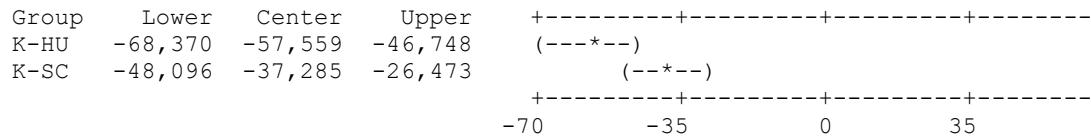
Group = E-Laff subtracted from:



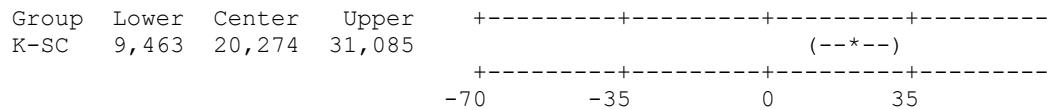
Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:

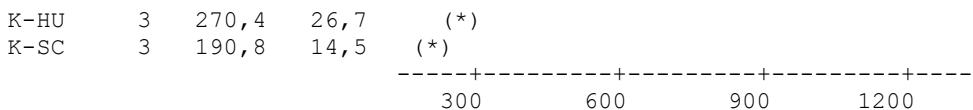


One-way ANOVA: Acetoin versus Group

Source	DF	SS	MS	F	P
Group	7	3145275	449325	1381,23	0,000
Error	15	4880	325		
Total	22	3150154			

S = 18,04 R-Sq = 99,85% R-Sq(adj) = 99,77%

Individual 95% CIs For Mean Based on Pooled StDev					
Level	N	Mean	StDev		
E-A	3	190,3	10,7	(*)	
E-Laff	3	175,9	16,0	(*)	
E-LT-SC	2	168,1	0,2	(*)	
E-SC	3	178,7	7,8	(*)	
K-13	3	200,7	19,1	(*)	
K-A	3	1292,3	26,8		(*)



Pooled StDev = 18,0

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-A	3	1292,3	A
K-HU	3	270,4	B
K-13	3	200,7	C
K-SC	3	190,8	C
E-A	3	190,3	C
E-SC	3	178,7	C
E-Laff	3	175,9	C
E-LT-SC	2	168,1	C

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 99,67%

Group = E-A subtracted from:

Group	Lower	Center	Upper				
--				-----+-----+-----+-----+-----			
E-Laff	-65,8	-14,4	37,0				(*)
E-LT-SC	-79,7	-22,2	35,3				(*)
E-SC	-63,0	-11,5	39,9				(*)
K-13	-41,0	10,4	61,8				(*)
K-A	1050,6	1102,0	1153,4				
*)							
K-HU	28,6	80,1	131,5				(*)
K-SC	-50,9	0,5	52,0				(*)
--				-----+-----+-----+-----+-----			
				-1200	-600	0	600

Group = E-Laff subtracted from:

Group	Lower	Center	Upper				
--				-----+-----+-----+-----+-----			
E-LT-SC	-65,3	-7,8	49,7				(*)
E-SC	-48,6	2,9	54,3				(*)
K-13	-26,6	24,8	76,2				(*)
K-A	1065,0	1116,4	1167,9				(*)
K-HU	43,0	94,5	145,9				(*)
K-SC	-36,5	14,9	66,4				(*)
--				-----+-----+-----+-----+-----			
				-1200	-600	0	600

Group = E-LT-SC subtracted from:

Group	Lower	Center	Upper			
E-SC	-46,9	10,7	68,2		(*)	
K-13	-24,9	32,6	90,1		(*)	
K-A	1066,7	1124,2	1181,7			
(*)						
K-HU	44,8	102,3	159,8		(*)	
K-SC	-34,8	22,7	80,2		(*)	
				+-----+-----+-----+		
				-1200	-600	0
						600

Group = E-SC subtracted from:

Group	Lower	Center	Upper			
K-13	-29,5	21,9	73,4		(*)	
K-A	1062,1	1113,5	1165,0			(*)
K-HU	40,2	91,6	143,1		(*)	
K-SC	-39,4	12,1	63,5		(*)	
				+-----+-----+-----+		
				-1200	-600	0
						600

Group = K-13 subtracted from:

Group	Lower	Center	Upper			
K-A	1040,2	1091,6	1143,1			(*)
K-HU	18,2	69,7	121,1		(*)	
K-SC	-61,3	-9,9	41,6		(*)	
				+-----+-----+-----+		
				-1200	-600	0
						600

Group = K-A subtracted from:

Group	Lower	Center	Upper			
K-HU	-1073,4	-1021,9	-970,5		(*)	
K-SC	-1152,9	-1101,5	-1050,0		(*)	
--				+-----+-----+-----+		
				-1200	-600	0
						600

Group = K-HU subtracted from:

Group	Lower	Center	Upper			
K-SC	-131,0	-79,6	-28,1		(*)	
				+-----+-----+-----+		
				-1200	-600	0
						600

One-way ANOVA: Geraniol versus Group

Source	DF	SS	MS	F	P
Group	3	304,14	101,38	19,62	0,000
Error	8	41,34	5,17		
Total	11	345,49			

S = 2,273 R-Sq = 88,03% R-Sq(adj) = 83,55%

Individual 95% CIs For Mean Based on
Pooled StDev

Level	N	Mean	StDev	
K-13	3	24,013	1,644	(-----*-----)
K-A	3	25,899	1,601	(-----*-----)
K-HU	3	25,284	1,424	(-----*-----)
K-SC	3	36,586	3,657	(-----*-----)

25,0 30,0 35,0 40,0

Pooled StDev = 2,273

Grouping Information Using Tukey Method

Group	N	Mean	Grouping
K-SC	3	36,586	A
K-A	3	25,899	B
K-HU	3	25,284	B
K-13	3	24,013	B

Means that do not share a letter are significantly different.

Tukey 95% Simultaneous Confidence Intervals
All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

Group = K-13 subtracted from:

Group	Lower	Center	Upper	
K-A	-4,060	1,886	7,832	(-----*-----)
K-HU	-4,675	1,271	7,217	(-----*-----)
K-SC	6,627	12,572	18,518	(-----*-----)

-10 0 10 20

Group = K-A subtracted from:

Group	Lower	Center	Upper	
K-HU	-6,561	-0,615	5,331	(-----*-----)
K-SC	4,740	10,686	16,632	(-----*-----)

-10 0 10 20

Group = K-HU subtracted from:

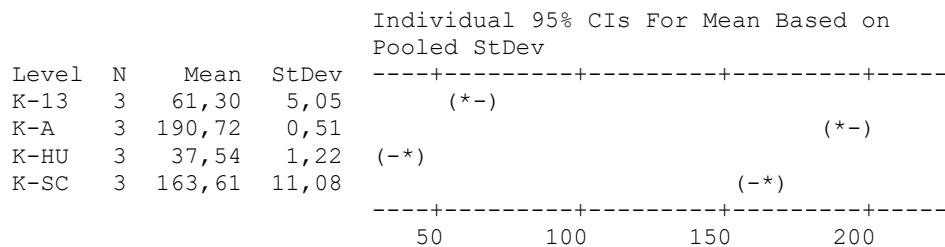
Group	Lower	Center	Upper	
K-SC	5,355	11,301	17,247	(-----*-----)

-10 0 10 20

One-way ANOVA: Syringol versus Group

Source	DF	SS	MS	F	P
Group	3	50904,7	16968,2	452,35	0,000
Error	8	300,1	37,5		
Total	11	51204,8			

S = 6,125 R-Sq = 99,41% R-Sq(adj) = 99,19%



Pooled StDev = 6,12

Grouping Information Using Tukey Method

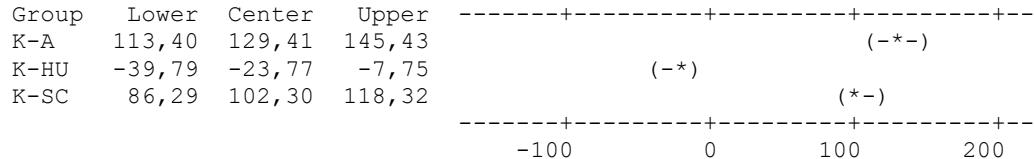
Group	N	Mean	Grouping
K-A	3	190,72	A
K-SC	3	163,61	B
K-13	3	61,30	C
K-HU	3	37,54	D

Means that do not share a letter are significantly different.

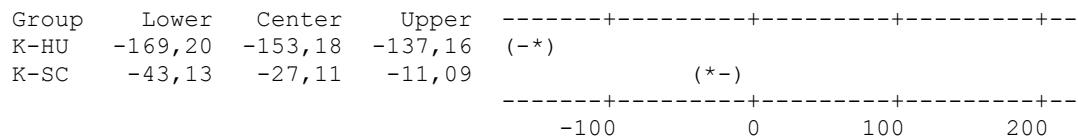
Tukey 95% Simultaneous Confidence Intervals All Pairwise Comparisons among Levels of Group

Individual confidence level = 98,74%

Group = K-13 subtracted from:



Group = K-A subtracted from:



Group = K-HU subtracted from:

Group	Lower	Center	Upper					
K-SC	110,05	126,07	142,09				(-*)	
				-100	0	100		200